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Preliminary Materials for the Integrated Risk Information System (IRIS)
Toxicological Review of Diisobutyl Phthalate (DIBP)
(CASRN No. 84-69-5)

September 2014

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National Center for Environmental Assessment
Office of Research and Development
U.S. Environmental Protection Agency
Washington, DC

Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

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ABBREVIATIONS

AGD	anogenital distance	IQR	interquartile range
aOR	adjusted odds ratio	IRIS	Integrated Risk Information System
BASC-PRS	Behavior Assessment System for Children—Parent Rating Scales	Koc	partition coefficient
BBP	butyl benzyl phthalate	LDL	low-density lipoprotein
BMI	body mass index	LH	luteinizing hormone
BP	blood pressure	LMW	low molecular weight
BPA	bisphenol A	LOD	level of detection
BRIEF	Behavior Rating Inventory of Executive Function	LOQ	level of quantification
BW	body weight	MBzP	mono-benzyl phthalate
CASRN	Chemical Abstracts Service Registry Number	MBP	monobutyl phthalate
CHAP	Chronic Hazard Advisory Panel	MCPP	mono-(3-carboxypropyl) phthalate
CI	confidence interval	MDI	mental delay index
CPSC	Consumer Product Safety Commission	MEHP	mono-(2-ethylhexyl) phthalate
DBP	dibutyl phthalate	MEP	monoethyl phthalate
DEP	di-ethyl phthalate	MHBP	mono-3-(3-carboxypropyl)phthalate
DEHP	di(2-ethylhexyl)phthalate	MIBP	monoisobutyl phthalate
DHEAS	dehydroepiandrosterone	MMP	monomethyl phthalate
DIBP	diisobutyl phthalate	MOA	mode of action
DINP	diisononyl phthalate	MOINP	oxo-(mono-oxoisonyl) phthalate
DnBP	dibutyl phthalate	MRI	magnetic resonance imaging
DNA	deoxyribonucleic acid	NCEA	National Center for Environmental Assessment
DPP	dipentyl phthalate	NHANES	National Health and Nutrition Examination Survey
DXA	dual energy x-ray absorptiometry	NHS	Nurses' Health Study
EPA	Environmental Protection Agency	NRC	National Research Council
FBG	fasting blood glucose	OR	odds ratio
FDA	Food and Drug Administration	ORD	Office of Research and Development
FSH	follicle stimulating hormone	PAH	polycyclic aromatic hydrocarbon
GD	gestational day	PCO	polycystic ovarian morphology
HbA1c	glycosolated hemoglobin	PCOS	polycystic ovarian syndrome
HCG	human chorionic gonadotropin	PDI	psychomotor delay index
HDL	high-density lipoprotein	PND	postnatal day
HERO	Health and Environmental Research Online	PPS	preputial separation
Hgb	hemoglobin	PVC	polyvinyl chloride
HOMA	homeostatic model assessment	RBC	red blood cell
HOMA-IR	homeostatic model assessment of insulin resistance	SD	standard deviation
HOME	Health Outcomes and Measures of the Environment	SE	standard error
IgE	immunoglobulin E	SHBG	sex-hormone binding globulin
ICC	intra-class correlation coefficient	T3	triiodothyronine
IM-GSM	grey scale media of the intima media complex	T4	thyroxine
IMT	intima media thickness	TSH	thyroid stimulating hormone
		VO	vaginal opening
		VOC	volatile organic compound
		WBC	white blood cell
		WHO	World Health Organization

PREFACE

This draft document presents preliminary materials for an assessment of diisobutyl phthalate (DIBP) prepared by the U.S. Environmental Protection Agency’s (EPA’s) Integrated Risk Information System (IRIS) Program. These preliminary materials include a planning and scoping summary, information on the approaches used to identify pertinent literature, results of the literature search, approaches for selection of studies for hazard identification, presentation of critical studies in evidence tables and exposure-response arrays, and mechanistic information for DIBP. This material is being released for public review and comment prior to a public meeting, providing an opportunity for the IRIS Program to engage in early discussions with stakeholders and the public on data that may be used to identify adverse health effects and characterize dose-response relationships.

The planning and scoping summary includes information on the uses of DIBP, occurrence of DIBP in the environment, and the rationale and scope for the development of the assessment. This information is responsive to recommendations in the 2009 National Research Council (NRC) report *Science and Decisions: Advancing Risk Assessment* ([NRC, 2009](#)) related to planning and scoping in the risk assessment process.

The preliminary materials are also responsive to the 2011 NRC report *Review of the Environmental Protection Agency’s Draft IRIS Assessment of Formaldehyde* ([NRC, 2011](#)). The IRIS Program’s implementation of the NRC recommendations is following a phased approach that is consistent with the NRC’s “Roadmap for Revision” as described in Chapter 7 of the formaldehyde review report. The NRC stated that “the committee recognizes that the changes suggested would involve a multi-year process and extensive effort by the staff of the National Center for Environmental Assessment and input and review by the EPA Science Advisory Board and others.” Phase 1 of implementation has focused on a subset of the short-term recommendations, such as editing and streamlining documents, increasing transparency and clarity, and using more tables, figures, and appendices to present information and data in assessments. Phase 1 also focused on assessments near the end of the development process and close to final posting. Phase 2 of implementation is focused on assessments that are in the beginning stages of assessment development. The IRIS DIBP assessment is in Phase 2 and represents a significant advancement in implementing the NRC recommendations. In the development of this assessment, many of the recommendations are being implemented in full, while others are being implemented in part. Achieving full and robust implementation of certain recommendations will be an evolving process with input and feedback from the public, stakeholders, and independent external peer review. Phase 3 of implementation will incorporate the longer-term recommendations made by the NRC, including the development of a standardized approach to describe the strength of evidence for noncancer effects.

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1 In May 2014, the NRC released their report reviewing the IRIS assessment development
2 process. As part of this review, the NRC reviewed current methods for evidence-based reviews and
3 made several recommendations with respect to integrating scientific evidence for chemical hazard
4 and dose-response assessments. In their report, the NRC states that EPA should continue to
5 improve its evidence-integration process incrementally and enhance the transparency of its
6 process. The committee did not offer a preference but suggests that EPA consider which approach
7 best fits its plans for the IRIS process. The NRC recommendations will inform the IRIS Program's
8 efforts in this area going forward. This effort is included in Phase 3 of EPA's implementation plan.

9 The literature search strategy, which describes the processes for identifying scientific
10 literature, screening studies for consideration, and identifying primary sources of health effects
11 data, is responsive to NRC recommendations regarding the development of a systematic and
12 transparent approach for identifying the primary literature for analysis. The preliminary materials
13 also describe EPA's approach for the selection of critical studies to be included in the evidence
14 tables, as well as the approach for evaluating methodological features of studies that will be
15 considered in the overall evaluation and synthesis of evidence for each health effect. The
16 development of these materials is in response to the NRC recommendation to thoroughly evaluate
17 critical studies with standardized approaches that are formulated and based on the type of research
18 (e.g., observational epidemiology or animal bioassays). In addition, NRC recommendations for
19 standardized presentation of key study data are addressed by the development of the preliminary
20 evidence tables and preliminary exposure-response arrays for primary health effect information.

21 EPA welcomes all comments on the preliminary materials in this document, including the
22 following:

- 23 • the clarity and transparency of the materials;
- 24 • the approach for identifying pertinent studies;
- 25 • the selection of critical studies for data extraction to preliminary evidence tables and
26 exposure-response arrays;
- 27 • any methodological considerations that could affect the interpretation of or confidence in
28 study results; and
- 29 • any additional studies published or nearing publication that may provide data for the
30 evaluation of human health hazard or dose-response relationships.

31 The preliminary evidence tables and exposure-response arrays should be regarded solely as
32 representing the data on each endpoint that have been identified as a result of the draft literature
33 search strategy. They do not reflect any conclusions as to hazard identification or dose-response
34 assessment.

35 After obtaining public input and conducting additional study evaluation and data
36 integration, EPA will revise these materials to support the hazard identification and dose-response
37 assessment in a draft Toxicological Review that will be made available for public comment.

1. INTRODUCTION

This introduction contains a planning and scoping summary for the Integrated Risk Information System (IRIS) assessment of diisobutyl phthalate (DIBP). The planning and scoping summary includes information on the properties, sources, and uses of DIBP, occurrence and fate of DIBP in the environment, potential for human exposure, and the rationale for the development of this assessment.

1.1. DIBP IN THE ENVIRONMENT

1.1.1. Production and Use

DIBP (Figure 1-1) is used as a plasticizer (HSDB, 2013) in a wide range of materials including polyvinyl chloride (PVC) formulations; paints; lacquers; varnish; paper, pulp and board industry; as a softener; in viscosity adjustment; nail polish; cosmetics; lubricants; carpets; clothing treatments; rubber dentistry settings; as a fuel stabilizer; as a concrete additive; explosive materials; and printing inks. DIBP has also been classified by the Food and Drug Administration (FDA) as an indirect food additive through its use as a component of adhesives. Because DIBP has similar properties to di-n-butyl phthalate (DBP), it can be used as a substitute for DBP (HSDB, 2013). Approximately 500,000 pounds were manufactured in the United States in 2012 (<http://www.epa.gov/oppt/cdr/index.html>). In July 2014, the Consumer Product Safety Commission's (CPSC) Chronic Hazard Advisory Panel (CHAP) recommended that DIBP be permanently banned from use in children's toys and child care articles at levels greater than 0.1% (CHAP, 2014).

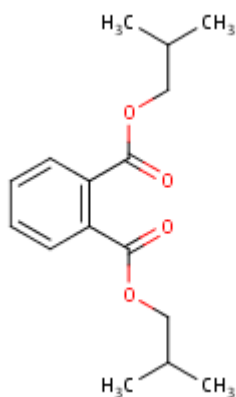


Figure 1-1. Chemical structure of DIBP (HSDB, 2013).

1 **1.1.2. Environmental Fate**

2 If released to air, DIBP will exist in both the vapor and particulate phases in the atmosphere.
3 Vapor-phase DIBP will be photolytically degraded with a half-life of about 1.2 days, and particulate-
4 phase DIBP will be removed from the atmosphere by wet or dry deposition ([HSDB, 2013](#)). In soil,
5 DIBP is expected to have low mobility due to a moderately high organic carbon partition coefficient
6 (Koc). Biodegradation in aerobic soil and water is expected to occur over days or weeks. Anaerobic
7 biodegradation rates are expected to be slower. Volatilization from moist soil or water is expected
8 to be an important fate process for DIBP, but volatilization from dry soil is not expected. If released
9 into water, DIBP is expected to adsorb to sediments and solids, and volatilization from water
10 surfaces is expected to be an important process. An estimated bioconcentration factor of 240
11 suggests that there is a potential for the chemical to concentrate in aquatic organisms, but
12 metabolism in the organisms can reduce accumulation ([HSDB, 2013](#)). As noted by [Wormuth et al.](#)
13 [\(2006\)](#), the majority of phthalates that are found in the environment come from their slow releases
14 from plastics and other phthalate-containing articles. Certain waste streams, sludges, and
15 industrially contaminated sites, however, may contain higher levels of phthalates than other sites.

16 **1.1.3. Human Exposure Pathways**

17 The routes by which humans are exposed to phthalates and the magnitude of individual
18 phthalate exposures have changed over time as the quantities and uses of the various phthalates
19 have changed. Human exposure to phthalates occurs mainly in occupational or household settings
20 because they are used and released from products in the home environment. Environmental
21 concentrations of phthalates are typically the highest in house dust, and they may be present in
22 food due to the use of phthalates in packaging and food preparation materials. For most phthalates,
23 food ingestion is the dominant pathway of exposure, with dust exposures (ingestion and dermal
24 contact) and inhalation also being important in some circumstances. Infant and toddler exposures
25 occur due to teething and playing with plastic toys that contain phthalates ([Wormuth et al., 2006](#)).

26 The presence of parent phthalates or their metabolites in a body matrix, such as blood or
27 urine, provides evidence of exposure to that chemical. The predominant metabolite of DIBP in
28 humans is monoisobutyl phthalate (MIBP). [Zota et al. \(2014\)](#) evaluated the prevalence and
29 temporal trends of MIBP in urine samples collected as part of the National Health and Nutrition
30 Examination Survey (NHANES) conducted between 2001 and 2010. MIBP was found in 72% of the
31 samples in the 2001–2002 cycle and 96% of the samples in the 2009–2010 cycle, and increased in
32 concentration over time, starting at about 2.4 ng/mL in the 2001–2002 cycle, and rising to about
33 7.8 ng/mL in the 2009–2010 cycle.

34 Intake exposures can be estimated on a pathway-basis by combining exposure media
35 concentrations and contact rates. Using this approach, [Clark et al. \(2011\)](#) estimated median intakes
36 of DIBP for various lifestages as defined by the authors: between 0.75 and 1.0 µg/kg-day for teens
37 (12–19 years of age) and adults (20–70 years of age), based on ingestion of food, drinking water,
38 dust/soil, and inhalation of air; and between 1.3 and 2.6 µg/kg-day for infants (0–0.5 years of age),
39 toddlers (ages 0.5–4 years of age), and children (5–11 years of age). The exposure was found to be

1 dominated by food, with inhalation of indoor air also important. The intakes determined by [Clark](#)
2 [et al. \(2011\)](#) were higher than those found by [Wormuth et al. \(2006\)](#), who determined intakes for
3 these age ranges at about ≤ 0.5 $\mu\text{g}/\text{kg}\text{-day}$. [Clark et al. \(2011\)](#) attributed this difference to use of
4 higher food concentrations in the estimates.

5 [Wittassek et al. \(2011\)](#) reported median intakes of DIBP in the range of 0.1–1.7 $\mu\text{g}/\text{kg}\text{-day}$
6 based on a literature survey of urinary biomonitoring data and intake estimates provided therein.
7 Their review included a single study in the United States of a cohort of pregnant woman that found
8 median intakes at 0.1 $\mu\text{g}/\text{kg}\text{-day}$. Three other studies from Germany had median intakes ranging
9 from 1.1 to 1.7 $\mu\text{g}/\text{kg}\text{-day}$. [Qian et al. \(2014\)](#) used NHANES 2007–2008 and found a median intake
10 of 0.2 $\mu\text{g}/\text{kg}\text{-day}$ and a 95th percentile intake of 0.9 $\mu\text{g}/\text{kg}\text{-day}$. [Christensen et al. \(2014b\)](#) combined
11 the data from NHANES 2005–2008 and found similar results to [Qian et al. \(2014\)](#), with a median
12 over that time span of 0.2 $\mu\text{g}/\text{kg}\text{-day}$ and a 95th percentile intake of 0.8 $\mu\text{g}/\text{kg}\text{-day}$.

13 1.2. SCOPE OF THE ASSESSMENT

14 The National Research Council (NRC) has recommended that, “[c]umulative risk assessment
15 based on common adverse outcomes is a feasible and physiologically relevant approach to the
16 evaluation of the multiplicity of human exposures and directly reflects EPA’s mission to protect
17 human health” ([NRC, 2008, p11](#)). They envisioned facilitating the process by “defining the groups
18 of agents that should be included for a given outcome” ([NRC, 2008, p12](#)). In humans, the NRC cited
19 results from NHANES that demonstrate exposure to multiple phthalates in most people ([NRC, 2008,](#)
20 [p23-25](#)). A recent review of human exposure to eight phthalates estimated that indoor air
21 contributed to approximately 25% of DIBP exposure in children ([CHAP, 2014, Appendix E1, p35](#)).
22 The unique exposure scenarios and potential sensitivities of children contribute to the need for an
23 assessment of phthalate toxicity. This IRIS assessment will help to inform EPA programs and
24 regions of the potentially unique vulnerabilities of children to DIBP exposure and enable future
25 cumulative risk assessments that assess effects on human health outcomes that might be associated
26 with DIBP and other phthalates. There is currently no IRIS assessment of DIBP.

2. METHODS FOR IDENTIFYING AND SELECTING STUDIES

The [NRC \(2011\)](#) recommended that the U.S. Environmental Protection Agency (EPA) develop a detailed search strategy utilizing a graphical display documenting how initial search findings are narrowed to the final studies that are selected for further evaluation on the basis of inclusion and exclusion criteria. Following these recommendations, a literature search and screening strategy was applied to identify literature related to characterizing the health effects of diisobutyl phthalate (DIBP). This strategy consisted of a search of online scientific databases and other sources, casting a wide net in order to identify all potentially pertinent studies. In subsequent steps, references were screened to exclude papers not pertinent to an assessment of the health effects of DIBP, and remaining references were sorted into categories for further evaluation. Section 2.1 describes the literature search and screening strategy in detail. The [NRC \(2011\)](#) further recommended that after studies are identified for review by utilizing a transparent search strategy, the next step is to summarize the details and findings of the most pertinent studies in the evidence tables. The NRC suggested that such tables should provide a link to the references, and include details of the study population, methods, and key findings. This approach provides for a systematic and concise presentation of the evidence. The NRC also recommended that the methods and findings should then be evaluated with a standardized approach. The approach that was outlined identified standard issues for the evaluation of epidemiological and experimental animal studies. Section 2.2 describes the approach taken for DIBP for selecting studies to be included in the preliminary evidence tables and exposure-response arrays. Section 3 presents the selected studies in preliminary evidence tables and exposure-response arrays, arranged by health effect.

2.1. DRAFT LITERATURE SEARCH AND SCREENING STRATEGY

The literature search for DIBP was conducted in four online scientific databases (PubMed, Web of Science, Toxline, and Toxic Substances Control Act Test Submissions (TSCATS2)) in February of 2013; the search was repeated in March of 2014. This document is complete through March 2014. Additional updates will be performed at regular (e.g., 6-month) intervals. The detailed search approach, including the search strings and number of citations identified per database, is presented in Table 2-1. The search strings and search terms described for DIBP captured studies using the parent compound and metabolites (i.e., the active metabolite, monoisobutyl phthalate [MIBP]). This search of online databases identified 504 citations (after electronically eliminating duplicates). The computerized database searches were also supplemented by a manual search of citations from other regulatory documents (Table 2-2);

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1 343 citations were obtained using these additional search strategies. In total, 809 citations were
 2 identified using online scientific databases and additional search strategies.

3

4 **Table 2-1. Database search strategy for DIBP**

Database (search date)	Keywords ^a
PubMed 03/2014 02/2013	dibp OR (mibp AND phthalate) OR "diisobutylphthalate" OR "di-isobutyl phthalate" OR "84-69-5" OR "diisobutyl phthalate" OR "di(i-butyl)phthalate" OR "di-iso-butyl phthalate" OR "isobutyl phthalate" OR "phthalic acid diisobutyl ester" OR ("diisobutyl ester" AND phthalate) OR "1,2-benzenedicarboxylic acid bis(2-methylpropyl) ester" OR "1,2-benzenedicarboxylic acid 1,2-bis(2-methylpropyl) ester" OR "monoisobutyl phthalate" OR "mono(i-butyl)phthalate" OR "mono-iso-butyl phthalate" OR "phthalic acid monoisobutyl ester" OR "1,2-benzenedicarboxylic acid, mono(2-methylpropyl) ester" OR "2-[(2-methylpropoxy)carbonyl]benzoic acid" OR "1,2-benzenedicarboxylic acid, mono(2-methylpropyl) ester (9CI)" OR "isobutyl hydrogen phthalate" OR "1,2-benzenedicarboxylic acid 1-(2-methylpropyl) ester"
Web of Science 03/2014 02/2013	TS=dibp OR (TS=mibp AND TS=phthalate) OR TS="diisobutylphthalate" OR TS="di-isobutyl phthalate" OR TS="84-69-5" OR TS="diisobutyl phthalate" OR TS="di(i-butyl)phthalate" OR TS="di-iso-butyl phthalate" OR TS="isobutyl phthalate" OR TS="phthalic acid diisobutyl ester" OR (TS="diisobutyl ester" AND TS=phthalate) OR TS="1,2-benzenedicarboxylic acid bis(2-methylpropyl) ester" OR TS="1,2-benzenedicarboxylic acid 1,2-bis(2-methylpropyl) ester" OR TS="monoisobutyl phthalate" OR TS="mono(i-butyl)phthalate" OR TS="mono-iso-butyl phthalate" OR TS="phthalic acid monoisobutyl ester" OR TS="1,2-benzenedicarboxylic acid, mono(2-methylpropyl) ester" OR TS="2-[(2-methylpropoxy)carbonyl]benzoic acid" OR TS="1,2-benzenedicarboxylic acid, mono(2-methylpropyl) ester (9CI)" OR TS="isobutyl hydrogen phthalate" OR TS="1,2-benzenedicarboxylic acid 1-(2-methylpropyl) ester"
Toxline 03/2014 02/2013	Split into 4 separate search strings: @TERM+@rn+84-69-5 @AND+mibp+phthalate @AND+"diisobutyl ester"+phthalate @OR+(dibp+"diisobutylphthalate"+"di-isobutyl+phthalate"+"diisobutyl+phthalate"+"di(i-butyl)phthalate"+"di-iso-butyl+phthalate"+"isobutyl+phthalate"+"phthalic+acid+diisobutyl+ester"+"1,2-benzenedicarboxylic+acid+bis(2-methylpropyl)+ester"+"1,2-benzenedicarboxylic+acid+1,2-bis(2-methylpropyl)+ester"+"monoisobutyl+phthalate"+"mono(i-butyl)phthalate"+"mono-iso-butyl+phthalate"+"phthalic+acid+monoisobutyl+ester"+"1,2-benzenedicarboxylic+acid,+mono(2-methylpropyl)+ester"+"2-[(2-methylpropoxy)carbonyl]benzoic+acid"+"1,2-benzenedicarboxylic+acid,+mono(2-methylpropyl)+ester+(9CI)"+"isobutyl+hydrogen+phthalate"+"1,2-benzenedicarboxylic+acid+1-(2-methylpropyl)+ester")
TSCATS2 03/2014	(2000-) 84-69-5

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6 ^aThe search strings and search terms described above captured studies using the parent compound and the
 7 metabolite MIBP.

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Table 2-2. Summary of additional search strategies for DIBP

System used	Selected key reference(s) or sources	Date	Additional references identified
Manual search of citations from regulatory documents	CPSC (2010) . Toxicity Review for Diisobutyl phthalate (DIBP). Bethesda, MD: Consumer Product Safety Commission.	3/2014	9 citations added
Web of Science, forward search	Hannas et al. (2011) . Dose-response assessment of fetal testosterone production and gene expression levels in rat testes following in utero exposure to diethylhexyl phthalate, diisobutyl phthalate, diisoheptyl phthalate, and diisononyl phthalate. <i>Toxicol Sci.</i> 123(1):206-16.	3/2014	2 citations added
	Saillenfait et al. (2008) . Diisobutyl phthalate impairs the androgen-dependent reproductive development of the male rat. <i>Reprod Toxicol.</i> 26(2):107-15.	3/2014	1 citation added
	Ray et al. (2012) . Ovarian development in Wistar rat treated prenatally with single dose diisobutyl phthalate. <i>Bratisl Lek Listy.</i> 113(10):577-82.	3/2014	0 citations added
	Kleinsasser et al. (2001a) . Genotoxicity of di-butyl-phthalate and di-iso-butyl-phthalate in human lymphocytes and mucosal cells. <i>Teratog Carcinog Mutagen.</i> 21(3):189-96.	3/2014	1 citation added
Web of Science, backward search	Hannas et al. (2011) . Dose-response assessment of fetal testosterone production and gene expression levels in rat testes following in utero exposure to diethylhexyl phthalate, diisobutyl phthalate, diisoheptyl phthalate, and diisononyl phthalate. <i>Toxicol Sci.</i> 123(1):206-16.	3/2014	1 citation added
	Saillenfait et al. (2008) . Diisobutyl phthalate impairs the androgen-dependent reproductive development of the male rat. <i>Reprod Toxicol.</i> 26(2):107-15.	3/2014	1 citation added
	Ray et al. (2012) . Ovarian development in Wistar rat treated prenatally with single dose diisobutyl phthalate. <i>Bratisl Lek Listy.</i> 113(10):577-82.	3/2014	4 citations added
	Kleinsasser et al. (2001a) . Genotoxicity of di-butyl-phthalate and di-iso-butyl-phthalate in human lymphocytes and mucosal cells. <i>Teratog Carcinog Mutagen.</i> 21(3):189-96.	3/2014	2 citations added
Snowball search	DIBP references in previous assessment or previously added to the HERO project page	4/2014	45 citations added
Background Check	Searched a combination of CASRNs and synonyms on the following databases: ACGIH (http://www.acgih.org/home.htm) ATSDR (http://www.atsdr.cdc.gov/substances/index.asp) CalEPA Office of Environmental Health Hazard Assessment (http://www.oehha.ca.gov/risk.html)	2/2013, update 3/2014	17 citations added

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System used	Selected key reference(s) or sources	Date	Additional references identified
	<p>OEHHA Toxicity Criteria Database http://www.oehha.ca.gov/tcdb/index.asp Biomonitoring California-Priority Chemicals http://www.oehha.ca.gov/multimedia/biomon/pdf/PriorityChemsCurrent.pdf Biomonitoring California-Designated Chemicals http://www.oehha.ca.gov/multimedia/biomon/pdf/DesignatedChemCurrent.pdf Cal/ECOTOX database http://www.oehha.ca.gov/scripts/cal_ecotox/CHEMLIST.ASP OEHHA Fact Sheets http://www.oehha.ca.gov/public_info/facts/index.html Non-cancer health effects Table (RELs) and Cancer Potency Factors (Appendix A and Appendix B) http://www.oehha.ca.gov/air/hot_spots/index.html</p> <p>CPSC (http://www.cpsc.gov)</p> <p>eChemPortal http://www.echemportal.org/echemportal/participant/page.action?pageID=9</p> <p>Environment Canada – Search entire site if not found below: http://www.ec.gc.ca/default.asp?lang=En&n=ECD35C36</p> <p>Toxic Substances Managed under CEPA http://www.ec.gc.ca/toxiques-toxics/Default.asp?lang=En&n=98E80CC6-1 Screening Assessment reports Risk Management reports Final Assessments (http://www.ec.gc.ca/lcpe-cepa/default.asp?lang=En&xml=09F567A7-B1EE-1FEE-73DB-8AE6C1EB7658) Draft Assessments (http://www.ec.gc.ca/lcpe-cepa/default.asp?lang=En&xml=6892C255-5597-C162-95FC-4B905320F8C9)</p> <p>EPA Acute Exposure Guideline Levels http://www.epa.gov/oppt/aegl/pubs/chemlist.htm</p> <p>EPA – IRISTrack/New Assessments and Reviews EPA NSCEP (http://www.epa.gov/ncepihom/) EPA RfD/RfC and CRAVE meeting notes EPA Science Inventory (http://cfpub.epa.gov/si/) FDA (http://www.fda.gov/) Federal Docket (www.regulations.gov) Health Canada First Priority List Assessments (http://www.hc-sc.gc.ca/ewh-semt/pubs/contaminants/psl1-lsp1/index-eng.php) Health Canada Second Priority List Assessments http://www.hc-sc.gc.ca/ewh-semt/pubs/contaminants/psl2-lsp2/index-eng.php</p> <p>IARC (http://monographs.iarc.fr/htdig/search.html)</p>		

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System used	Selected key reference(s) or sources	Date	Additional references identified
	ITER (TERA database) http://iter.ctcnet.net/publicurl/pub_search_list.cfm NAP – Search Site (http://www.nap.edu/) NRC – AEGLs via NAP search for “Acute Exposure Guideline Level” and the chemical NCI (http://www.cancer.gov) National Institute for Environmental Health Sciences (NIEHS) http://www.niehs.nih.gov/ NICNAS (PEC only covered by eChemPortal) http://www.nicnas.gov.au/industry/aics/search.asp NIOSH (http://www.cdc.gov/niosh/topics/) NIOSHTIC 2 (http://www2a.cdc.gov/nioshtic-2/) NTP - RoC, status, results, and management reports http://ntpsearch.niehs.nih.gov/query.html OSHA http://www.osha.gov/dts/chemicalsampling/toc/toc_chemsam.html RTECS http://www.ccohs.ca/search.html		

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2 These citations were screened using the title, abstract, and in limited instances, full text for
3 pertinence to examining the health effects of DIBP exposure. The citations were then screened
4 using inclusion criteria (Table 2-3) describing specific information to help identify primary source
5 health effect data and mechanistic and/or genotoxic data, as well as resources useful in preparation
6 of the DIBP package. The process for screening the literature search is described below and is
7 shown graphically in Figure 2-1:

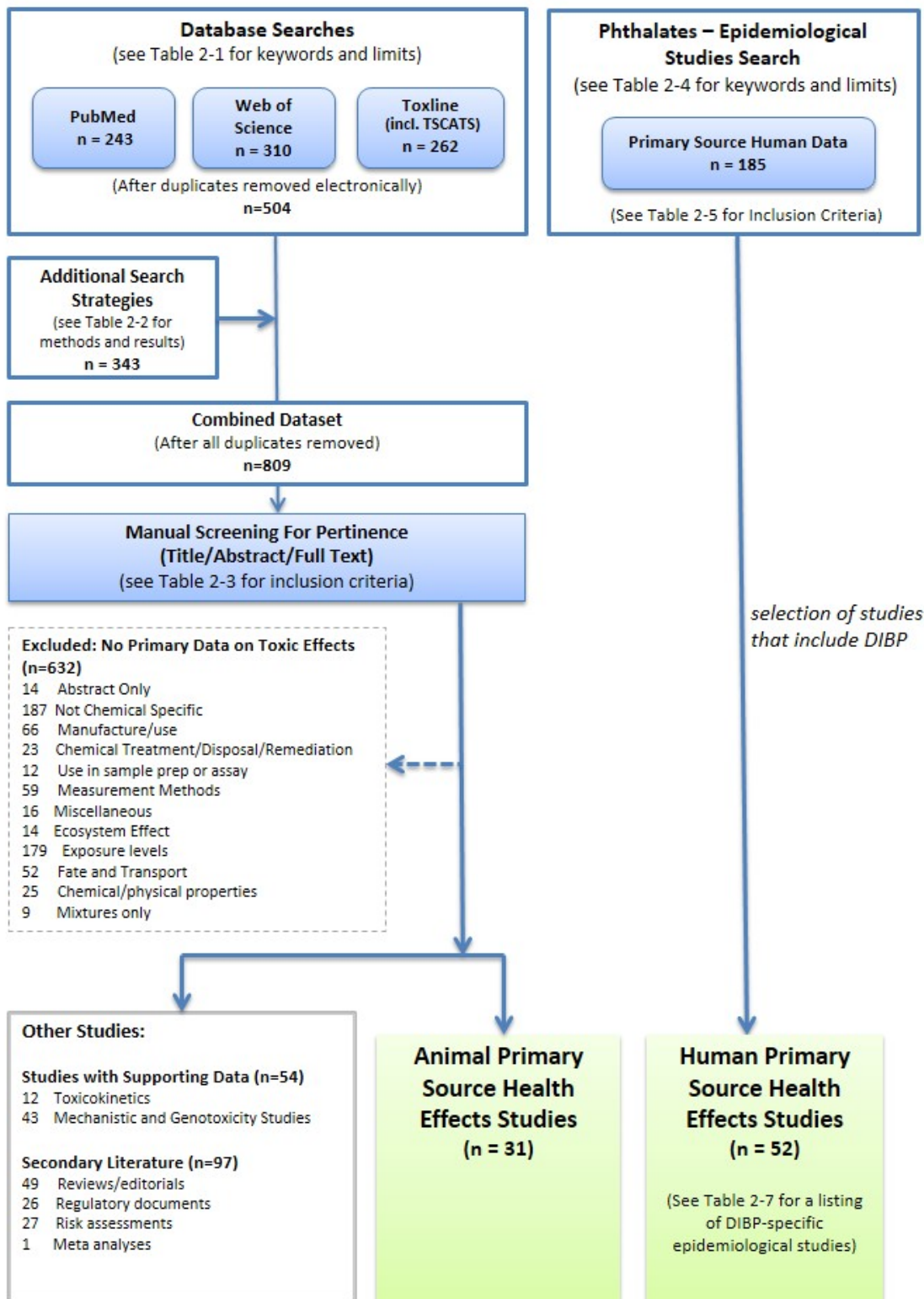
- 8 • 31 references were identified as animal studies with health effects data and were
9 considered for data extraction to evidence tables and exposure-response arrays.
- 10 • 54 references were identified as supporting studies; of these, 12 were toxicokinetic studies
11 and 43 were mechanistic and genotoxicity studies.
- 12 • 97 references were identified as secondary literature (e.g., reviews and editorials, risk
13 assessments, meta analyses, and regulatory documents); these references were kept as
14 additional resources for development of the Toxicological Review.
- 15 • 632 references were excluded because these studies did not include primary source data
16 evaluating DIBP in relation to any kind of toxicity or health endpoint, and did not provide
17 either supporting information (e.g., toxicokinetic or mechanistic/genotoxicity data) or
18 secondary literature information (see Figure 2-1 and Table 2-3 for inclusion categories and
19 criteria).

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1 Note that some studies were identified as belonging to multiple categories. As a result, the
2 total number of studies in a given category may be less than the sum of the individual studies listed
3 in subcategories. For example, the category “Studies with Supporting Data” included one study that
4 contained information relevant to both the toxicokinetics and mechanistic and/or genotoxicity
5 subcategories.

6 Among the studies identified in the DIBP literature searches, there were a number of
7 foreign language studies. Based on a review of the English titles and, when available, English
8 abstracts, two of the foreign language articles, [Ma et al. \(2013b\)](#) and [Iijo \(1975\)](#), were tagged as
9 toxicity studies and four foreign language articles, [Ma et al. \(2010\)](#), [Ma et al. \(2013c\)](#), [Kleinsasser et](#)
10 [al. \(1999\)](#), and [Kleinsasser et al. \(2001b\)](#), were tagged as mechanistic and genotoxicity studies. The
11 other foreign language articles were excluded (tagged to Excluded: No primary data on toxic
12 effects). [Ma et al. \(2013b\)](#) is a report of neurotoxicological effects after DIBP exposure. With the
13 exception of one study ([University of Rochester, 1954](#)) that assessed brain weight, the [Ma et al.](#)
14 [\(2013b\)](#) article was the only available neurotoxicological study; this article was translated into
15 English ([certified translation; Ma et al., 2013a](#)). The remaining five foreign language articles
16 (above), tagged to toxicity studies or mechanistic and genotoxicity studies, have not yet been
17 translated or considered for inclusion in either evidence or mechanistic tables. These studies will
18 be further evaluated and considered during the development of the draft assessment of the
19 available evidence of DIBP-induced health effects.
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2 Note: Studies containing multiple information categories were sorted into multiple tags. For this reason, the
3 subcategory numbers do not always add up to the category total.

4 **Figure 2-1. Literature search approach for DIBP.**

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Table 2-3. Inclusion criteria used to identify animal studies of health-related endpoints, supporting data, or secondary literature

Inclusion criteria ^a
<ul style="list-style-type: none">• Did the study evaluate effects of DIBP or its metabolites known to be formed in humans?• Did the study evaluate effects in a tissue (organ) or cells derived from a tissue (organ)?• Did the study evaluate cellular, biochemical or molecular effects relevant to any mode of action? <p style="text-align: center;">or</p> <ul style="list-style-type: none">• Does the study include information from other agencies, risk assessments, or reviews that would aid in the development of a toxicological review of DIBP?

^aIf the answer is “no” to any of these criteria questions, the study was placed under “No Primary Data on Toxic Effects.”

Thirty-six human studies were also identified from the initial literature search using the search strings presented in Table 2-1. However, work being done concurrently on the development of other phthalate preliminary materials revealed that this set of DIBP epidemiology studies was incomplete. Epidemiology studies frequently examine multiple compounds (e.g., metabolites of several different phthalates). The indexing terms and abstracts may not include a comprehensive list of all of the specific phthalates examined, resulting in the inappropriate exclusion of studies and the potential for introduction of bias in the selection process. Specifically, “negative” studies (i.e., studies that did not demonstrate an association between exposure and disease) are potentially more likely to be missed than “positive” studies. This issue did not arise in the search process for experimental (animal toxicology) studies, for which the test compound is virtually always identified through search terms or key word searches of abstracts.

Another issue encountered in the development of the search and screening process for the phthalate epidemiology studies relates to the duplication of efforts involved in the development of EPA’s health assessments for several individual phthalates (e.g., dibutyl phthalate [DBP], DIBP, butyl benzyl phthalate [BBP], di(2-ethylhexyl)phthalate [DEHP], di-ethyl phthalate [DEP], diisononyl phthalate [DINP], and dipentyl phthalate [DPP]). In contrast to animal toxicology studies, most of the epidemiology studies examine more than one phthalate, resulting in considerable overlap in the sets of studies identified using individual-phthalate search terms. Full text screening of the same studies identified in multiple searches results in an inefficient use of resources.

For these reasons, EPA developed a process for identifying epidemiological studies evaluating phthalates by performing a single broad search to create a listing of epidemiological studies of all phthalates mentioned above, from which the selection of studies examining potential health effects of an individual phthalate could be drawn. This list records each of the phthalates included in the study, based on information in the methods section of the paper, and the outcome(s)

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1 examined. This literature search for epidemiological studies examining phthalates in relation to
 2 health-related endpoints (from which the DIBP studies were drawn) was conducted in PubMed,
 3 Web of Science, and ToxNet databases in June 2013, using keywords and limits described in
 4 Table 2-4; the search was updated in December 2013 and in June 2014. For this search, “phthalate”
 5 (and related terms) rather than names of specific phthalates was used as the foundation of the
 6 search, along with terms designed specifically to identify epidemiological studies. These terms
 7 were based on terms used in previously identified epidemiology studies of six different phthalates.

8 **Table 2-4. Summary of search terms: targeted epidemiology search**

Database, search date	Terms	Hits
June 2013 search PubMed 06/2013 No date restriction	(phthalate OR phthalates OR phthalic acid) AND (human OR case-control OR pregnancy OR cohort OR workers OR children OR survey)	Imported: 2,505 After duplicates deleted: 2,482
Web of Science 06/2013 No date restriction	(TS="phthalic acid" OR TS="phthalate" OR TS="phthalates") AND (TS="humans" OR TS="human" OR TS="case-control" OR TS="pregnancy" OR TS="cohort" OR TS="workers" OR TS="child" OR TS="children" OR TS="survey")	Imported: 1,840 After duplicates deleted: 1,836
ToxNet 06/2013 No date restriction	(phthalate OR phthalates OR phthalic acid) AND (human OR case-control OR pregnancy OR cohort OR workers OR children OR survey)	Imported: 2,505 After duplicates deleted: 2,426
Merged Reference Set	Merged dataset, with duplicates eliminated through electronic screen	4,127
	Epidemiology articles meeting inclusion criteria	127
December 2013 search	PubMed Web of Science ToxNet Merged Reference Set Additional epidemiology articles meeting inclusion criteria	155 249 114 350 22
June 2014 search	PubMed Web of Science ToxNet (was not searched because no articles have been found solely through this source in all the previous searches) Merged Reference Set Additional epidemiology articles meeting inclusion criteria	184 409 0 494 24

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1 More than 4,000 citations were identified through this search. These were then screened
2 using inclusion criteria describing specific population (i.e., human), exposure measures,
3 comparison, and health effects (Table 2-5). Note that other studies obtained in the search, for
4 example mechanistic and pharmacokinetic studies, are excluded from consideration with respect to
5 the specific objective of this search (i.e., identification of epidemiology studies), but could be
6 included in other steps in the assessment. Duplicate citations of the same article were excluded,
7 and articles written in a language other than English were retained for subsequent review. Earlier
8 analyses that are updated in a subsequent paper (e.g., with a larger sample size) are not included as
9 a primary paper, but may be used as background material regarding study methods.

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1 **Table 2-5. Inclusion criteria used to identify epidemiology studies of health-**
 2 **related endpoints**

Inclusion criteria
<ul style="list-style-type: none"> • Is the study population humans? <div style="text-align: center;">and</div> • Is exposure to one or more phthalate (parent compound or metabolite(s)^a... <ul style="list-style-type: none"> - measured in air, dust, or biological tissue? - based on knowledge of industrial hygiene (occupational settings)? - based on knowledge of specific contamination sites or accidental exposure? <div style="text-align: center;">and</div> • Does the study compare a health effect in higher versus lower or no exposure? <div style="text-align: center;">and</div> • Does the study include a measure of one or more primary health effect endpoints relating to... <ul style="list-style-type: none"> - sexual differentiation measures (e.g., male genital malformations, anogenital distance, gender-related play behavior) - male reproductive effects (e.g., steroidal and gonadotropin hormone levels, measures of male-mediated infertility)? - female reproductive effects (e.g., steroidal and gonadotropin hormone levels, measures of female-mediated infertility, gynecological conditions)? - pregnancy outcomes (e.g., birth weight, gestation age)? - puberty (male and female) (e.g., timing of development, precocious puberty, gynecomastia)? - neurodevelopment (infants and children) (e.g., standardized tests of reflexes, behavior, and intelligence)? - thyroid effects (e.g., thyroid stimulating hormone and thyroid hormones, subclinical and clinical thyroid disease)? - immune system effects (e.g., asthma, allergies, immunoglobulin E (IgE) levels, skin prick tests)? - pulmonary function (e.g., standardized test of lung volume, diffusing capacity)? - neurological effects (adults) (e.g., peripheral neuropathy, vision or hearing or other sensory tests)? - liver effects (e.g., cholestasis, biomarkers of liver function)? - kidney effects (e.g., end stage renal disease, biomarkers of kidney function)? - diabetes and measures of insulin resistance? - obesity (and other measures of adiposity)? - cardiovascular disease (cause-specific incidence or mortality)? - cardiovascular risk factors (e.g., triglyceride and lipid levels, blood pressure or hypertension)? - cancer (cause-specific incidence or mortality)? <div style="text-align: center;">or</div> • Does the study include a measure of one or more secondary health effect endpoints (to be considered within context of mechanistic evidence) relating to... <ul style="list-style-type: none"> - oxidative stress? - inflammation? - gene expression?

3
 4 ^aFor DIBP, the primary metabolite of interest is MIBP.
 5

6 One hundred and seventy-three epidemiological studies examining one or more phthalates
 7 in relation to one or more endpoints were identified by the searches conducted through June 2014

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1 (127 in the initial search, 22 in the December 2013 update, and 24 in the June 2014 update;
 2 Figure 2-1). Other strategies were also used to supplement this broad search for epidemiology
 3 studies of phthalates), resulting in the identification of 12 additional publications (Table 2-6), for a
 4 total of 185 epidemiological studies. From this set of all of the epidemiological studies examining
 5 any phthalate, 52 studies analyzed one or more health effects in relation to a measure of DIBP
 6 (Table 2-7).

7 **Table 2-6. Summary of additional search strategies for epidemiology studies**
 8 **of phthalate exposure in relation to health-related endpoints**

Approach used	Date performed	Number of additional citations identified
Testing and refinement of search terms based on terms used for the identified articles within each category	June 2014	6
Review of references cited in the identified list of epidemiology studies (“backward” search)	July 2014	1
Electronic forward search through Web of Science of one to three studies within each health endpoint category (early studies within each category generally selected to maximize potential for citation in subsequent publications) ^a	July 2014	5

9
 10 ^aThe following studies were used to conduct the forward searches: ([Trasande et al. \(2013c\)](#); [James-Todd et al. \(2012\)](#); [Lind and Lind \(2011\)](#); [Boas et al. \(2010\)](#); [Cho et al. \(2010\)](#); [Engel et al. \(2010\)](#); [Lopez-Carrillo et al. \(2010\)](#); [Wolff et al. \(2010\)](#); [Adibi et al. \(2009\)](#); [Chou et al. \(2009\)](#); [Hatch et al. \(2008\)](#); [Wolff et al. \(2008\)](#); [Meeker et al. \(2007\)](#); [Stahlhut et al. \(2007\)](#); [Hauser et al. \(2006\)](#); [Reddy et al. \(2006\)](#); [Jonsson et al. \(2005\)](#); [Swan et al. \(2005\)](#); [Bornehag et al. \(2004\)](#); [Hoppin et al. \(2004\)](#); [Aschengrau et al. \(1998\)](#); [Heineman et al. \(1992\)](#); [Nielsen et al. \(1989\)](#); [Nielsen et al. \(1985\)](#)).

17 **TABLE 2-7. Primary source epidemiological studies examining health effects of**
 18 **DIBP**

Outcome category	Reference ^a	DIBP measure
Sexual differentiation measures (Table 3-1)	Swan (2008) Swan et al. (2010)	MIBP (maternal urine) MIBP (maternal urine)
Male reproductive (semen parameters, infertility, and hormones) (Tables 3-2 and 3-3)	Buck Louis et al. (2014) Joensen et al. (2012) Kranvogel et al. (2014) Mendiola et al. (2011) Wirth et al. (2008)	MIBP (urine) MIBP (urine) MIBP (urine) MIBP (urine) MIBP (urine)
Male pubertal development (Table 3-4)	Mieritz et al. (2012) Mouritsen et al. (2013b)	MIBP (maternal urine) MIBP (urine) Sum MIBP + MBP (urine) ^a
Female pubertal development (Table 3-5)	Frederiksen et al. (2012) Hart et al. (2013) Lomenick et al. (2010) Mouritsen et al. (2013b)	Sum MIBP + MBP (urine) ^a MIBP (maternal serum) MIBP (urine) Sum MIBP + MBP(urine) ^a

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Outcome category	Reference^a	DIBP measure
Female reproductive (infertility, hormones, gynecological conditions) (Tables 3-6 and 3-7)	Buck Louis et al. (2013) Hart et al. (2013) Sathyanarayana et al. (2014) Upson et al. (2013)	MIBP (urine) MIBP (maternal serum) MIBP (maternal urine) MIBP (urine)
Pregnancy outcomes (fetal growth, preterm birth) (Table 3-8)	Ferguson et al. (2014c) Ferguson et al. (2014a) Huang et al. (2014b) Meeker et al. (2009) Philippat et al. (2012) Wolff et al. (2008)	MIBP (maternal urine) MIBP (maternal urine) DIBP (cord blood) MIBP (maternal urine) MIBP (maternal urine) MIBP (maternal urine)
Immune: allergy (rhinitis, eczema) (Table 3-9)	Ait Bamai et al. (2014) Bornehag et al. (2004) Callesen et al. (2014b) Callesen et al. (2014a) Hoppin et al. (2013) Sun et al. (2009)	DIBP (dust) DIBP (dust) MIBP (urine) DIBP (dust) MIBP (urine) DIBP (dust)
Immune: asthma (Table 3-10)	Ait Bamai et al. (2014) Bertelsen et al. (2013) Callesen et al. (2014b) Callesen et al. (2014a) Hoppin et al. (2013) Sun et al. (2009)	DIBP (dust) MIBP (urine) MIBP (urine) DIBP (dust) MIBP (urine) DIBP (dust)
Neurodevelopment (Table 3-11)	Braun et al. (2014) Engel et al. (2010) Kobrosly et al. (2014) Téllez-Rojo et al. (2013) Whyatt et al. (2012)	MIBP (maternal urine) MIBP (maternal urine) MIBP (maternal urine) MIBP (maternal urine) MIBP (maternal urine)
Thyroid (Table 3-12)	Dirtu et al. (2013) Meeker and Ferguson (2011)	MIBP (urine) MIBP (urine)
Obesity (Table 3-13)	Buser et al. (2014) Dirtu et al. (2013) Hart et al. (2013) Kasper-Sonnenberg et al. (2012) Lind et al. (2012b) Olsén et al. (2012) Svensson et al. (2011) Teitelbaum et al. (2012) Trasande et al. (2013a) Wang et al. (2013)	MIBP (urine) MIBP (urine) MIBP (maternal serum) Sum MIBP + OH-MIBP (urine) MIBP (serum) MIBP (serum) MIBP (urine) MIBP (urine) MIBP (urine) MIBP (urine) MIBP (urine)
Diabetes and insulin resistance (Table 3-14)	Huang et al. (2014a) James-Todd et al. (2012) Lind et al. (2012a) Olsén et al. (2012) Svensson et al. (2011) Trasande et al. (2013b)	MIBP (urine) MIBP (urine) MIBP (serum) MIBP (serum) MIBP (urine) MIBP (urine)
Other cardiovascular disease risk factors (Table 3-15)	Lind and Lind (2011) Shiue (2014) Trasande et al. (2013c)	MIBP (serum) MIBP (urine) MIBP (urine)

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Outcome category	Reference ^a	DIBP measure
	Olsén et al. (2012)	MIBP (serum)
Cancer (Table 3-16)	Lopez-Carrillo et al. (2010)	MIBP (urine)

1
2 ^aIncluded in DIBP tables because in this population, at this time, MIBP concentrations were greater than
3 monobutyl phthalate (MBP) concentrations.
4

5 The literature for both epidemiological and animal studies will be regularly monitored for
6 the publication of new studies. The documentation and results for this supplementary search can
7 be found on the Health and Environmental Research On-line (HERO) website¹
8 (<http://hero.epa.gov/DIBP> and <http://hero.epa.gov/phthalates-humanstudies>).

9 **2.2. SELECTION OF CRITICAL STUDIES IN EARLY STAGES OF DRAFT**
10 **DEVELOPMENT**

11 **2.2.1. General Approach**

12 Each study retained following the literature search and screen was evaluated for aspects of
13 design, conduct, or reporting that could affect the interpretation of results and the overall
14 contribution to the synthesis of evidence for determination of hazard potential. Much of the key
15 information for conducting this evaluation can generally be found in the study's methods section
16 and in how the study results are reported. Importantly, this evaluation does not consider study
17 results or, more specifically, the direction or magnitude of any reported effects. For example,
18 standard issues for evaluation of experimental animal data identified by the NRC and adopted in
19 this approach include consideration of the species and sex of animals studied, dosing information
20 (dose spacing, dose duration, and route of exposure), endpoints considered, and the relevance of
21 the endpoints to the human endpoints of concern. Similarly, observational epidemiologic studies in
22 this approach for evaluation should consider the following:

- 23 • Approach used to identify the study population and the potential for selection bias.
24 • Study population characteristics and the generalizability of findings to other populations.

¹HERO is a database of scientific studies and other references used to develop EPA's risk assessments aimed at understanding the health and environmental effects of pollutants and chemicals. It is developed and managed in EPA's Office of Research and Development (ORD) by the National Center for Environmental Assessment (NCEA). The database includes more than 1,400,000 scientific articles from the peer-reviewed literature. New studies are added continuously to HERO.

Note: The HERO database will be regularly updated as additional references are identified during assessment development. Therefore, the numbers of references (by tag) displayed on the HERO webpage for DIBP may not match the numbers of references identified in Figure 2-1 (current through March 2014).

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- 1 • Approach used for exposure assessment and the potential for information bias, whether
2 differential (nonrandom) or nondifferential (random).
- 3 • Approach used for outcome identification and any potential bias.
- 4 • Appropriateness of analytic methods used.
- 5 • Potential for confounding to have influenced the findings.
- 6 • Precision of estimates of effect.
- 7 • Availability of an exposure metric that is used to model the severity of adverse response
8 associated with a gradient of exposures.

9 To facilitate the evaluation outlined above, evidence tables are constructed that
10 systematically summarize the important information from each study in a standardized tabular
11 format as recommended by the [NRC \(2011\)](#). In general, the evidence tables include all studies that
12 inform the overall synthesis of evidence for hazard potential. At this early stage of study
13 evaluation, the goal is to be inclusive. Exclusion of studies may unnecessarily narrow subsequent
14 analyses by eliminating information that might later prove useful. Premature exclusion might also
15 give a false sense of the consistency of results across the database of studies by unknowingly
16 reducing the diversity of study results. However, there may be situations in which the initial review
17 of the available data will lead to a decision to focus on a particular set of health effects and to
18 exclude others from further evaluation.

19 **2.2.2. Exclusion of Studies**

20 After the literature search was manually screened for pertinence, studies were excluded if
21 fundamental flaws were identified in their design, conduct, or reporting. The DIBP experimental
22 animal database consists of studies designed to examine repeat-dose intraperitoneal or oral toxicity
23 (including subchronic and short-term duration studies) and endpoint-specific toxicities (including
24 reproductive and developmental toxicity). Four studies administered DIBP via the intraperitoneal
25 route of exposure. These studies were excluded from the DIBP evidence tables because the
26 intraperitoneal route of exposure is generally considered less relevant to human health exposure.
27 The remaining studies involved administration of DIBP in the diet or via gavage administration.
28 Acute studies are generally less pertinent for characterizing health hazards associated with chronic
29 exposure. There was one acute study that was excluded from the evidence tables. Two BASF
30 reports identified in the literature searches could not be obtained and thus, could not be evaluated
31 for inclusion in the evidence tables ([BASE, 2003, 1961](#)). For these reasons, these studies are not
32 summarized in the preliminary evidence tables. Nevertheless, with the exception of the studies that
33 could not be obtained, the studies will still be evaluated as possible sources of supporting health
34 effects information during assessment development. Experimental animal studies that were
35 sources of short-term, subchronic, or chronic health effects were evaluated for potential flaws in
36 their design, reporting, or conduct. As a result, one study, [Ma et al. \(2013b\)](#) (English translation

1 cited as [Ma et al. \(2013a\)](#)), was removed from consideration in the assessment because of
2 incomplete description of experimental methods that leads to uncertainty in the results. Another
3 study, [Eastman Kodak \(1978\)](#), a one-page data summary, was excluded because it does not provide
4 detailed data reporting.

5 The remaining studies are all sources of health effects data that may be used in the
6 assessment. The 20 studies summarized in the evidence tables are considered the “critical”
7 studies from which the study methods and results are presented in preliminary evidence tables
8 and exposure-response arrays (Section 3). There were also a few cases of the same study data
9 being contained in multiple reports; in those cases, the studies are listed together in the evidence
10 tables.

11 **2.3. STUDY CHARACTERISTICS THAT WILL BE CONSIDERED IN THE** 12 **FUTURE EVALUATION AND SYNTHESIS OF THE CRITICAL** 13 **EPIDEMIOLOGICAL STUDIES FOR DIBP**

14 Several considerations will be used in EPA’s evaluation of epidemiological studies of human
15 health effects of DIBP. These considerations include aspects of the study design affecting the
16 internal or external validity of the results (e.g., population characteristics and representativeness,
17 exposure and outcome measures, confounding, data analysis), focusing on specific types of bias
18 (e.g., selection bias; information bias due to exposure misclassification) and other considerations
19 that could otherwise influence or limit the interpretation of the data. A study is externally valid if
20 the study results for the study population can be extrapolated to external target populations. An
21 internally valid study is free from different types of biases, and is a prerequisite for generalizing
22 study results beyond the study population. These issues are outlined in the IRIS Preamble, and are
23 described below.

24 **2.3.1. Study Population**

25 Evaluation of study population characteristics (including key socio-demographic variables
26 and study inclusion criteria) can be used to evaluate external validity (i.e., generalizability) and to
27 facilitate comparison of results across different study populations. Some aspects of the selection
28 process may also affect the interval validity of a study, resulting in a biased effect estimate.

29 The general considerations for evaluating issues relating to the study population include
30 adequate documentation of participant recruitment, including eligibility criteria and participation
31 rates, missing data, and loss to follow-up. This information is used to evaluate internal study
32 validity related to selection bias. Different types of selection bias that may occur include the
33 healthy worker effect, differential loss to follow up, Berkson’s bias (relating to selection of
34 participants in hospital-based case-control studies), and participation bias. It is important to note
35 that low participation rates, or differences in participation rates between exposed and non-exposed
36 groups or between cases and controls, is not evidence of selection bias. Rather, selection bias arises
37 from a differential pattern of participation with respect to both the exposure and the outcome, i.e.,
38 patterns of participation that would result in a biased effect estimate. An example of differential

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1 participation would be when people with high levels of exposure and the outcome of interest are
2 more likely to participate than people with low levels of exposure and the outcome.

3 The available DIBP studies have generally examined metabolites from many different
4 phthalates within the context of research on environmental exposures. Most of these studies rely
5 on objective exposure measures (e.g., biomonitoring data), some of which are collected prior to
6 onset of the outcomes being examined (e.g., in the prospective pregnancy cohort studies). Study
7 participants generally do not have knowledge of the study hypothesis or their exposure to DIBP and
8 thus, knowledge of exposure or exposure level is unlikely to result in differential participation with
9 respect to outcomes. These study features should minimize the potential for selection bias.
10 However, EPA will consider the possibility that a particular concern about the specific sources of
11 DIBP, in conjunction with knowledge of specific health outcomes, may motivate people to
12 participate in a study or to continue participation throughout a follow-up period. In the absence of
13 evidence that any of these scenarios is likely to occur in a study, EPA will not consider selection bias
14 as a limitation of a study.

15 **2.3.2. Exposure Considerations**

16 General considerations for evaluating exposure include: (1) identifying how exposure can
17 occur (e.g., exposure sources, routes, and media); (2) determining appropriate critical exposure
18 period(s) for the outcomes under study; (3) evaluating variability in the exposure metrics of
19 interest (e.g., temporal and spatial variability for environmental measures or inter-individual
20 variability for biomonitoring data) that can impact different types of exposure metrics (e.g.,
21 cumulative, average, or peak exposure); (4) determining if an appropriate analytical methodology
22 was employed (e.g., choice of biological matrix, sampling protocol, quantification approach);
23 (5) evaluating the choice of exposure surrogate evaluated (e.g., constituent chemical or
24 group/mixture); and (6) evaluating the classification of individuals into exposure categories. These
25 six considerations help determine the accuracy and precision of the exposure estimates, and the
26 likelihood of measurement error with respect to the exposure metrics used. Nondifferential
27 misclassification of exposure categories, for example, can also result from measurement error and
28 is expected to predominantly result in attenuated effect estimates ([Blair et al., 2007](#)).

29 Some common sources of exposure to DIBP include cosmetics, food, and food packaging
30 ([Zota et al., 2014](#)) with the primary route of exposure occurring through ingestion and some
31 exposure occurring via inhalation and dermal routes (see Section 1.1.3). Thus, exposure to DIBP is
32 typically from multiple sources, and occurs episodically on a daily basis. Exposure to DIBP may be
33 increasing; a recent study of the U.S. general population found that urinary concentrations of the
34 DIBP metabolite MIBP have increased over time and were 206% higher in 2009–2010 compared to
35 2001–2002 ([Zota et al., 2014](#)).

36 Urine provides an integrated measure of phthalate exposure from all sources.
37 Measurement of DIBP metabolites, rather than the parent compound, is preferred because the
38 parent compound is metabolized very quickly and does not provide an accurate measure of
39 exposure. The simple monoester metabolite, monoisobutyl phthalate (MIBP) is the most commonly

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1 measured DIBP metabolite in epidemiologic studies. MIBP accounts for an estimated 70.3% of the
2 urinary excretion of DIBP; this value is based on human data from a controlled dosing study of a
3 single volunteer ([Koch et al., 2012](#)). EPA considers the use of MIBP to be a good proxy for total
4 DIBP exposure.

5 Although urine measures are most commonly used in epidemiological studies of phthalate
6 exposure, measures in serum, semen, and breast milk have also been used. Studies examining DIBP
7 metabolites in breast milk or serum have generally reported low levels of detection. One study in
8 Taiwan reported that MIBP above the limit of detection was found in 33.3% of breast milk samples
9 from 30 women. The detection rate in 30 cord blood samples in this study was 100%, but the
10 correlation between MIBP measured in cord blood and maternal urine was -0.11 (Pearson
11 correlation of log-transformed levels) ([Lin et al., 2011](#)). [Hogberg et al. \(2008\)](#) reported that few
12 breast milk (2 out of 42) or serum (3 out of 36) samples in a study in Sweden had detectable MIBP
13 concentrations. Another study conducted among 60 men ages 18–26 years found that 33.3% of
14 serum samples and 16.9% of seminal plasma samples had MIBP concentrations above the limit of
15 detection ([Frederiksen et al., 2010](#)). The Spearman correlation coefficient between urine and
16 serum concentrations was 0.39; the correlation between urine and seminal plasma concentrations
17 was not calculated because of the low detection rate for the latter samples ([Frederiksen et al.,](#)
18 [2010](#)). The lower detection rate in tissues other than urine reduces EPA’s confidence in DIBP
19 metabolite measures in these biological matrices.

20 Given their first-order kinetics with half-lives on the order of hours [3.9 hours for MIBP in
21 ([Koch and Angerer, 2007](#))], urinary phthalate metabolite concentrations peak shortly after
22 exposure. Thus, for single-time exposure scenarios (rather than multi-source, multiple time
23 exposure scenarios), urine sampled during this time of peak concentration could lead to
24 overestimates of average daily intake, and conversely, measurements made after concentrations
25 have peaked and declined could lead to underestimates of intake. One study conducted among
26 139 pregnant women in Puerto Rico included measurement of MIBP found that specific gravity
27 adjusted concentrations were lower in samples collected from 9 am to noon (geometric mean 9.4)
28 compared with samples collected in early morning, early afternoon, or evening (geometric means
29 13–14) ([Cantonwine et al., 2014](#)). Urinary measures of DIBP metabolite concentrations in
30 epidemiological studies are generally conducted using spot urine samples (i.e., collected at time of a
31 clinic or study examination visit) rather than at a specified time (e.g., first morning void) or in 24-
32 hour urine samples. Although the time of sample collection described above may affect the
33 accuracy of an estimated intake for a single individual, studies of other phthalates (e.g., DEHP) have
34 demonstrated that on a group level, spot urine samples provide a reasonable approximation of
35 concentrations that would have been observed using full-day urine samples ([Christensen et al.,](#)
36 [2012](#)) and that a single spot sample was reliable in ranking subjects according to tertile of MIBP
37 ([Teitelbaum et al., 2008](#)). Based on this information, EPA does not consider the reliance on spot
38 urine samples for exposure estimation (including ranking of individuals into different DIBP
39 categories) to be a major limitation for epidemiological studies. However because of the potential
40 for greater inaccuracy of estimates in the “tails” of the distribution, EPA will include additional

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1 considerations (e.g., discussion of analysis of residuals, outliers) when evaluating analyses based on
2 use of DIBP metabolites as continuous measures.

3 Another potential limitation of measurement of DIBP metabolites in urine is the
4 reproducibility of phthalate metabolite concentrations over time; that is, how well does a single
5 measure reflect the key exposure metric (average, peak) for the critical exposure window of
6 interest. For many short-lived chemicals, considerable temporal variability in exposure level is
7 expected, and thus, repeated measures in the critical exposure window are preferred over a single
8 measurement. Reproducibility is usually evaluated with the intraclass correlation coefficient (ICC),
9 a measure of the ‘between-individual’ variance divided by the total variance (between and within
10 individuals). A higher ICC indicates greater reproducibility (i.e., lower within-person variance). An
11 ICC of 0.51 for MIBP was reported in a study of 25 Hmong women ages 19–51 years with samples
12 collected 2-4 weeks apart ([Peck et al., 2010](#)). In studies of reproducibility of measures during
13 pregnancy, [Cantonwine et al. \(2014\)](#) reported ICCs of 0.35 and 0.34 (unadjusted and specific gravity
14 adjusted) when comparing urine samples taken at approximately 18, 22, and 26 weeks of gestation.
15 ICCs of 0.36 and 0.38, respectively, were seen before pregnancy and in early pregnancy ([Braun et
16 al., 2012](#)), and an ICC of approximately 0.5 was seen over a 6-week period in the last trimester
17 ([Adibi et al., 2008](#)). Among women participating in the Nurses’ Health Study (NHS) (in 2000–2001
18 for NHS and in 1996-1999 for NHS II), the ICC for samples collected 1-3 years apart was 0.30 for all
19 samples, and was 0.29 for first-morning samples ([Townsend et al., 2013](#)). Data for children are
20 sparse, limiting the ability to examine this source of uncertainty in this population. One study
21 evaluated variability in children aged 6–10 years old over a 6-month period ([Teitelbaum et al.,
22 2008](#)) and found a relatively low ICC (0.21 unadjusted, 0.28 creatinine-adjusted). The available
23 data highlight the value of repeated exposure measures collected during the appropriate critical
24 period for the outcome(s) under study. Based on these studies, however, EPA does not consider the
25 use of a single measurement to be a major limitation in studies in adults in which the measure of
26 exposure is closely aligned with the relevant window(s) of exposure, if known, for the effect under
27 study. EPA has greater uncertainty, however, about measurements taken outside of the relevant
28 time window (e.g., several years after diagnosis, or the difference between first and third trimesters
29 of pregnancy), and about measurements taken in children.

30 Some studies present analyses using a combined measure based on summation of MIBP and
31 monobutyl phthalate (MBP), as a measure of both DIBP and DBP, respectively. The relative
32 contribution of DIBP to this total has varied over time (as the use of DIBP has increased), and can
33 vary between populations (e.g., greater use of DIBP compared with DBP in some countries). EPA
34 includes studies in the DIBP evidence tables using this summed exposure measure in situations in
35 which the concentration of MIBP is greater than that of MBP, but recognizes that this measure
36 introduces an additional source of exposure misclassification. Other studies present analyses using
37 a combined “low molecular weight” phthalate measure based on the summation of MIBP, MBP, and
38 monoethyl phthalate (MEP) (reflecting exposure to the parent compounds of DIBP, DBP, and DEP,
39 respectively). Because MIBP does not represent a major contributor to this summation
40 measurement, EPA has not included data from these studies in the DIBP evidence tables.

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1 EPA will also consider the potential for differential misclassification of biomarker measures
2 of exposure; for example, in situations in which a health outcome (e.g., diagnosis with diabetes or
3 cancer) could lead to a behavioral change that results in a change in DIBP exposure. This type of
4 scenario adds an additional challenge to the interpretation of the DIBP metabolites as valid
5 measures of exposure in a relevant time window(s) with respect to disease development.

6 The distribution of exposure will also be considered in evaluating individual studies and
7 when comparing results among groups of studies. One consideration is the contrast of exposure
8 levels (i.e., the difference between “high” and “low”): a study with a very narrow contrast may not
9 have sufficient variability to detect an effect that would be seen over a broader range. Another
10 consideration is the absolute level of exposure, as different effect estimates may be expected in
11 studies examining different exposure levels even if they had similar exposure contrasts.

12 **2.3.3. Primary Outcome Measures**

13 The general considerations for evaluating issues relating to accuracy, reliability, and
14 biological relevance of outcomes include adequate length of follow-up to evaluate the outcomes of
15 interest, and use of appropriate ascertainment methods to classify individuals with regard to the
16 outcome (e.g., high sensitivity and specificity). With respect to continuous measures, such as
17 hormone concentrations or semen parameters, EPA will consider, in addition to assessing whether
18 reported parameters are outside normal physiological range, evidence of smaller changes in the
19 distribution of a parameter that may represent an effect on a population level [e.g., as is the case for
20 early childhood exposure to lead and decrements in intelligence as measured by IQ ([U.S. EPA,
21 2013](#))].

22 Issues relating to assessment of the specific primary health effects are discussed below and
23 summarized in Table 2-8 at the end of Section 2.3.

24 ***Sexual Differentiation***

25 Cryptorchidism and hypospadias are two disorders of the development of the male
26 reproductive system. Cryptorchidism, or undescended testes, can be present at birth (congenital
27 cryptorchidism) or can occur later during infancy and childhood (acquired cryptorchidism).
28 Surgical correction (orchiopexy) is recommended in cases of cryptorchidism that do not resolve
29 during infancy because long-term complications include impaired sperm production and increased
30 risk of testicular cancer ([Virtanen et al., 2007](#)). Retractable testes can move back and forth between
31 the scrotum and the abdomen; this condition usually resolves by puberty and is not associated with
32 reproductive or other complications. Classification criteria for cryptorchidism that involve
33 testicular positioning are commonly used in clinical research ([John Radcliffe Hospital
34 Cryptorchidism Study Group, 1988](#); [Scorer, 1964](#)). EPA will consider the definition used and age
35 range in interpreting studies of cryptorchidism or related outcomes.

36 In animal toxicology studies, anogenital distance (AGD) is a routine marker to assess
37 endocrine disruption; this marker has only recently been adapted for use in epidemiological
38 studies. One study in adult men reported associations between decreased AGD and measures

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1 relating to infertility ([Eisenberg et al., 2011](#)); most studies have used this measure in infants,
2 however, as a marker of endocrine environment during development. It is important to consider
3 general size, in addition to sex, in the evaluation of AGD, for example by incorporating birth weight
4 or length (e.g., calculation of “anogenital index” by dividing anogenital distance by weight). With
5 regard to reproducibility of this measure, a low degree of between-observer variability was found
6 using a standardized protocol and trained observers ([Romano-Riquera et al., 2007](#); [Salazar-
7 Martinez et al., 2004](#)). Because of the importance of size and age in the interpretation of this
8 measure, EPA has greater confidence in studies with measures taken at birth or over a narrow age
9 range and lesser confidence in studies among a group spanning a larger age range.

10 Gender-related behaviors, as measured by the Pre-School Activities Inventory ([Golombok
11 and Rust, 1993](#)) or other scales, has been examined in relation to direct or indirect measures of
12 fetal testosterone levels, including studies of DIBP. This outcome measure has been examined in
13 studies of relatively rare genetic conditions (e.g., congenital adrenal hyperplasia and complete
14 androgen insensitivity syndrome), as well as in studies focusing on the normal variability seen in
15 the general population ([reviewed in Hines, 2006](#)). EPA will consider evidence pertaining to the
16 reliability and validity of the Pre-School Activities Inventory in its evaluation of studies using this
17 scale.

Male and Female Reproductive Outcomes

19 The DIBP literature includes studies of reproductive and gonadotropin hormone levels in
20 men and studies of semen parameters that can be indicative of reduced fertility. The details of the
21 laboratory procedures, including information on the basic methods, level of detection, and
22 coefficient of variation, are important considerations for hormone assays and measures of semen
23 parameters. The World Health Organization (WHO) laboratory methods for analysis of sperm
24 counts and semen parameters ([see, for example, WHO, 1999](#)) are generally recognized as standards
25 in this field. EPA will consider studies that reference these methods, regardless of which revision
26 used, to be reliable measures.

27 Much of the focus of the research on male steroidal and gonadotropin hormones in the DIBP
28 database concerns testosterone. One issue with respect to these measures is the estimation method
29 used for free testosterone. Based on the analysis by [Vermeulen et al. \(1999\)](#), EPA will consider
30 estimates based on total testosterone divided by immunoassay-derived sex-hormone binding
31 globulin (SHBG) levels to be most reliable.

32 The DIBP literature also includes studies of reproductive hormones in women. In addition
33 to the general considerations regarding hormone assays noted above, timing within a menstrual
34 cycle for studies of pre- and peri-menopausal women, and timing with respect to gestational age for
35 studies of women during pregnancy, are also be an important considerations for interpretation of
36 reproductive hormone concentrations.

37 Another female reproductive outcome included in the DIBP literature is endometriosis.
38 Endometriosis can be symptomless, or can lead to surgical intervention; it is often diagnosed as
39 part of a work-up for infertility. Variability in clinical presentation and in access and use of health

1 care services present considerable challenges to conducting epidemiological studies of this
2 condition ([Holt and Weiss, 2000](#)). Confirmation of “case” and “control” status (i.e., presence or
3 absence of endometriosis) by ultrasound or clinical evaluation is recommended to reduce outcome
4 misclassification, and representation of the source population should be carefully considered.

5 Infertility is generally defined clinically and for research purposes as the inability to
6 conceive a clinically-recognized pregnancy after 12 months of intercourse of regular frequency
7 without use of contraceptives. Fecundity or fecundability are terms for the capacity for
8 reproduction. “Time to pregnancy” (i.e., the number of cycles of unprotected intercourse before
9 conception) has been used as a measure of fecundability in studies of environmental and
10 occupational exposures ([Baird et al., 1986](#); [Baird and Wilcox, 1985](#)). Time to pregnancy is a
11 measure of a couple’s fecundability, incorporating effects that can be manifested through the male
12 or female (or both). Considerations in time to pregnancy studies include the source of data (i.e.,
13 retrospective or prospective designs) and incorporation of information on “non-pregnancy
14 planners” ([Weinberg et al., 1994](#)).

15 ***Timing of Male and Female Puberty, and Conditions of Unusual Pubertal Development***

16 Pubertal development in humans is often assessed using timing of peak height velocity
17 (“growth spurt”) and secondary markers of sexual development. Secondary markers for females
18 include breast development (thelarche) and pubic hair development (pubarche), and age at first
19 period (menarche). Secondary markers for males include gonadal development (gonadarche) and
20 pubic hair development, and age at first sperm emission (spermarche).

21 Evaluation of breast, pubic hair, and gonadal development is frequently performed using
22 the Tanner stages ([Marshall and Tanner, 1970, 1969](#)), which places the individual in one of five
23 stages, ranging from pre-pubertal (stage 1) to adult maturation (stage 5). However, the process of
24 this staging is not straightforward, and is most reliable when performed by trained personnel
25 (rather than by the individual or a parent, for example) ([Slough et al., 2013](#); [Schlossberger et al.,
26 1992](#); [Espeland et al., 1990](#)). Age at menarche is considered to more reliable when assessed via
27 self-report ([Koprowski et al., 2001](#)), although reliability may decrease with increasing time since
28 menarche ([Cooper et al., 2006](#)). Additionally, hormone levels may sometimes be used to evaluate
29 pubertal development. Individuals may vary widely in the timing of these developmental
30 milestones.

31 Several clinical syndromes are known to disrupt the timing and order of markers of
32 pubertal development. Considerations in the diagnosis of either precocious or delayed puberty
33 include the diagnostic criteria used and the source of the information (e.g., whether collected from
34 medical records or from self- or parental report). For females, precocious puberty is usually
35 defined as the onset of puberty before the age of 8 years, while delayed puberty is usually defined
36 as the lack of pubertal development by the age of 13 years ([Marshall and Tanner, 1969](#));
37 corresponding ages in males are before the age of 9 years for precocious puberty and lack of
38 pubertal development by the age of 14 years for delayed puberty ([Marshall and Tanner, 1970](#)).
39 Clinical evaluation would involve hormone assays to distinguish between gonadotropin dependent

1 (“central”), gonadotropin independent (“peripheral”), or a combination of both ([Traggiai and](#)
2 [Stanhope, 2003](#)) forms of these conditions.

3 ***Pregnancy-Related Outcomes***

4 Infant birth weight and gestational age are two outcomes commonly used in reproductive
5 epidemiology studies. EPA considers analyses of the various indices for both outcomes (fetal
6 growth and gestational age) to be informative with respect to hazard identification, but will
7 consider each separately as they address different issues. Gestational duration can be measured as
8 a continuous outcome or dichotomous outcome such as preterm birth. Preterm births include
9 infants delivered earlier than 37 gestational weeks, and those delivered earlier than 32 gestational
10 weeks are classified as very preterm births. Different measures of fetal growth restriction are often
11 examined in epidemiological studies. In addition to the continuous measure of birth weight,
12 another commonly used measure of fetal growth restriction is the categorical variable of low birth
13 weight (defined as <2,500 g). Small for gestational age (defined as birth weight less than the 10th
14 percentile for the gestational birth weight distribution) is considered a better measure of fetal
15 growth rate as it takes into consideration gestational duration, and would be preferred over a
16 measure of birth weight in a study that includes preterm births. Birth weight and gestational
17 duration can also be examined as continuous variables, often in analysis that excludes preterm or
18 low birth weight births, so that the focus of the analysis is on variability within the “normal” range.

19 EPA considers birth weight obtained from medical records to be a reliable source as this is a
20 very accurate and precise measurement. Although more prone to measurement error than birth
21 weight measures, gestational age can be estimated from several approaches. Some of these include
22 ultrasonography, estimates based on date of last menstrual period based on maternal recall, or
23 from clinical examination based on antenatal or newborn assessments (which may include an
24 ultrasound). Menstrual dating of gestational age dependent on maternal recall of the last menstrual
25 period can be subject to considerable measurement error in some cases, so ultrasonography-based
26 estimates may be considered more accurate ([Savitz et al., 2002](#); [Taipale and Hiilesmaa, 2001](#)).

27 ***Immune-Related Outcomes: Allergy and Asthma***

28 Skin prick testing is a standard method for assessing atopy (allergic disease) used in some
29 epidemiologic studies. Other studies use an assessment protocol based on reported history of
30 symptoms (e.g., rhinitis, hay fever) or specific types of allergies. These can be considered
31 complementary types of measures: skin prick tests provide information on a defined set of
32 potential antigens to which a person may be exposed, and symptom-based evaluations provide
33 information on experiences of individuals and the variety of exposures they encounter. Studies
34 comparing questionnaire responses with skin prick tests in children have reported relatively high
35 specificity (89–96%) and positive predictive value (69–77%) for self-reported history of pollen or
36 pet dander allergy or for answers to a combination of questions incorporating itchy eyes with nasal
37 congestion in the absence of a cold or flu ([Braun-Fahrlander et al., 1997](#); [Dotterud et al., 1995](#)). The
38 validity was somewhat lower for a more restricted set of questions (nasal congestion in the absence

1 of a cold or flu; specificity 83%, positive predictive value 52%) ([Braun-Fahrländer et al., 1997](#)).
2 Based on these data, EPA considers allergy history based only on rhinitis symptoms to have a
3 greater likelihood of outcome misclassification compared with those based on a combination of
4 symptoms.

5 Epidemiologic studies of asthma typically use a questionnaire-based approach to define
6 asthma based on symptoms relating to wheezing episodes or shortness of breath, reported history
7 of asthma attacks, or use of asthma medication, usually for a period defined as “current” or in the
8 past year. Much of this work is based upon the American Thoracic Society questionnaire ([Ferris,
9 1978](#)) or subsequent instruments that built upon this work, including the International Society of
10 Arthritis and Allergies in Children Questionnaire and the European Community Respiratory Health
11 Survey. These questionnaire-based approaches have been found to have an adequate level of
12 specificity and positive predictive value for use in etiologic research ([Ravault and Kauffmann, 2001](#);
13 [Pekkanen and Pearce, 1999](#); [Burney et al., 1989](#); [Burney and Chinn, 1987](#)). EPA considers
14 outcomes defined over a recent time period (e.g., symptoms in the past 12 months) to be more
15 relevant within the context of concurrent exposure measurements compared with outcomes
16 defined over a lifetime (e.g., ever had asthma).

17 **Neurodevelopment**

18 With respect to neurodevelopmental outcomes, a major consideration is the assessment
19 tool(s) used by the study investigators; details of the assessment method, or references providing
20 this information, should be provided. In addition, EPA also looks for discussion of (or reference to)
21 validation studies and the appropriateness of the tool for evaluation in the specific study population
22 (e.g., age range, language).

23 **Thyroid**

24 Thyroid-related endpoints examined in epidemiological studies of DIBP include thyroid
25 hormones (triiodothyronine, T3, and thyroxine, T4) and thyroid stimulating hormone (TSH) (or
26 thyrotropin) produced by the pituitary.

27 As with other hormone assays, the details of the laboratory procedures, including
28 information on the basic methods, limit of detection, and coefficient of variation, are important
29 considerations for the hormone assays. Thyroid hormones are generally measured in serum,
30 although they may also be measured in dried blood spots, such as are collected from newborn
31 infants in screening for congenital hypothyroidism. Studies in older age groups have also shown a
32 very high correlation ($r = 0.99$) between thyroid hormone levels measured in dried blood spots and
33 levels in serum ([Hofman et al., 2003](#)).

34 With respect to thyroid hormones, time of day and season of sampling are two main
35 potential sources of variability. For example, serum TSH measured shortly after midnight may be
36 as much as twice as high as the value measured in late afternoon ([Brabant et al., 1991](#); [Weeke and
37 Gundersen, 1978](#)). The evidence with respect to seasonal variability is mixed ([Plasqui et al., 2003](#);
38 [Nicolau et al., 1992](#); [Simoni et al., 1990](#); [Behall et al., 1984](#); [Postmes et al., 1974](#)) and this effect is

1 likely to be smaller than that of time of day. The impact of these sources of variation will depend on
2 whether they are also related to DIBP (i.e., whether DIBP levels vary diurnally or seasonally). If this
3 is the case, failure to address these factors in the design or analysis could result in confounding of
4 the observed association, with the direction of this bias determined by the direction of the
5 association between these factors and DIBP. If this is not the case, the lack of consideration of time
6 of day or seasonality would result in greater variability in the hormone measures, and would thus
7 result in more imprecise (but not biased) estimates was located. EPA has not found studies
8 examining seasonal variation in DIBP levels. With respect to variability relating to time of day, as
9 noted previously, one study of 139 pregnant women in Puerto Rico reported lower concentrations
10 of specific gravity-adjusted MIBP in samples collected from 9 am to noon (geometric mean of 9.4)
11 compared with samples collected in early morning, early afternoon, or evening (geometric means of
12 13–14) ([Cantonwine et al., 2014](#)). Based on these data, EPA has greater confidence in thyroid
13 hormone studies that consider time of sample collection in the analysis, but recognizes the limited
14 nature of the available data pertaining to this issue.

15 ***Obesity***

16 Most of the studies of obesity measures in the DIBP database are based on body mass index
17 (BMI, calculated as kg/m²) or waist circumference using measurements taken as part of the data
18 collection protocol. BMI is highly correlated with body fat, and standardized cut-points have been
19 established for characterization of “normal” (BMI between 18.5 and 24.9 kg/m²), “overweight”
20 (BMI between 25.0 and 29.9 kg/m²) and “obese” (BMI ≥ 30.0 kg/m²) categories. Waist
21 circumference is also highly correlated with body fat, and is a more direct measure of abdominal
22 obesity. EPA notes that use of self-reported weight (e.g., report of pre-pregnancy weight) would
23 not be considered to be as reliable as actual measurements.

24 ***Diabetes and Measure of Insulin Resistance***

25 In the DIBP database, diabetes has been assessed by a variety of biomarkers of glucose and
26 insulin and by self-report of diabetes diagnosis. Oral glucose tolerance testing and glycosolated
27 hemoglobin (HbA1c) are used clinically and in epidemiological research ([Selvin et al., 2011](#)). Self-
28 report of prevalent diabetes can have high sensitivity and specificity in comparison to diagnosed
29 diabetes based on validated medical record data ([Oksanen et al., 2010](#); [Leikauf and Federman,](#)
30 [2009](#)). The biomarker-based classifications, however, offer an added advantage of being able to
31 include undiagnosed disease. EPA will consider these points in assessing the reliability and validity
32 of the diabetes measures used in the studies. None of the currently available studies assessed
33 diabetes through cause of death data; sensitivity of diabetes assessed using cause of death data is
34 low, even if underlying and other contributing cause of death fields are included ([Cheng et al.,](#)
35 [2008](#)).

36 Insulin resistance, a marker of diabetes risk, can be measured using the homeostatic model
37 assessment (HOMA) method, a physiologically-based structural model, using fasting glucose and
38 insulin or C-peptide concentrations. HOMA is a validated tool for the estimation of insulin

1 resistance in epidemiology studies, and requires a single measurement of fasting glucose and
2 insulin ([Wallace et al., 2004](#)). Although the mean of three samples taken at 5-minute intervals
3 results in a more precise estimate, insulin resistance estimated using a single baseline
4 measurement is well correlated with that using the mean of three measurements when used to
5 estimate a group mean. Therefore, EPA does not consider the use of a single measurement as an
6 input to the HOMA model to be a limitation.

7 **Cancer**

8 With respect to studies of cancer, EPA considers the source of the outcome data (e.g., cause
9 of death data, hospital cancer registry data, hospital discharge data, histopathology reports) in its
10 evaluation of the accuracy of the data. An additional issue is the validity of mortality data as a
11 representation of cancer incidence; mortality data for cancer types with a high survival rate may
12 underrepresent disease incidence, require additional considerations with respect to determining
13 appropriate time windows of exposure, and may lead to biased risk estimates if survival is related
14 to exposure.

15 **2.3.4. Confounding**

16 The general considerations for evaluating issues relating to potential confounding include
17 consideration of which factors may be potential confounders (i.e., those which are strongly related
18 to both the exposure and the outcome under consideration, and are not intermediaries on a causal
19 pathway), adequate control for these potential confounders in the study design or analysis, and
20 where appropriate, quantification of the potential impact of mismeasured or unmeasured
21 confounders. Uncontrolled confounding by factors that are positively associated with both the
22 exposure (e.g., DIBP) and health endpoint of interest, and those that are inversely associated with
23 both exposure and health endpoint, will result in an upward bias of the effect estimate.
24 Confounding by factors that are positively associated with exposure and inversely associated with
25 the health endpoint (or vice versa) will result in a downward bias of the effect estimate.

26 **Potential Confounding by Other Phthalates**

27 Few studies have reported results of analyses evaluating the correlation between MIBP and
28 metabolites of other phthalates. In an analysis conducted by EPA of 5,109 samples from the
29 2003–2008 National Health and Nutrition Examination Survey (NHANES) participants aged ≥6
30 years, the pairwise Spearman correlation coefficient between MIBP and MEP (the primary
31 metabolite of DEP) was low (0.33). A more moderate correlation was seen with the DEHP
32 metabolites (correlations of approximately 0.5); higher correlations were seen with MBzP (the
33 primary metabolite of BBP, correlation coefficient = 0.58) and MBP (the primary metabolite of DBP;
34 correlation = 0.72). Similar or somewhat lower correlations were seen between MIBP and other
35 phthalate metabolites in a small study (n = 45) of men seen in an infertility clinic ([Wirth et al.,
36 2008](#)), in 319 pregnancy women ([Whyatt et al., 2012](#)), and in 600 reproductive age women in a
37 study of endometriosis ([Buck Louis et al., 2013](#)). EPA will evaluate the potential for confounding by

1 examining the similarity of the results seen with different metabolites. Thus, for example, lack of
2 adjustment for mono-benzyl phthalate (MBzP) would not be considered a limitation in a study in
3 which an association was seen with MIBP that was not seen with MBzP; however this lack of
4 adjustment would be considered a limitation if an association of similar or higher magnitude was
5 seen for both of metabolites.

6 ***Potential Confounding by Demographic Factors***

7 Age, race/ethnicity, and sex are considered important explanatory factors for most types of
8 outcomes measured in epidemiological research. In NHANES 2009–2010 data, urinary MIBP levels
9 decreased with age (geometric means of 13.2, 8.63, and 7.45 µg/g-creatinine, respectively, in ages
10 6–11, 12–19 and ≥20 years) ([CDC, 2013](#)). Concentrations were lower levels in males compared
11 with females (geometric means of 6.99 and 9.05 µg/g-creatinine, respectively, in males and
12 females), and variability by ethnicity was also observed, with lower levels in non-Hispanic whites
13 (geometric mean of 7.12 µg/g-creatinine) compared with non-Hispanic blacks and Mexican
14 Americans (geometric means of 10.1 and 9.27 µg/g-creatinine, respectively). EPA will consider
15 these differences in assessing the potential influence of demographic factors on observed effect
16 estimates for DIBP.

17 ***Potential Confounding by Other Factors***

18 Some of the health effects under consideration may have strong associations with other risk
19 factors. For example, smoking is associated with increased risk of low birth weight and preterm
20 births, and with infertility. Abstinence time is strongly related to sperm concentration measures.
21 In evaluating the potential for confounding by any of these factors, EPA will review evidence
22 pertaining to the strength and direction of its association with DIBP (or its metabolites).

23 **2.3.5. Data Analysis**

24 The general considerations for evaluating issues relating to data analysis include adequate
25 documentation of statistical assumptions and analytic approach (including addressing skewness of
26 exposure or outcome variable and shape of exposure-response), consideration of sample size and
27 statistical power, and use of appropriate statistical methods for the study design.

28 One other issue, specific to much of the DIBP literature, concerns the optimal approach to
29 addressing urinary volume or dilution in the analysis of spot urine or first morning void samples.
30 Options include use of creatinine- or specific gravity-adjusted metabolite concentrations, or use of
31 unadjusted concentrations. Although use of some kind of correction factor has been advocated for
32 studies of obesity ([Goodman et al., 2014](#)), a simulation study reported that creatinine-adjusted
33 exposure measures may produce biased effect estimates for outcomes that are strongly related to
34 factors affecting creatinine levels, of which obesity is a prime example ([Christensen et al., 2014a](#)).
35 EPA recognizes the lack of consensus at this time, as well as the need for continued research into
36 the potential bias introduced by different analytic approaches. Based on current understanding of
37 this issue, EPA prefers results using unadjusted concentration for outcomes strongly related to

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1 creatinine levels; for other outcomes, EPA does not have a basis for preferring one type of analysis
 2 over another.

3 **Table 2-8. General and outcome-specific considerations for DIBP study**
 4 **evaluation**

General considerations	
Study population	<ul style="list-style-type: none"> • Study population and setting: geographic area, site, time period, age and sex distribution, other details as needed (may include race/ethnicity, socioeconomic status) • Recruitment process; exclusion and inclusion criteria, knowledge of study hypothesis; knowledge of exposure and outcome • Participation rates: total eligible; participation at each stage and for final analysis group and denominators used to make these calculations • Length of follow-up, loss to follow-up • Comparability: participant characteristic data by group, data on non-participants
Exposure	<ul style="list-style-type: none"> • Biological matrix or target tissue/organ (e.g., urine, serum, semen, breast milk) • Level of detection (LOD) or level of quantitation (LOQ) • Exposure distribution (e.g., central tendency, interquartile range), proportion < LOD
Analysis	<ul style="list-style-type: none"> • Consideration of data distribution including skewness of exposure and outcome measures • Consideration of influence of “tails” in analysis based on continuous exposure measure • Consideration of analytic approaches exploring different shapes of exposure-response • Consideration of values below LOD or LOQ • Consideration of creatinine or other approach to adjust for urine volume. • Presentation of effect estimates, rather than statement regarding presence or absence of statistical significance
Outcome-specific considerations	
<i>Sexual differentiation</i>	<ul style="list-style-type: none"> • AGD: protocol, training procedures, standardization and inter-rater reliability • Cryptorchidism: definition • Gender related play behavior: reliability and validity of measurement scale
Measures	
Consideration of confounding	<ul style="list-style-type: none"> • AGD: variability by size (e.g., birth weight), sex, age; temporal trends in DIBP exposure if study spans several years and includes a wide age range • Cryptorchidism, preterm birth
Relevant exposure time window(s)	<ul style="list-style-type: none"> • In utero for outcomes assessed in infancy; for acquired cryptorchidism, other time window(s) during childhood may also be relevant

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<i>Steroid and gonadotropin hormones (adults; sex-specific)</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Type of assay Sensitivity/detection limits, coefficient of variation; number of samples below LOD
	<ul style="list-style-type: none"> Age, day or phase of menstrual cycle (if cycling)
	<ul style="list-style-type: none"> Up to 6 months preceding hormone sample collection
<i>Sperm parameters</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Type of assay (e.g., WHO protocol)
	<ul style="list-style-type: none"> Age, smoking, BMI, abstinence time (consider if these are related to exposure)
	<ul style="list-style-type: none"> Up to 6 months preceding semen sample collection
<i>Infertility</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Definition, source of data
	<ul style="list-style-type: none"> Age, smoking, alcohol use, heavy metal exposure, radiation time (consider if these are related to exposure)
	<ul style="list-style-type: none"> Time preceding and during attempt to become pregnant
<i>Timing of puberty</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Source of data (e.g., self-report, physician assessment)
	<ul style="list-style-type: none"> Age, sex, ethnicity, body size, nutritional status (consider if these are related to exposure)
	<ul style="list-style-type: none"> In utero? Up to 12 months preceding transition from one stage to another stage?
<i>Gestational age</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Source of data and estimation procedure (ultrasound; last menstrual period or clinical assessment)
	<ul style="list-style-type: none"> Smoking, pregnancy complications, assisted reproduction technologies (consider if these are related to exposure)
	<ul style="list-style-type: none"> In utero
<i>Birth weight</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Source of data (e.g., medical records, birth certificate)
	<ul style="list-style-type: none"> Gestational age, maternal age, ethnicity, nutritional intake, smoking, maternal height/BMI, (consider if these are related to exposure)
	<ul style="list-style-type: none"> In utero
<i>Immune – allergy and asthma</i> Measures Consideration of confounding Relevant exposure time window(s)	<ul style="list-style-type: none"> Number of allergens used in skin prick testing or allergen-specific IgE assay; sensitivity/specificity of specific questions used in history assessment
	<ul style="list-style-type: none"> Age, family history (consider if these are related to exposure)
	<ul style="list-style-type: none"> For current conditions (e.g., asthma in past 12 months): up to 12 months

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	preceding outcome assessment
<i>Neurobehavioral</i>	
Measures	<ul style="list-style-type: none"> Standardized assessment tool, validation studies for specific study population (e.g., age group, geographic location) Blinding of assessor to exposure
Consideration of confounding	<ul style="list-style-type: none"> Age, sex, socioeconomic status
Relevant exposure time window(s)	<ul style="list-style-type: none"> In utero; early childhood
<i>Thyroid</i>	
Measures	<ul style="list-style-type: none"> Assay used and evidence from validation studies, if available Sensitivity/detection limits, coefficient of variation; number of samples below LOD Time of day and season when samples for thyroid hormone (and TSH) collected
Consideration of confounding	<ul style="list-style-type: none"> Age, sex, smoking, iodine, radiation exposure (consider if these are related to exposure)
Relevant exposure time window(s)	<ul style="list-style-type: none"> Varies by lifestage (i.e., infants, children, adults)
<i>Obesity</i>	
Measures	<ul style="list-style-type: none"> Source of data (e.g., measured or self-reported weight and height)
Consideration of confounding	<ul style="list-style-type: none"> Age, sex, ethnicity, caloric intake, physical activity (consider if these are related to exposure)
Relevant exposure time window(s)	<ul style="list-style-type: none"> Not established (likely to be more than one, including in utero)
<i>Diabetes and insulin resistance</i>	
Measures	<ul style="list-style-type: none"> Source of data (e.g., biomarkers of insulin or glucose, medical records, self-report)
Consideration of confounding	<ul style="list-style-type: none"> Age, sex, ethnicity
Relevant exposure time window(s)	<ul style="list-style-type: none"> Not established (likely to be more than one, including in utero)

1

2 **2.4. STUDY CHARACTERISTICS THAT WILL BE CONSIDERED IN THE**
3 **FUTURE EVALUATION AND SYNTHESIS OF THE CRITICAL**
4 **EXPERIMENTAL STUDIES FOR DIBP**

5 Beyond the initial methodological screening described above in Section 2.2.2,
6 methodological aspects of a study’s design, conduct, and reporting will be considered again in the
7 overall evaluation and synthesis of the pertinent data that will be developed for each health effect.
8 Some general questions that will be considered in evaluating experimental animal studies are
9 presented in Table 2-9. These questions are, for the most part, broadly applicable to all
10 experimental studies.

1 **Table 2-9. Questions and relevant experimental information for the**
2 **evaluation of experimental animal studies**

Methodological feature	Question(s) considered
Test animal	Based on the endpoint(s) in question, are concerns raised regarding the suitability of the species, strain, or sex of the test animals on study?
Experimental setup	Are the timing, frequency and duration of exposure, as well as animal age and experimental group allocation procedures/group size for each endpoint evaluation, appropriate for the assessed endpoint(s)?
Exposure	Are the exposure conditions and controls informative and reliable for the endpoint(s) in question, and are they sufficiently specific to the compound of interest?
Endpoint evaluation procedures	Do the procedures used to evaluate the endpoint(s) in question conform to established protocols, or are they biologically sound? Are they sensitive for examination of the outcome(s) of interest?
Outcomes, data, and reporting	Were data reported for all pre-specified endpoint(s) and study groups, or were any data excluded from presentation/analyses?

3
4 Note: “Outcome” refers to findings from an evaluation (e.g., steatosis), whereas “endpoint” refers to the
5 evaluation itself (e.g., liver histopathology).
6

7 Evaluation of some specific methodological features identified in Table 2-9 such as
8 exposure, is likely to be relatively independent of outcome. Other methodological features, in
9 particular those related to experimental setup and endpoint evaluation procedures, are generally
10 outcome specific (i.e., reproductive and developmental toxicity). In general, experimental animal
11 studies will be compared against traditional assay formats (e.g., those used in guideline studies),
12 with deviations from the protocol evaluated in light of how the deviations could alter interpretation
13 of the outcome in question. A full evaluation of all critical studies will be performed as part of the
14 critical review and synthesis of evidence for hazard identification for each of the health endpoints
15 identified in the evidence tables presented in Section 3.
16

3. PRELIMINARY EVIDENCE TABLES AND EXPOSURE-RESPONSE ARRAYS

3.1. DATA EXTRACTION FOR EPIDEMIOLOGICAL AND EXPERIMENTAL STUDIES: PREPARATION OF PRELIMINARY EVIDENCE TABLES

The evidence tables present data from studies related to a specific outcome or endpoint of toxicity. At a minimum, the evidence tables include the relevant information for comparing key study characteristics such as study design, exposure metrics, and dose-response information. Evidence tables will serve as an additional method for presenting and evaluating the suitability of the data to inform hazard identification for DIBP during the analysis of hazard potential and utility of the data for dose-response evaluation. For each critical study selected, key information on the study design, including characteristics that inform study quality, and study results pertinent to evaluating the health effects from subchronic and chronic oral exposure to DIBP are summarized in preliminary evidence tables.

Epidemiological studies are presented first where each study per table is listed in reverse chronological order. Animal studies are then presented where each study per health endpoint is presented in alphabetical order by study author, followed by species and strain. Most results are presented as the percent change from the control group; an asterisk (*) indicates a result that has been calculated and reported by study authors to be statistically significant compared to controls ($p < 0.05$). Unless otherwise noted in a footnote, doses presented in the animal evidence tables were those reported by the study authors.

The information in the preliminary evidence tables is also displayed graphically in preliminary exposure-response arrays. In these arrays, a significant effect (indicated by a filled circle) is based on statistical significance by the study authors. The complete list of references considered in preparation of these materials can be found on the Health and Environmental Research Online (HERO) website at (<https://hero.epa.gov/DIBP> and <http://hero.epa.gov/phthalates-humanstudies>).

1 **3.2. EPIDEMIOLOGICAL STUDIES**

2 **3.2.1. Sexual Differentiation Measures**

3 **Table 3-1. Evidence pertaining to DIBP and sexual differentiation effects in**
 4 **humans**

Reference and study design	Results								
<i>Anogenital distance (AGD)</i>									
<p>Swan (2008) (United States; Minnesota, Missouri, California) Population: 106 boys from birth cohort study (Study for Future Families), 2000–2002, mean age 12.8 mo (0–36 mo) Outcome: AGD (to posterior genitalia) measured at 0–36 mo (mean 70.4 mm, 7.1 mm/kg) Exposure: Maternal urine sample, 3rd trimester MIBP in urine (ng/mL):</p> <table border="0" data-bbox="190 808 782 871"> <tr> <td></td> <td>Median</td> <td>75th percentile</td> </tr> <tr> <td>Unadjusted</td> <td>2.5</td> <td>5.1</td> </tr> </table> <p>Analysis: Regression analysis using mixed model adjusting for age and weight percentile Related references: Swan et al. (2005) (exposure data and analysis of smaller sample size with less robust method of adjustment for variation by size)</p>		Median	75 th percentile	Unadjusted	2.5	5.1	<p>Percent change in AGD per interquartile increase in MIBP concentration (<i>p</i>-value)</p> <table border="0" data-bbox="782 598 1427 640"> <tr> <td>MIBP</td> <td>-3.5 (0.097)</td> </tr> </table>	MIBP	-3.5 (0.097)
	Median	75 th percentile							
Unadjusted	2.5	5.1							
MIBP	-3.5 (0.097)								
<i>Cryptorchidism or testicular position</i>									
<p>Swan (2008) (United States; Minnesota, Missouri, California) Population: 106 boys from birth cohort study (Study for Future Families), 2000–2002, mean age 12.8 mo (0–36 mo) Outcome: Incomplete testicular descent assessed at clinical exam (10% prevalence) Exposure: Maternal urine sample, 3rd trimester MIBP in urine (ng/mL):</p> <table border="0" data-bbox="190 1396 782 1459"> <tr> <td></td> <td>Median</td> <td>75th percentile</td> </tr> <tr> <td>Unadjusted</td> <td>2.5</td> <td>5.1</td> </tr> </table> <p>Analysis: Logistic regression, adjusting for age and weight percentile Related references: Swan et al. (2005) (exposure data)</p>		Median	75 th percentile	Unadjusted	2.5	5.1	<p>MIBP reported as not associated with testicular position (quantitative results not reported)</p>		
	Median	75 th percentile							
Unadjusted	2.5	5.1							

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Reference and study design	Results																		
<i>Gender-related play</i>																			
<p>Swan et al. (2010) (United States; Minnesota, Missouri, California, Iowa) Population: 145 children from birth cohort study (Study for Future Families), 2000–2002 and 2002–2005 (Iowa), ages 4–7 yrs; second follow-up Outcome: Gender-specific play based on Pre-School Activities Inventory (24 items completed by parent or caregiver; subscores of male-oriented items and female-oriented items and a composite score consisting of male summation minus the female summation scores) Exposure: Maternal urine sample, 3rd trimester Unadjusted MIBP in urine (ng/mL):</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> <td align="center">75th percentile</td> </tr> <tr> <td>Boys</td> <td align="center">2.4</td> <td align="center">5.1</td> </tr> <tr> <td>Girls</td> <td align="center">2.8</td> <td align="center">5.0</td> </tr> </table> <p>Analysis: Regression analysis using Generalized Linear Models, considering creatinine, sex and age of child, maternal age, parental education, number of same and opposite sex siblings, ethnicity, clinic location, and parental attitude as potential covariates Related references: Swan et al. (2005) (exposure data)</p>		Median	75 th percentile	Boys	2.4	5.1	Girls	2.8	5.0	<p>Regression coefficient (95% CI) for pre-school activities index scores and log-transformed MIBP (adjusted for child’s age, mother’s age, mother’s education, parents’ attitude toward boy’s play, and interaction between education and attitude; negative value indicates less masculine play behavior with higher metabolite level)</p> <table border="0"> <tr> <td></td> <td align="center">Boys</td> <td align="center">Girls</td> </tr> <tr> <td>Masculine</td> <td align="center">-1.65 (-4.57, 1.28)</td> <td align="center">1.04 (-1.75, 3.82)</td> </tr> <tr> <td>Composite</td> <td align="center">-4.53 (-8.12, -0.94)</td> <td align="center">0.38 (-3.86, 4.63)</td> </tr> </table>		Boys	Girls	Masculine	-1.65 (-4.57, 1.28)	1.04 (-1.75, 3.82)	Composite	-4.53 (-8.12, -0.94)	0.38 (-3.86, 4.63)
	Median	75 th percentile																	
Boys	2.4	5.1																	
Girls	2.8	5.0																	
	Boys	Girls																	
Masculine	-1.65 (-4.57, 1.28)	1.04 (-1.75, 3.82)																	
Composite	-4.53 (-8.12, -0.94)	0.38 (-3.86, 4.63)																	

- 1
- 2 CI = confidence interval; MIBP = monoisobutyl phthalate

1 **3.2.2. Male Reproductive Effects in Humans**

2 **Table 3-2. Evidence pertaining to DIBP and semen parameters or infertility in**
 3 **adult men or couples**

Reference ^a and study design	Results													
<p>Kranvogel et al. (2014) (Slovenia) Population: 136 men from couples seeking infertility treatment (mean age 36.2 yrs, range 24–54 yrs), 2012 Outcome: Semen analysis Exposure: Urine sample, collected at same time as semen sample MIBP in urine:</p> <table align="center"> <tr> <td></td> <td>Median</td> <td>Maximum</td> </tr> <tr> <td>Unadjusted (µg/L)</td> <td>21.6</td> <td>161.8</td> </tr> <tr> <td>Cr-adjusted (µg/g Cr)</td> <td>20.8</td> <td>119.2</td> </tr> </table> <p>Analysis: Spearman correlation</p>		Median	Maximum	Unadjusted (µg/L)	21.6	161.8	Cr-adjusted (µg/g Cr)	20.8	119.2	<p>Spearman correlation coefficient, MIBP and sperm parameters:</p> <table align="center"> <tr> <td>Sperm concentration</td> <td>–0.044</td> </tr> <tr> <td>Sperm motility</td> <td>–0.075</td> </tr> </table> <p>($p > 0.05$ for both parameters)</p>	Sperm concentration	–0.044	Sperm motility	–0.075
	Median	Maximum												
Unadjusted (µg/L)	21.6	161.8												
Cr-adjusted (µg/g Cr)	20.8	119.2												
Sperm concentration	–0.044													
Sperm motility	–0.075													
<p>Joensen et al. (2012) (Denmark) Population: 881 men from general population, assessed at military conscript exam*, 2007–2009, median age 19.1 yrs (5th–95th percentile: 18.4–22.0 yrs) Outcome: Semen analysis Exposure: Urine sample, collected at same time as semen sample MIBP in urine (ng/mL):</p> <table align="center"> <tr> <td></td> <td>Median</td> <td>95th percentile</td> </tr> <tr> <td>Unadjusted</td> <td>58</td> <td>173</td> </tr> </table> <p>Analysis: Linear regression, considering age, BMI, smoking, alcohol consumption, ethnicity, BMI squared, in utero exposure to tobacco smoke, previous or current diseases, recent fever, recent use of medication, abstinence time, and time from ejaculation to analysis as potential covariates *As reported by Ravnborg et al. (2011)</p>		Median	95 th percentile	Unadjusted	58	173	<p>Results for individual phthalate metabolites (including MIBP) reported as “few significant associations” with sperm volume, count, or percentage progressively motile sperm (quantitative results not reported). Sperm concentration analysis adjusted for abstinence time (volume, concentration, and count); sperm motility analysis adjusted for time from ejaculation to analysis (progressively motile); analysis of percent of morphologically normal sperm was unadjusted</p>							
	Median	95 th percentile												
Unadjusted	58	173												
<p>Wirth et al. (2008) (United States, Michigan) Population: 45 male partners seen in infertility clinic, time period not reported; mean age 34 yrs Outcome: Semen analysis Exposure: Urine sample, collected at same time as semen sample (all between 7 and 11 am) MIBP in urine (ng/mL) (percentile):</p> <table align="center"> <tr> <td>Median</td> <td>75th percentile</td> <td>95th percentile</td> </tr> <tr> <td>5.8</td> <td>10.0</td> <td>17.9</td> </tr> </table> <p>Analysis: Dichotomized outcomes (above and below WHO reference values), MIBP dichotomized at median; age, education (three levels), income (three levels), race, BMI (three levels), current smoking status, and alcohol use (two levels) considered as potential confounders;</p>	Median	75 th percentile	95 th percentile	5.8	10.0	17.9	<p>The combined measure for MIBP and MBP was not associated with any sperm parameter, nor was MIBP when analyzed individually (data not shown)</p>							
Median	75 th percentile	95 th percentile												
5.8	10.0	17.9												

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Table 3-3. Evidence pertaining to DIBP and reproductive hormones in adult men

Reference and study design	Results						
<p>Joensen et al. (2012) (Denmark) Population: 881 men from general population, assessed at military conscript exam*, 2007–2009, median age 19.1 yrs (5th–95th percentile: 18.4–22.0 yrs) Outcome: Serum steroidal and gonadotropin hormones Exposure: Urine sample, collected at same time as serum sample for hormone analysis MIBP in urine (ng/mL):</p> <table align="center"> <tr> <td></td> <td>Median</td> <td>95th percentile</td> </tr> <tr> <td>Unadjusted</td> <td>58</td> <td>173</td> </tr> </table> <p>Analysis: Linear regression considering age, BMI, smoking, alcohol consumption, time of blood sampling, assay type, ethnicity, BMI squared, in utero exposure to tobacco smoke, previous or current diseases, recent fever, and recent use of medication as potential covariates *As reported by Ravnborg et al. (2011)</p>		Median	95 th percentile	Unadjusted	58	173	<p>Results for individual phthalate metabolites (including MIBP) reported as “few significant associations” with free testosterone, estradiol, SHBG, LH, inhibin-B, or FSH (quantitative results not reported); analyses adjusted for age, BMI, smoking, alcohol consumption, and time of blood sampling (and assay type for inhibin-B only)</p>
	Median	95 th percentile					
Unadjusted	58	173					
<p>Mendiola et al. (2011) (United States; Minnesota, Missouri, California, Iowa, New York) Population: 425 men whose partners enrolled in birth cohort study (Study for Future Families), 1999–2005, mean age 32 yrs Outcome: Serum steroidal and gonadotropin hormones Exposure: Urine sample, collected at same time as serum sample for hormone analysis MIBP in urine (ng/mL) (distribution not reported) Analysis: Pearson correlation of log(10)-transformed MIBP and hormone measures; linear regression considering age, age square, BMI, smoking status, ethnicity, urinary creatinine concentration, time of sample collection, time of collection squared, season, educational level, center, and stressful life events)</p>	<p>Authors reported “little or no association with metabolites of phthalate other than DEHP” [including MIBP] with testosterone, estradiol, SHBG, LH, inhibin-B, or FSH (quantitative results not reported)</p>						

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DEHP = diethylhexyl phthalate; FSH = follicle-stimulating hormone; LH = luteinizing hormone; MOINP = oxo-(mono-oxoisobutyl) phthalate; SD = standard deviation; SHBG = sex hormone binding globulin

1 3.2.3. Male Pubertal Development in Humans

2 Table 3-4. Evidence pertaining to DIBP and the timing of male puberty or sex
3 hormones in boys

Reference and study design	Results																																																		
<p>Ferguson et al. (2014b) (Mexico) Population: 115 boys ages 8–14 yrs from a birth cohort (Early Life Exposure in Mexico to Environmental Toxicants, participants enrolled during first trimester 1994–2004), follow-up initiated in 2010 Outcome: Adrenarche or puberty, based on Tanner staging by physician (pubic hair stage ≥ 2; genitalia stage ≥ 2 or testicular volume >3 mL); serum hormone level Exposure: Maternal urine sample (n = 107) from third trimester or child’s urine sample (n = 113) collected at time of Tanner staging and serum collection Unadjusted MIBP in urine (ng/mL): <table border="1" data-bbox="190 871 771 976"> <thead> <tr> <th></th> <th>Median</th> <th>95th percentile</th> </tr> </thead> <tbody> <tr> <td>Maternal sample</td> <td>1.83</td> <td>6.64</td> </tr> <tr> <td>Child’s sample</td> <td>9.61</td> <td>36.1</td> </tr> </tbody> </table> Analysis: Logistic regression for analysis of puberty onset, adjusting for variables shown in results column; linear regression for analysis of hormone levels, considering age, BMI z-score, socioeconomic status, and maternal smoking as potential covariates</p>		Median	95 th percentile	Maternal sample	1.83	6.64	Child’s sample	9.61	36.1	<p>OR (95% CI) for adrenarche or puberty per interquartile increase in ln-transformed MIBP (adjusted for child age, BMI z-score, and urine specific gravity)</p> <p style="text-align: center;">Exposure basis</p> <table border="1" data-bbox="771 556 1421 892"> <thead> <tr> <th></th> <th>Maternal urine (prenatal)</th> <th>Child urine</th> </tr> </thead> <tbody> <tr> <td>Tanner stage or testicular volume</td> <td></td> <td></td> </tr> <tr> <td>Pubic hair (stage ≥ 2)</td> <td>0.29 (0.07, 1.30)</td> <td>0.76 (0.32, 1.81)</td> </tr> <tr> <td>Genitalia (stage ≥ 2)</td> <td>0.71 (0.37, 1.35)</td> <td>0.76 (0.39, 1.49)</td> </tr> <tr> <td>Testicular volume (>3 mL)</td> <td>1.60 (0.70, 3.65)</td> <td>2.17 (0.81, 5.82)</td> </tr> </tbody> </table> <p>Percent change (95% CI) in serum hormone level per interquartile increase in ln-transformed MIBP (adjusted for urine specific gravity, child age, and BMI z-score)</p> <p style="text-align: center;">Exposure basis</p> <table border="1" data-bbox="771 1060 1421 1451"> <thead> <tr> <th></th> <th>Maternal urine (prenatal)</th> <th>Child urine</th> </tr> </thead> <tbody> <tr> <td>Serum hormone</td> <td></td> <td></td> </tr> <tr> <td>Testosterone</td> <td>5.12 (–23.3, 44.0)</td> <td>–26.2 (–45.6, 0.16)</td> </tr> <tr> <td>Free testosterone</td> <td>1.69 (–26.7, 41.1)</td> <td>–27.9 (–47.8, –0.60)</td> </tr> <tr> <td>SHBG</td> <td>5.72 (–5.18, 17.9)</td> <td>2.20 (–8.41, 14.1)</td> </tr> <tr> <td>DHEAS</td> <td>–2.02 (–15.9, 14.1)</td> <td>3.02 (–11.4, 19.8)</td> </tr> <tr> <td>Estradiol</td> <td>–1.94 (–11.2, 8.23)</td> <td>–12.3 (–20.2, –3.54)</td> </tr> <tr> <td>Inhibin B</td> <td>–1.98 (–12.7, 10.1)</td> <td>2.73 (–8.24, 15.0)</td> </tr> </tbody> </table>				Maternal urine (prenatal)	Child urine	Tanner stage or testicular volume			Pubic hair (stage ≥ 2)	0.29 (0.07, 1.30)	0.76 (0.32, 1.81)	Genitalia (stage ≥ 2)	0.71 (0.37, 1.35)	0.76 (0.39, 1.49)	Testicular volume (>3 mL)	1.60 (0.70, 3.65)	2.17 (0.81, 5.82)		Maternal urine (prenatal)	Child urine	Serum hormone			Testosterone	5.12 (–23.3, 44.0)	–26.2 (–45.6, 0.16)	Free testosterone	1.69 (–26.7, 41.1)	–27.9 (–47.8, –0.60)	SHBG	5.72 (–5.18, 17.9)	2.20 (–8.41, 14.1)	DHEAS	–2.02 (–15.9, 14.1)	3.02 (–11.4, 19.8)	Estradiol	–1.94 (–11.2, 8.23)	–12.3 (–20.2, –3.54)	Inhibin B	–1.98 (–12.7, 10.1)	2.73 (–8.24, 15.0)
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Inhibin B	–1.98 (–12.7, 10.1)	2.73 (–8.24, 15.0)																																																	
<p>Mouritsen et al. (2013b) (Denmark) Population: Boys from population-based cohort (COPENHAGEN Puberty Study), 2006–2010; age 11 yrs (53 boys) or 13 yrs (31 boys) Outcome: Adrenarche or puberty, based on Tanner staging by physician (pubarche = pubic hair stage ≥ 2 and testicular volume >3 mL); serum hormone level Exposure: Urine sample, first morning sample; data reported in Mouritsen et al. (2013a), Supplemental Material MIBP + MBP in urine (ng/mL)^a: <table border="1" data-bbox="190 1816 771 1896"> <thead> <tr> <th></th> <th>Geometric mean</th> <th>Maximum</th> </tr> </thead> <tbody> <tr> <td></td> <td>118</td> <td>676</td> </tr> </tbody> </table> </p>		Geometric mean	Maximum		118	676	<p>Median age (yrs) at development by ΣMIBP + MBP level (evaluation at 11 yrs)</p> <table border="1" data-bbox="771 1543 1421 1690"> <thead> <tr> <th></th> <th>Low</th> <th>High</th> </tr> </thead> <tbody> <tr> <td>Pubarche</td> <td>12.3</td> <td>11.0 ($p < 0.05$)</td> </tr> <tr> <td>Testicular volume >3 mL</td> <td>11.5</td> <td>11.1</td> </tr> </tbody> </table> <p>Median hormone concentration by MIBP + MBP level (evaluation at 11 yrs)</p> <table border="1" data-bbox="771 1774 1421 1896"> <thead> <tr> <th></th> <th>Low</th> <th>High</th> </tr> </thead> <tbody> <tr> <td>Testosterone (nmol/L)</td> <td><0.23</td> <td><0.23</td> </tr> <tr> <td>DHEAS (μmol/L)</td> <td>2.02</td> <td>1.61</td> </tr> </tbody> </table>				Low	High	Pubarche	12.3	11.0 ($p < 0.05$)	Testicular volume >3 mL	11.5	11.1		Low	High	Testosterone (nmol/L)	<0.23	<0.23	DHEAS (μ mol/L)	2.02	1.61																								
	Geometric mean	Maximum																																																	
	118	676																																																	
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																		
(based on larger sample of 84 boys) Analysis: Two-tailed Mann-Whitney U-test for comparisons between groups, comparing median hormone levels and pubertal stage in “high” and “low” phthalate groups (based on above or below group mean excretion)	Adione (nmol/L)	1.28	1.22																
	Estradiol (pmol/L)	<18	<18																
	FSH (IU/L)	1.28	1.68																
	LH (IU/L)	0.28	0.27																
	Median age (yrs) at development by MIBP + MBP level (evaluation at 13 yrs)																		
		Low	High																
	Pubarche	12.5	12.1																
	Testicular volume >3 mL	11.6	11.6																
	Median age (yrs) at development by MIBP + MBP level (evaluation at 13 yrs)																		
		Low	High																
	Testosterone (nmol/L)	5.1	7.7																
	DHEAS (μmol/L)	2.61	3.64																
	Adione (nmol/L)	2.96	3.85																
	Estradiol (pmol/L)	19	37																
	FSH (IU/L)	2.4	2.5																
	LH (IU/L)	1.8	1.4																
Mieritz et al. (2012) (Denmark) Population: 38 boys with pubertal gynecomastia and 190 age-matched controls drawn from 555 boys from population-based cohort (COPENHAGEN Puberty Study), 2006–2008; ages 6–19 yrs Outcome: Anthropometry, pubertal stage (pubic hair and genital development), presence of gynecomastia, and serum testosterone Exposure: Urine sample, first morning sample MIBP in urine (ng/mL): <table border="0" style="margin-left: 40px;"> <tr> <td></td> <td align="center">Median</td> <td align="center">95th percentile</td> </tr> <tr> <td>Group 3</td> <td align="center">74.88</td> <td align="center">229.1</td> </tr> </table> (boys without gynecomastia, all ages) Analysis: Two-tailed Mann-Whitney U-test for comparisons between groups; linear regression with age adjustment for association with serum testosterone; probit analysis with phthalate concentrations divided in quartiles for analysis of puberty timing		Median	95 th percentile	Group 3	74.88	229.1	MIBP concentration (ng/mL) by group <table border="0" style="margin-left: 40px;"> <tr> <td></td> <td align="center">Group 1 (n = 38)</td> <td align="center">Group 2 (n = 189)</td> <td align="center">Group 3 (n = 517)</td> </tr> <tr> <td>Median</td> <td align="center">68.50</td> <td align="center">73.96</td> <td align="center">74.88</td> </tr> <tr> <td>95th percentile</td> <td align="center">178.8</td> <td align="center">199.5</td> <td align="center">229.1</td> </tr> </table> Group 1 = boys with palpable gynecomastia Group 2 = boys without palpable gynecomastia (age-matched) Group 3 = boys without palpable gynecomastia (all ages) No association between MIBP concentration and timing of puberty or serum testosterone level (quantitative results not reported)		Group 1 (n = 38)	Group 2 (n = 189)	Group 3 (n = 517)	Median	68.50	73.96	74.88	95 th percentile	178.8	199.5	229.1
	Median	95 th percentile																	
Group 3	74.88	229.1																	
	Group 1 (n = 38)	Group 2 (n = 189)	Group 3 (n = 517)																
Median	68.50	73.96	74.88																
95 th percentile	178.8	199.5	229.1																

1
 2 ^aIn this population at this time, MIBP tended to be present at higher concentrations than MBP; EPA includes these
 3 studies in the DIBP tables, but recognizes the exposure misclassification introduced by the use of the summed
 4 concentration exposure measure.
 5

6 DHEAS = dehydroepiandrosterone; EPA = Environmental Protection Agency
 7

1 3.2.4. Female Pubertal Development in Humans

2 Table 3-5. Evidence pertaining to DIBP and timing of female puberty or sex
3 hormones in girls

Reference and study design	Results																						
<i>Precocious puberty and premature thelarche</i>																							
<p>Frederiksen et al. (2012) (Denmark) Population: 24 girls with precocious puberty (n = 13 with central precocious puberty, n = 6 with early normal puberty, n = 5 with premature thelarche) from outpatient clinic, 2008–2009 and 184* age-matched controls from population-based cohort (COPENHAGEN Puberty Study), recruited from high schools 2006–2008; age 7.4–9.9 yrs Outcome: Precocious puberty, early normal puberty, or premature thelarche based on Tanner staging by physician Exposure: Urine sample (child’s), first morning sample collected at clinical evaluation MIBP and MBP in urine (ng/mL)^a, controls (analysis based on sum of these two metabolites):</p> <table border="0" data-bbox="190 919 552 1018"> <tr> <td></td> <td>Median</td> <td>95th percentile</td> </tr> <tr> <td>MIBP</td> <td>81</td> <td>241</td> </tr> <tr> <td>MBP</td> <td>51</td> <td>153</td> </tr> </table> <p>(based on larger sample of 725 controls) Analysis: Urine concentrations in cases and controls compared with Mann-Whitney U test *Study reports number of controls inconsistently; text reports 164 controls, while Table 4 reports 184</p>		Median	95 th percentile	MIBP	81	241	MBP	51	153	<p>Median (range) ΣMIBP and MBP metabolites in urine (ng/mL) in cases and controls</p> <table border="0" data-bbox="776 520 1421 651"> <tr> <td></td> <td colspan="2">Precocious puberty</td> <td>(p-value)</td> </tr> <tr> <td>Controls</td> <td></td> <td></td> <td></td> </tr> <tr> <td>147 (22–2,195)</td> <td>94 (32–383)</td> <td></td> <td>(p < 0.01)</td> </tr> </table>		Precocious puberty		(p-value)	Controls				147 (22–2,195)	94 (32–383)		(p < 0.01)	
	Median	95 th percentile																					
MIBP	81	241																					
MBP	51	153																					
	Precocious puberty		(p-value)																				
Controls																							
147 (22–2,195)	94 (32–383)		(p < 0.01)																				
<p>Lomenick et al. (2010) (United States, Ohio and Kentucky) Population: 28 girls with central precocious puberty, 28 age- and race-matched controls; all recruited from pediatric endocrinology clinic, 2005–2008; mean age 7 yrs Outcome: Central precocious puberty defined based on clinical standards (appearance of physical characteristics of puberty before 8 yrs of age, with laboratory confirmation of central origin of breast development); no cases had received medical treatment prior to urine sample collection Exposure: Urine sample (child’s), collected at clinical evaluation MIBP in urine of controls:</p> <table border="0" data-bbox="190 1686 552 1785"> <tr> <td></td> <td>Mean ± SE</td> </tr> <tr> <td>Unadjusted (ng/mL)</td> <td>22.6 ± 7.6</td> </tr> <tr> <td>Cr-adjusted (µg/g Cr)</td> <td>20.2 ± 4.9</td> </tr> </table> <p>Analysis: MIBP concentrations in cases and controls compared with Wilcoxon rank-sum test</p>		Mean ± SE	Unadjusted (ng/mL)	22.6 ± 7.6	Cr-adjusted (µg/g Cr)	20.2 ± 4.9	<table border="0" data-bbox="776 1203 1421 1459"> <tr> <td></td> <td></td> <td>Central precocious puberty</td> <td>(p-value)</td> </tr> <tr> <td></td> <td>Controls</td> <td></td> <td></td> </tr> <tr> <td>Unadjusted (ng/mL)</td> <td>22.6 ± 7.6</td> <td>15.4 ± 2.9</td> <td>(0.77)</td> </tr> <tr> <td>Cr-adjusted (µg/g Cr)</td> <td>20.2 ± 4.9</td> <td>16.5 ± 2.1</td> <td>(0.96)</td> </tr> </table>			Central precocious puberty	(p-value)		Controls			Unadjusted (ng/mL)	22.6 ± 7.6	15.4 ± 2.9	(0.77)	Cr-adjusted (µg/g Cr)	20.2 ± 4.9	16.5 ± 2.1	(0.96)
	Mean ± SE																						
Unadjusted (ng/mL)	22.6 ± 7.6																						
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		Central precocious puberty	(p-value)																				
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Cr-adjusted (µg/g Cr)	20.2 ± 4.9	16.5 ± 2.1	(0.96)																				

Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																								
		Estradiol (pmol/L)	194	131																					
	FSH (IU/L)	4.9	5.8 ($p < 0.05$)																						
	LH (IU/L)	3.8	3.8																						
<p>Frederiksen et al. (2012) (Denmark) Population: 725 healthy girls ages 5.6–19.1 yrs from COPENHAGEN Puberty Study cohort, recruited from high schools during 2006–2008 Outcome: Stage of breast or pubic hair development based on Tanner staging by physician; Serum steroid and gonadotropin hormones Exposure: Urine sample (child’s), collected at time of pubertal stage assessment Unadjusted MIBP and MBP in urine (ng/mL)^a, all 725 participants:</p> <table border="0"> <tr> <td></td> <td>Median</td> <td>95th percentile</td> </tr> <tr> <td>MIBP</td> <td>81</td> <td>241</td> </tr> <tr> <td>MBP</td> <td>51</td> <td>153</td> </tr> </table> <p>Analysis: Probit analysis, results verified using Pool-Adjacent-Violators algorithm</p>		Median	95 th percentile	MIBP	81	241	MBP	51	153	<p>Mean age (95% CI) (yrs) at entry into breast stage 2 or pubic hair stage 2, by quartile of ΣMIBP + MBP metabolites:</p> <table border="0"> <tr> <td>ΣMIBP + MBP quartile</td> <td>Breast stage 2 (n = 394)</td> <td>Pubic hair stage (n not reported)</td> </tr> <tr> <td>1 (low)</td> <td>10.12 (9.61, 10.62)</td> <td>10.83 (10.54, 11.12)</td> </tr> <tr> <td>2</td> <td>9.97 (9.48, 10.46)</td> <td>10.97 (10.67, 11.28)</td> </tr> <tr> <td>3</td> <td>9.89 (9.40, 10.37)</td> <td>11.22 (10.93, 11.52)</td> </tr> <tr> <td>4 (high)</td> <td>9.79 (9.30, 10.30)</td> <td>11.54*(11.21, 11.88)</td> </tr> </table> <p>*Significantly different from quartile 1; $p < 0.05$</p> <p>Levels of FSH, LH, estradiol, and testosterone were similar across ΣMIBP + MBP metabolite exposure groups when adjusted for age distribution (quantitative results not reported)</p>	Σ MIBP + MBP quartile	Breast stage 2 (n = 394)	Pubic hair stage (n not reported)	1 (low)	10.12 (9.61, 10.62)	10.83 (10.54, 11.12)	2	9.97 (9.48, 10.46)	10.97 (10.67, 11.28)	3	9.89 (9.40, 10.37)	11.22 (10.93, 11.52)	4 (high)	9.79 (9.30, 10.30)	11.54*(11.21, 11.88)
	Median	95 th percentile																							
MIBP	81	241																							
MBP	51	153																							
Σ MIBP + MBP quartile	Breast stage 2 (n = 394)	Pubic hair stage (n not reported)																							
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2	9.97 (9.48, 10.46)	10.97 (10.67, 11.28)																							
3	9.89 (9.40, 10.37)	11.22 (10.93, 11.52)																							
4 (high)	9.79 (9.30, 10.30)	11.54*(11.21, 11.88)																							

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2 ^aIn this population at this time, MIBP tended to be present at higher concentrations than MBP; EPA includes these
3 studies in the DIBP tables, but recognizes the exposure misclassification introduced by the use of the summed
4 concentration exposure measure.
5

1 **3.2.5. Female Reproductive Effects in Humans**

2 **Table 3-6. Evidence pertaining to DIBP and reproductive hormones in adult**
 3 **women**

Reference and study design	Results	
<i>Maternal hormones during pregnancy</i>		
<p>Sathyanarayana et al. (2014) (United States; Minnesota, Missouri, California) Population: 180 mothers from birth cohort (Study for Future Families), recruited during pregnancy, 1999–2002 Outcome: Serum hormone levels, samples collected during prenatal clinic visit Exposure: Maternal urine sample, collected during 2nd or 3rd trimester MIBP in urine (ng/mL): Median 75th percentile Unadjusted 2.7 4.85 Analysis: Linear regression, log-transformed MIBP and log-transformed hormone level</p>	Regression coefficient (95% CI) for change in maternal log-transformed serum hormone level with unit increase in log-transformed MIBP, stratified by sex of fetus	
	Mothers with male fetus (n = 94)	Mothers with female fetus (n = 86)
	Testosterone (total)	-0.03 (-0.18, 0.13)
	Testosterone (free)	-0.10 (-0.28, 0.07)
	Estradiol	-0.03 (-0.20, 0.14)
	0.003	0.03 (-0.14, 0.20)
<p>Hart et al. (2013) (Australia) Population: 123 mothers from birth cohort (Western Australian Pregnancy Cohort), whose mothers were recruited at 18 wks of gestation between 1989 and 1991 Outcome: Reproductive and gonadotropin hormone levels in maternal serum collected at 18 and 34–36 wks of gestation Exposure: Maternal serum samples (n = 123) collected at 18 and 34–36 wks of gestation (combined aliquot from both time periods) MIBP in serum (ng/mL): Median 90th percentile MIBP 1.77 6.16 Analysis: Correlation between quartiles of serum MIBP and log-transformed hormone levels</p>	Correlation coefficient between log-transformed maternal serum hormone level and quartiles of MIBP in maternal serum	
	At 18 wks of gestation (n = 119)	At 34–36 wks of gestation (n = 114)
	Androstene-dione (nmol/L)	0.023
	DHEAS (µmol/L)	-0.060
	Testosterone (pmol/L)	-0.042
	SHBG (nmol/L)	0.003
	Free testosterone (pmol/L)	-0.101
	Free testosterone index	0.108
	-0.061	-0.020
	-0.051	-0.063
	-0.064	
	<i>p</i> > 0.10 for all correlations	

4

Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results
	Uterine volume (mL) $r \leq 0.20$ ($p \geq 0.17$) Ovarian volume (cm ³) $r \leq 0.10$ ($p \geq 0.29$) Antral follicle count $r \leq 0.12$ ($p \geq 0.20$) Authors reported no association between MIBP and polycystic ovarian syndrome using either definition (quantitative results not reported).

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PCO = polycystic ovarian morphology; PCOS = polycystic ovarian syndrome

1 3.2.6. Pregnancy Outcomes in Humans

2 Table 3-8. Evidence pertaining to DIBP and pregnancy outcomes in humans

Reference and study design	Results			
<i>Fetal growth (birth weight, birth length, head circumference)</i>				
<p>Huang et al. (2014b) (China) Population: 207 women delivering at one hospital in Chongqing between 2011 and 2012, aged 18–35 yrs and with no history of tobacco or alcohol use; mean age 28 yrs Outcome: Standard clinical measures at birth Exposure: Cord blood sample DIBP in cord blood (µg/L) Median 75th percentile 95th percentile All samples 16.7 26.9 114 Analysis: Linear regression, adjusting for variables shown in results column</p>	Regression coefficient (95% CI) for change in clinical measurement at birth per unit increase in ln-transformed DIBP (µg/L) (adjusted for gestational age):			
		Girls		Boys
	Birth weight (g)	-27 (-90, 36)	-87 (-195, 200)	
	Birth length (cm)	-0.06 (-0.45, 0.33)	-0.75 (-1.35, -0.15)	
Head circumference (mm)	-3.85 (-9.47, 1.76)	-2.76 (-7.62, 2.11)		
<p>Philippat et al. (2012) (France) Population: 72 cases with undescended testis or hypospadias, 215 matched controls from two birth cohorts (EDEN and PELAGIE), 2002–2006 Outcome: Standard clinical measurements at birth Exposure: Maternal urine sample, collected between 6 and 19 (PELAGIE) or between 24 and 30 (EDEN) wks of gestation MIBP in urine (ng/mL): Median 95th percentile Measured 45.9 219.0 Standardized* 64.7 365.3 Analysis: Cases and controls combined for this analysis; weighted linear regression using tertiles or ln-transformed urine concentrations, adjusting for variables shown in results column; analysis by tertiles for evaluation of possible non-monotonic relationship; analyses corrected for oversampling of malformation cases *Standardized for sampling conditions and gestational age at collection</p>	Regression coefficient (95% CI) for change in birth outcome by MIBP tertile and per unit change in ln-MIBP (standardized, ng/mL) (adjusted for gestational duration, maternal pre-pregnancy weight and height, maternal smoking, maternal education, parity, recruitment center, urine creatinine, and mode of delivery as potential covariate; head circumference model also adjusted for mode of delivery)			
	MIBP tertile (µg/L)	Birth weight (g)	Birth length (cm)	Head circumference (cm)
	1 (<48.2)	0 (referent)	0 (referent)	0 (referent)
	2 (48.2–97.9)	61 (-77, 200)	0.4 (-0.3, 1.1)	-0.1 (-0.6, 0.4)
	3 (≥97.9)	-31 (-190, 129)	0.3 (-0.4, 1.0)	0.2 (-0.5, 0.9)
	(trend p-value)	(0.48)	(0.54)	(0.40)
	ln (MIBP)	-44 (-110, 23)	0.0 (-0.3, 0.3)	-0.1 (-0.4, 0.1)
	<p>Wolff et al. (2008) (United States, New York City) Population: 382 singleton live births without medical complications from birth cohort (Mt. Sinai Children’s Environmental Health study), 1998–2002 Outcome: Standard clinical measurements at birth Exposure: Maternal urine sample, third trimester MIBP in urine (ng/mL): Median 75th percentile Unadjusted 6.2 12 Analysis: Linear regression, adjusting for variables shown in results column</p>	Regression coefficient (95% CI) for change in birth outcome with unit increase in ln-MIBP (ng/mL) (adjusted for race/ethnicity, infant sex, gestational age at delivery, ln-creatinine, prenatal smoking, pre-pregnancy BMI, maternal education, and marital status)		
Birth weight (g)		-14 (-57, 28)		
Birth length (cm)		0.04 (-0.19, 0.28)		
Head circumference (cm)		0.05 (-0.11, 0.21)		
Restricted to observations with creatinine ≥20 mg/dL				

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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																											
<i>Preterm birth (<37 wks) and gestational age</i>																												
<p>(Ferguson et al. (2014a); Ferguson et al. (2014c)) (United States; Boston) Population: 130 cases, 352 controls from pregnancy cohort (study of predictors of pre-eclampsia, enrolled during first trimester, 2006–2008); controls randomly selected from among those delivering ≥37 wks of gestation; mean age 33 yrs Outcome: Preterm birth (<37 wks of gestation; gestation estimated from first trimester ultrasound); additional analysis of subgroup with spontaneous preterm labor or preterm premature rupture of membranes (“spontaneous preterm,” n = 57) Exposure: Maternal urine sample, one to three samples collected at median times of 9.7, 17.9, or 26.0 wks of gestation; geometric mean of results from visits 1–3 used in analyses. MIBP in urine, SG-adjusted (µg/L):</p> <table border="0"> <tr> <td></td> <td align="center">Geometric mean</td> <td align="center">75th percentile</td> </tr> <tr> <td>Controls</td> <td align="center">6.71</td> <td align="center">10.3</td> </tr> <tr> <td>All cases</td> <td align="center">6.85</td> <td align="center">10.5</td> </tr> </table> <p>Analysis: Logistic regression (ln-transformed metabolites), considering average specific gravity, maternal age, race/ethnicity, education level, health insurance provider, BMI at first study visit, smoking status, alcohol use, parity, use of assisted-reproductive technology, and sex of infant as potential covariates Related reference: Ferguson et al. (2014c) (analysis by individual sample results for the four visits)</p>		Geometric mean	75 th percentile	Controls	6.71	10.3	All cases	6.85	10.5	<p>OR (95% CI) for preterm birth per unit increase in ln-transformed MIBP (adjusted for average specific gravity, maternal age, race/ethnicity, education level, and insurance provider)</p> <table border="0"> <tr> <td>All preterm</td> <td align="center">0.98</td> <td align="center">(0.72, 1.34)</td> </tr> <tr> <td>Spontaneous preterm</td> <td align="center">1.52</td> <td align="center">(0.97, 2.38)</td> </tr> </table> <p>[Results weaker than those seen with DEHP metabolites]</p> <p>Results by study visit from Ferguson et al. (2014c), all pre-term births</p> <table border="0"> <tr> <td>Visit 1</td> <td align="center">0.92</td> <td align="center">(0.57, 1.47)</td> </tr> <tr> <td>Visit 2</td> <td align="center">0.88</td> <td align="center">(0.54, 1.41)</td> </tr> <tr> <td>Visit 3</td> <td align="center">0.75</td> <td align="center">(0.50, 1.13)</td> </tr> <tr> <td>Visit 4</td> <td align="center">0.66</td> <td align="center">(0.28, 1.55)</td> </tr> </table>	All preterm	0.98	(0.72, 1.34)	Spontaneous preterm	1.52	(0.97, 2.38)	Visit 1	0.92	(0.57, 1.47)	Visit 2	0.88	(0.54, 1.41)	Visit 3	0.75	(0.50, 1.13)	Visit 4	0.66	(0.28, 1.55)
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<p>Huang et al. (2014b) (China) Population: 207 women delivering at one hospital in Chongqing between 2011 and 2012; aged 18–35 yrs and with no history of tobacco or alcohol use; mean age 28 yrs Outcome: Preterm birth (<37 wks of gestation; gestational age estimated from last menstrual period) Exposure: Cord blood sample DIBP in cord blood (µg/L)</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> <td align="center">75th percentile</td> <td align="center">95th percentile</td> </tr> <tr> <td>All samples</td> <td align="center">16.7</td> <td align="center">26.9</td> <td align="center">114</td> </tr> </table> <p>Analysis: Logistic and linear regression, adjusting for variables shown in results column</p>		Median	75 th percentile	95 th percentile	All samples	16.7	26.9	114	<p>OR (95% CI) for preterm delivery comparing ln-DIBP above and below the median (adjusted for maternal age, BMI, frequency of prenatal exam, and pregnancy history), with additional stratification by history of intravenous infusions</p> <table border="0"> <tr> <td>Total sample (n = 207)</td> <td align="center">6.01</td> <td align="center">(3.24, 11.17)</td> </tr> <tr> <td>No intravenous infusions (n = 154)</td> <td align="center">4.78</td> <td align="center">(1.68, 13.57)</td> </tr> <tr> <td>Intravenous infusions (n = 53)</td> <td align="center">6.07</td> <td align="center">(2.66, 13.83)</td> </tr> </table> <p>[History of intravenous infusions present in 26% of total and 55% of preterm birth group]</p> <p>Regression coefficient (95% CI) for change in gestational age (wks) per unit increase in ln-transformed DIBP (µg/L) (adjusted for maternal age, BMI, frequency of prenatal examination, history of intravenous infusions therapy, and pregnancy history)</p> <p align="center">-0.75 (-1.03, -0.46)</p>	Total sample (n = 207)	6.01	(3.24, 11.17)	No intravenous infusions (n = 154)	4.78	(1.68, 13.57)	Intravenous infusions (n = 53)	6.07	(2.66, 13.83)										
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																		
<p>Meeker et al. (2009) (Mexico) Population: 30 cases, 30 controls (term births) from pregnancy cohort, 2001–2003. Outcome: Preterm birth (<37 wks of gestation), determined using maternal recall of last menstrual period Exposure: Maternal urine sample, third trimester MIBP in urine, among term births</p> <table border="0"> <tr> <td></td> <td align="center" colspan="2">Median 75th percentile</td> </tr> <tr> <td>Unadjusted</td> <td align="center">2.0</td> <td align="center">4.1</td> </tr> <tr> <td>SG-adjusted (µg/L)</td> <td align="center">2.3</td> <td align="center">5.0</td> </tr> <tr> <td>Cr-adjusted (µg/g Cr)</td> <td align="center">3.7</td> <td align="center">6.6</td> </tr> </table> <p>Analysis: Logistic regression, considering maternal age, pre-pregnancy BMI, parity, education, marital status, infant’s sex, and gestational age at urine sample as potential covariates</p>		Median 75 th percentile		Unadjusted	2.0	4.1	SG-adjusted (µg/L)	2.3	5.0	Cr-adjusted (µg/g Cr)	3.7	6.6	<p>OR (95% CI) for preterm birth by MIBP above compared with below the median (adjusted for marital status, maternal education, and infant sex and gestational age at time of urine sample)</p> <table border="0"> <tr> <td>Cr-unadjusted (µg/L)</td> <td align="center">3.6 (1.1, 12.2)</td> </tr> <tr> <td>SG-adjusted (µg/L)</td> <td align="center">2.0 (0.7, 6.0)</td> </tr> <tr> <td>Cr-adjusted (µg/g Cr)</td> <td align="center">1.5 (0.5, 4.5)</td> </tr> </table>	Cr-unadjusted (µg/L)	3.6 (1.1, 12.2)	SG-adjusted (µg/L)	2.0 (0.7, 6.0)	Cr-adjusted (µg/g Cr)	1.5 (0.5, 4.5)
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1 3.2.7. Immune Effects in Humans

2 Table 3-9. Evidence pertaining to DIBP and allergy/immune effects in humans

Reference and study design	Results																																																																																
<p>Ait Bamai et al. (2014)^a (Japan) Population: Children (n = 122, ages <15 yrs) and adults (n = 374, ages ≥15 yrs) living in 148 detached dwellings in which at least 25 mg of dust was collected; 2006 follow-up of 2003 baseline survey Outcome: Allergic condition assessed by self-administered questionnaire (positive response to: in the past 2 yrs have you been seen at a hospital for allergic rhinitis, allergic conjunctivitis, or atopic dermatitis?); parents completed questionnaires for children <6 yrs Exposure: Dust samples DIBP in dust (µg/g dust):</p> <table border="0" style="margin-left: auto; margin-right: auto;"> <tr> <td></td> <td style="text-align: center;">Median</td> <td style="text-align: center;">75th</td> <td></td> </tr> <tr> <td></td> <td style="text-align: center;">percentile</td> <td></td> <td></td> </tr> <tr> <td>Floor dust (n = 148)</td> <td style="text-align: center;">2.4</td> <td style="text-align: center;">5.5</td> <td></td> </tr> <tr> <td>Multi-surface dust (n = 120)</td> <td style="text-align: center;">1.9</td> <td style="text-align: center;">3.5</td> <td></td> </tr> </table> <p>Analysis: Generalized linear mixed effects model, considering gender, age strata (<15, ≥15 yrs), smoking status (personal and environmental tobacco smoke), furry pets in home, signs of dampness, Der 1 (not defined by authors), other phthalates dust, airborne fungi, formaldehyde, total VOC, and building characteristics as potential covariates</p>		Median	75 th			percentile			Floor dust (n = 148)	2.4	5.5		Multi-surface dust (n = 120)	1.9	3.5		<p>OR (95% CI) for allergic condition by tertile of DIBP in floor dust (µg/g dust)(adjusted for adjusted for gender, age strata, smoking status, dampness index, furry pets inside the home, Der 1, and sum of other phthalates)</p> <table border="0" style="width: 100%;"> <tr> <td style="width: 15%;">DIBP tertile</td> <td style="width: 25%;">Full sample</td> <td style="width: 25%;">Children</td> <td style="width: 35%;">Adults</td> </tr> <tr> <td colspan="4" style="text-align: center;">Allergic rhinitis</td> </tr> <tr> <td>1 (low)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> </tr> <tr> <td>2</td> <td>1.87 (0.83, 4.22)</td> <td>3.54 (0.86, 14.5)</td> <td>0.99 (0.47, 2.05)</td> </tr> <tr> <td>3 (high)</td> <td>1.05 (0.47, 2.32)</td> <td>2.30 (0.60, 8.89)</td> <td>0.48 (0.22, 1.02)</td> </tr> <tr> <td>(trend p-value)</td> <td>(0.91)</td> <td>(0.23)</td> <td>(0.06)</td> </tr> <tr> <td colspan="4" style="text-align: center;">Allergic conjunctivitis</td> </tr> <tr> <td>1 (low)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> </tr> <tr> <td>2</td> <td>1.07 (0.38, 3.01)</td> <td>1.97 (0.35, 11.1)</td> <td>0.59 (0.19, 1.8)</td> </tr> <tr> <td>3 (high)</td> <td>1.64 (0.64, 4.18)</td> <td>3.27 (0.68, 15.7)</td> <td>0.82 (0.31, 2.2)</td> </tr> <tr> <td>(trend p-value)</td> <td>(0.30)</td> <td>(0.14)</td> <td>(0.69)</td> </tr> <tr> <td colspan="4" style="text-align: center;">Atopic dermatitis</td> </tr> <tr> <td>1 (low)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> </tr> <tr> <td>2</td> <td>5.52 (1.68, 18.1)</td> <td>11.95 (1.37, 104)</td> <td>2.55 (0.89, 7.31)</td> </tr> <tr> <td>3 (high)</td> <td>4.84 (1.46, 16.0)</td> <td>15.0 (1.91, 118)</td> <td>1.56 (0.44, 5.53)</td> </tr> <tr> <td>(trend p-value)</td> <td>(0.01)</td> <td>(0.01)</td> <td>(0.49)</td> </tr> </table> <p>p-value for age interaction >0.05 for all endpoints</p> <p>No increased aORs (either in the full sample or stratified by age) were observed in analyses using DIBP measurements in multisurface dust.</p>	DIBP tertile	Full sample	Children	Adults	Allergic rhinitis				1 (low)	1.0 (referent)	1.0 (referent)	1.0 (referent)	2	1.87 (0.83, 4.22)	3.54 (0.86, 14.5)	0.99 (0.47, 2.05)	3 (high)	1.05 (0.47, 2.32)	2.30 (0.60, 8.89)	0.48 (0.22, 1.02)	(trend p-value)	(0.91)	(0.23)	(0.06)	Allergic conjunctivitis				1 (low)	1.0 (referent)	1.0 (referent)	1.0 (referent)	2	1.07 (0.38, 3.01)	1.97 (0.35, 11.1)	0.59 (0.19, 1.8)	3 (high)	1.64 (0.64, 4.18)	3.27 (0.68, 15.7)	0.82 (0.31, 2.2)	(trend p-value)	(0.30)	(0.14)	(0.69)	Atopic dermatitis				1 (low)	1.0 (referent)	1.0 (referent)	1.0 (referent)	2	5.52 (1.68, 18.1)	11.95 (1.37, 104)	2.55 (0.89, 7.31)	3 (high)	4.84 (1.46, 16.0)	15.0 (1.91, 118)	1.56 (0.44, 5.53)	(trend p-value)	(0.01)	(0.01)	(0.49)
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																																											
<p>(Callesen et al. (2014a); Callesen et al. (2014b))^a (Denmark)</p> <p>Population: 81 rhinoconjunctivitis cases, 88 atopic dermatitis cases, 242 healthy controls group from population-based survey (Indoor Environment and Children’s Health); ages 3–5 yrs</p> <p>Outcome: Clinical exam and parent interview; allergic rhinoconjunctivitis: recurrence of at least two or more nasal symptoms (pruritus, runny nose, sneezing spells >20, nasal stenosis/mouth breathing) and ocular symptoms (itching, conjunctival injection, or watery secretion in both eyes) when exposed to allergens; atopic dermatitis: presence of at least 3 of 4 major features and 3 of 23 minor features; 70% of rhinoconjunctivitis and 50% of atopic dermatitis cases were IgE positive based on 20 allergen tests</p> <p>Exposure: DIBP concentrations in dust samples from bedroom and day care centers; total DIBP exposure estimated as a weighted mass fraction</p> <p>DIBP in dust among controls (µg/g):</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> </tr> <tr> <td>Home</td> <td align="center">27.0</td> </tr> <tr> <td>Day care</td> <td align="center">22.6</td> </tr> <tr> <td>Area-weighted</td> <td align="center">27.2</td> </tr> </table> <p>(weighted by assumed hrs in each environment)</p> <p>Analysis: Mann-Whitney U-test</p> <p>Related study: Callesen et al. (2014b) (same study population, with exposure measured in urine sample from participants)</p> <p>MIBP in urine: median 74.2 ng/mL (controls)</p>		Median	Home	27.0	Day care	22.6	Area-weighted	27.2	<p>Median DIBP in dust (µg/g), by case-control status assessed by clinical examination</p> <table border="0"> <tr> <td></td> <td align="center" colspan="3">Cases</td> </tr> <tr> <td></td> <td align="center">Controls (n = 242)</td> <td align="center">Rhinoconjunctivitis (n = 81)</td> <td align="center">Atopic dermatitis (n = 88)</td> </tr> <tr> <td>Home</td> <td align="center">27.0</td> <td align="center">30.4</td> <td align="center">33.4</td> </tr> <tr> <td>Day care</td> <td align="center">22.6</td> <td align="center">22.3</td> <td align="center">22.5</td> </tr> <tr> <td>Area-weighted</td> <td align="center">27.2</td> <td align="center">26.8</td> <td align="center">33.1</td> </tr> </table> <p>Similar results when based on case status defined by parent-questionnaire data (n = 56 rhinoconjunctivitis, n = 83 atopic dermatitis)</p> <p>Results from Callesen et al. (2014b): OR (95% CI) by quartile of MIBP (urine sample), adjusting for sex, breastfeeding less than 3 mo, smoking in the home, and single allergic predisposition</p> <table border="0"> <tr> <td></td> <td align="center">Rhinoconjunctivitis (76 cases, 222 controls)</td> <td align="center">Atopic dermatitis (76 cases, 216 controls)</td> </tr> <tr> <td>1</td> <td align="center">1.0 (referent)</td> <td align="center">1.0 (referent)</td> </tr> <tr> <td>2</td> <td align="center">1.18 (0.54, 2.55)</td> <td align="center">1.11 (0.53, 2.34)</td> </tr> <tr> <td>3</td> <td align="center">0.89 (0.39, 2.02)</td> <td align="center">0.88 (0.41, 1.91)</td> </tr> <tr> <td>4</td> <td align="center">1.07 (0.52, 2.22)</td> <td align="center">0.97 (0.48, 1.94)</td> </tr> </table>		Cases				Controls (n = 242)	Rhinoconjunctivitis (n = 81)	Atopic dermatitis (n = 88)	Home	27.0	30.4	33.4	Day care	22.6	22.3	22.5	Area-weighted	27.2	26.8	33.1		Rhinoconjunctivitis (76 cases, 222 controls)	Atopic dermatitis (76 cases, 216 controls)	1	1.0 (referent)	1.0 (referent)	2	1.18 (0.54, 2.55)	1.11 (0.53, 2.34)	3	0.89 (0.39, 2.02)	0.88 (0.41, 1.91)	4	1.07 (0.52, 2.22)	0.97 (0.48, 1.94)
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<p>Hoppin et al. (2013)^a (United States, NHANES) Population: 2,325 participants in population-based survey (NHANES), 2005–2006; ages ≥6 yrs Outcome: Self-administered questionnaire current allergy symptoms (hay fever, allergy, itchy rash, rhinitis) in past yr; allergic sensitization as measured by serum IgE (19 allergen specific IgEs, ≥0.35kU/L) Exposure: Urine sample collected same day as serum sample MIBP in urine (µg/L): Percentile Median 75th 95th Children 8.93 16.38 45.97 Adults 5.42 10.53 28.98 Analysis: Logistic regression, adjusting for variables shown in results column and sampling weights; separate analyses for children (ages 6–17 yrs) and adults (>17 yrs)</p>	<p>Prevalence and OR (95% CI) for allergy symptoms and allergic sensitization per unit change in log-transformed urinary MIBP level (adjusted for age, race/ethnicity, gender, BMI, creatinine, and cotinine)</p> <p>Children (n = 779)</p> <table> <tr> <td>Hay fever (n = 23)</td> <td align="center">3.6%</td> <td align="center">0.12(0.04, 0.39)</td> </tr> <tr> <td>Rhinitis (n = 188)</td> <td align="center">27.6%</td> <td align="center">0.84 (0.53, 1.33)</td> </tr> <tr> <td>IgE sensitization (any)</td> <td align="center">46.1%</td> <td align="center">0.93 (0.51, 1.70)</td> </tr> </table> <p>Adults (n = 1,546)</p> <table> <tr> <td>Hay fever (n = 88)</td> <td align="center">7.4%</td> <td align="center">0.93 (0.46, 1.87)</td> </tr> <tr> <td>Rhinitis (n = 498)</td> <td align="center">35.4%</td> <td align="center">0.99 (0.76, 1.29)</td> </tr> <tr> <td>IgE sensitization (any)</td> <td align="center">44.0%</td> <td align="center">1.32 (0.99, 1.76)</td> </tr> </table> <p>Authors reported that adjustment for poverty income ratio did not alter ORs.</p>	Hay fever (n = 23)	3.6%	0.12(0.04, 0.39)	Rhinitis (n = 188)	27.6%	0.84 (0.53, 1.33)	IgE sensitization (any)	46.1%	0.93 (0.51, 1.70)	Hay fever (n = 88)	7.4%	0.93 (0.46, 1.87)	Rhinitis (n = 498)	35.4%	0.99 (0.76, 1.29)	IgE sensitization (any)	44.0%	1.32 (0.99, 1.76)
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<p>Sun et al. (2009)^a (China) Population: Cases of rhinitis (n = 240) or eczema (n = 61) and controls (n = 204 and 119 for rhinitis and eczema analysis, respectively), all students of Tianjin University who had participated in a cross-sectional study of allergic symptoms and environmental factors; 2006–2007 Outcome: Self-reported symptoms from questionnaire: rhinitis = in past 12 mo, had a problem with sneezing, or a runny, or a blocked nose when not having a cold or the flu, or sneezing, or a runny, or a blocked nose, or itchy-watery eyes after contact with furred animals or after contact with pollen; eczema = in past 12 mo, had an itchy rash; controls responded no to questions on asthma/wheeze, rhinitis, and eczema Exposure: Surface dust sample in dorm rooms DIBP in dust (µg/g): Median 75th percentile 20.24 34.77 Analysis: Logistic regression for OR considering age, gender, passive smoking, smoking, pet raising, atopy, and building age as potential covariates; Mann-Whitney U-test for comparison between DIBP concentrations of cases and controls;</p>	<p>OR for rhinitis and eczema comparing DIBP in dust (µg/g dust) above and below the median (adjusted for age, gender, smoking, atopy and building age) reportedly did not reach statistical significance (quantitative results not reported)</p> <p>Median Concentration DIBP in dust (µg/g dust)</p> <table> <thead> <tr> <th></th> <th align="center">Cases</th> <th align="center">Controls</th> </tr> </thead> <tbody> <tr> <td>Rhinitis</td> <td align="center">20.17</td> <td align="center">28.76*</td> </tr> <tr> <td>Eczema</td> <td align="center">28.68</td> <td align="center">22.56</td> </tr> </tbody> </table> <p>*p = 0.019 by Mann-Whitney test; p = 0.051 by t-test.</p>		Cases	Controls	Rhinitis	20.17	28.76*	Eczema	28.68	22.56									
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Reference and study design	Results									
t-test for comparisons between log-transformed concentrations										
<p>Bornehag et al. (2004) (Sweden)</p> <p>Population: 198 cases, 202 controls from population-based cohort (Dampness in Buildings and Health cohort); n = 10,852; 2001–2002; ages 3–8 yrs</p> <p>Outcome: Eczema, wheezing, or rhinitis (Cases report at least two incidents of eczema, or wheezing or rhinitis without a cold, in the preceding yr, and at follow-up 1.5 yrs later)</p> <p>Exposure: Surface dust samples from children’s bedrooms</p> <p>DIBP in dust (mg/g):</p> <p align="center">Median</p> <p>All homes 0.045</p> <p>Analysis: Mann-Whitney U-test for comparing concentrations in all homes; t-test for comparing log-transformed concentrations in homes with concentrations above detection limit.</p>	<p>Concentration in dust (mg/g dust)</p> <table border="0"> <thead> <tr> <th></th> <th align="center">Median, all homes (n = 346)</th> <th align="center">Geometric mean (95% CI), homes with phthalate > detection limit (n = 290)</th> </tr> </thead> <tbody> <tr> <td>Controls</td> <td align="center">0.048</td> <td align="center">0.055 (0.046, 0.065)</td> </tr> <tr> <td>Cases (all)</td> <td align="center">0.042</td> <td align="center">0.058 (0.048, 0.070)</td> </tr> </tbody> </table> <p>$p > 0.4$ in both tests</p>		Median, all homes (n = 346)	Geometric mean (95% CI), homes with phthalate > detection limit (n = 290)	Controls	0.048	0.055 (0.046, 0.065)	Cases (all)	0.042	0.058 (0.048, 0.070)
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^aAdditional results for this study presented in asthma table.

aOR = adjusted odds ratio; IgE = immunoglobulin E; NHANES = National Health and Nutrition Examination Survey;
VOC = volatile organic compound

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2

Table 3-10. Evidence pertaining to DIBP and asthma/wheezing and hypersensitivity in humans

Reference and study design	Results																															
<p>Ait Bamai et al. (2014)^a (Japan) Population: Children (n = 122, ages <15 yrs) and adults (n = 374, ages ≥15 yrs) living in 148 detached dwellings in which at least 25 mg of dust was collected; 2006 follow-up of 2003 baseline survey. Outcome: Bronchial asthma assessed by self-administered questionnaire (positive response to: in the past 2 yrs have you been seen at a hospital for bronchial asthma?); parents completed questionnaires for inhabitants <6 yrs Exposure: Dust samples from floor and other surfaces DIBP (µg/g dust):</p> <table border="0" data-bbox="190 808 711 934"> <tr> <td></td> <td style="text-align: center;">Median</td> <td style="text-align: center;">75th percentile</td> </tr> <tr> <td>Floor dust (n = 148)</td> <td style="text-align: center;">2.4</td> <td style="text-align: center;">5.5</td> </tr> <tr> <td>Multi-surface dust (n = 120)</td> <td style="text-align: center;">1.9</td> <td style="text-align: center;">3.5</td> </tr> </table> <p>Analysis: Generalized linear mixed effects model, considering gender, age strata (<15, ≥15 yrs), smoking status (personal and environmental tobacco smoke), furry pets in home, signs of dampness, Der 1 (not defined by authors), other phthalates dust, airborne fungi, formaldehyde, total VOC, and building characteristics as potential covariates.</p>		Median	75 th percentile	Floor dust (n = 148)	2.4	5.5	Multi-surface dust (n = 120)	1.9	3.5	<p>OR (95% CI) for bronchial asthma by tertile of DIBP in floor dust (adjusted for adjusted for gender, age strata, smoking status, dampness index, furry pets inside the home, Der 1, and sum of other phthalate dusts)</p> <table border="0" data-bbox="711 472 1421 766"> <thead> <tr> <th></th> <th style="text-align: center;">Full sample</th> <th style="text-align: center;">Children</th> <th style="text-align: center;">Adults</th> </tr> </thead> <tbody> <tr> <td>1 (low)</td> <td style="text-align: center;">1.0 (referent)</td> <td style="text-align: center;">1.0 (referent)</td> <td style="text-align: center;">1.0 (referent)</td> </tr> <tr> <td>2</td> <td style="text-align: center;">2.25 (0.48, 10.57)</td> <td style="text-align: center;">4.37 (0.36, 53.6)</td> <td style="text-align: center;">1.16 (0.16, 8.17)</td> </tr> <tr> <td>3 (high)</td> <td style="text-align: center;">5.09 (1.17, 22.15)</td> <td style="text-align: center;">8.94 (0.86, 93.0)</td> <td style="text-align: center;">2.90 (0.52, 16.2)</td> </tr> <tr> <td>(trend <i>p</i>-value)</td> <td style="text-align: center;">(0.03)</td> <td style="text-align: center;">(0.067)</td> <td style="text-align: center;">(0.22)</td> </tr> </tbody> </table> <p><i>p</i>-value for age interaction = 0.51</p> <p>No significantly increased aORs (either in the full sample or stratified by age) were observed in analyses of bronchial asthma using DIBP measurements in multisurface dust.</p>				Full sample	Children	Adults	1 (low)	1.0 (referent)	1.0 (referent)	1.0 (referent)	2	2.25 (0.48, 10.57)	4.37 (0.36, 53.6)	1.16 (0.16, 8.17)	3 (high)	5.09 (1.17, 22.15)	8.94 (0.86, 93.0)	2.90 (0.52, 16.2)	(trend <i>p</i> -value)	(0.03)	(0.067)	(0.22)
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<p>(Callesen et al. (2014a); Callesen et al. (2014b))^a (Denmark)</p> <p>Population: 72 asthma cases, 242 healthy controls group from population-based survey (Indoor Environment and Children’s Health); ages 3–5 yrs</p> <p>Outcome: Clinical exam and parent interview; asthma: recurrence of at least two of the three symptoms: cough, wheeze, and shortness of breath within the previous 12 mo (symptoms other than those triggered by respiratory infections); and doctor diagnosis of asthma in combination with ongoing treatment; 47% of asthma cases were IgE positive based on 20 allergen tests</p> <p>Exposure: DIBP concentrations in dust samples from bedroom and day care centers; total DIBP exposure estimated as a weighted mass fraction</p> <p>DIBP in dust among controls (µg/g):</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> </tr> <tr> <td>Home</td> <td align="center">27.0</td> </tr> <tr> <td>Day care</td> <td align="center">22.6</td> </tr> <tr> <td>Area-weighted</td> <td align="center">27.2</td> </tr> </table> <p>(weighted by assumed hrs in each environment)</p> <p>Analysis: Mann-Whitney U-test</p> <p>Related study: Callesen et al. (2014b) (same study population, with exposure measured in urine sample from participants MIBP in urine: median 74.2 ng/mL (controls)</p>		Median	Home	27.0	Day care	22.6	Area-weighted	27.2	<p>Median DIBP in dust (µg/g), by case-control status assessed by clinical examination</p> <table border="0"> <tr> <td></td> <td align="center">Controls (n = 242)</td> <td align="center">Asthma (n = 72)</td> </tr> <tr> <td>Home</td> <td align="center">27.0</td> <td align="center">25.8</td> </tr> <tr> <td>Day care</td> <td align="center">22.6</td> <td align="center">21.5</td> </tr> <tr> <td>Area-weighted</td> <td align="center">27.2</td> <td align="center">25.7</td> </tr> </table> <p>Similar results when based on case status defined by parent-questionnaire data (n = 110 asthma cases)</p> <p>Results from Callesen et al. (2014b): OR (95% CI) by quartile of MIBP (urine sample), adjusting for sex, breastfeeding <3 mo, smoking in the home, and single allergic predisposition Bronchial asthma (60 cases, 216 controls)</p> <table border="0"> <tr> <td>1</td> <td align="center">1.0 (referent)</td> </tr> <tr> <td>2</td> <td align="center">0.49 (0.22, 1.09)</td> </tr> <tr> <td>3</td> <td align="center">0.91 (0.41, 1.69)</td> </tr> <tr> <td>4</td> <td align="center">0.61 (0.27, 1.34)</td> </tr> </table>		Controls (n = 242)	Asthma (n = 72)	Home	27.0	25.8	Day care	22.6	21.5	Area-weighted	27.2	25.7	1	1.0 (referent)	2	0.49 (0.22, 1.09)	3	0.91 (0.41, 1.69)	4	0.61 (0.27, 1.34)
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<p>Bertelsen et al. (2013) (Norway)</p> <p>Population: 623 children from birth cohort (Environment and Childhood Asthma study), born 1992–1993; children with current asthma over-sampled (follow-up 2001–2004); ages 10 yrs</p> <p>Outcome: Current asthma (parental report of history of asthma plus ≥1 of the following: dyspnea, chest tightness, and/or wheezing in previous 12 mo; use of asthma medications in previous 12 mo; positive exercise challenge test)</p> <p>Exposure: First morning urine sample (child’s), collected at study examination</p> <p>MIBP in urine (µg/L)</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> <td align="center">75th</td> <td align="center">95th</td> </tr> <tr> <td>Unadjusted</td> <td align="center">49.2</td> <td align="center">88.4</td> <td align="center">231.0</td> </tr> <tr> <td>SG-adjusted</td> <td align="center">50.1</td> <td align="center">90.5</td> <td align="center">239.6</td> </tr> </table> <p>Analysis: Logistic regression, adjusting for variables shown in the results column</p>		Median	75 th	95 th	Unadjusted	49.2	88.4	231.0	SG-adjusted	50.1	90.5	239.6	<p>OR (95% CI) for current asthma by quartile of MIBP (µg/L) (adjusted for urine specific gravity, sex, parental asthma, and household income)</p> <table border="0"> <tr> <td>1: ≤31.4 (referent)</td> <td align="center">1 (referent)</td> </tr> <tr> <td>2: >31.4–49.2</td> <td align="center">1.3 (0.74, 2.4)</td> </tr> <tr> <td>3: >49.2–88.4</td> <td align="center">1.4 (0.73, 2.5)</td> </tr> <tr> <td>4: >88.4</td> <td align="center">1.5 (0.80, 2.7)</td> </tr> </table> <p>Increase in odds of current asthma per log₁₀ IQR MIBP (95% CI) = 1.1 (0.87, 1.5)</p>	1: ≤31.4 (referent)	1 (referent)	2: >31.4–49.2	1.3 (0.74, 2.4)	3: >49.2–88.4	1.4 (0.73, 2.5)	4: >88.4	1.5 (0.80, 2.7)								
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Reference and study design	Results																												
<p>Hoppin et al. (2013)^a (United States, NHANES) Population: 2,325 participants in population-based survey (NHANES), 2005–2006; ages ≥6 yrs Outcome: Self-administered questionnaire (asthma, wheeze in past yr) Exposure: Urine sample collected same day as serum sample Unadjusted MIBP in urine (µg/L):</p> <table border="0"> <tr> <td></td> <td></td> <td align="center" colspan="2">Percentile</td> </tr> <tr> <td></td> <td align="center">Median</td> <td align="center">75th</td> <td align="center">95th</td> </tr> <tr> <td>Children</td> <td align="center">8.93</td> <td align="center">16.38</td> <td align="center">45.97</td> </tr> <tr> <td>Adults</td> <td align="center">5.42</td> <td align="center">10.53</td> <td align="center">28.98</td> </tr> </table> <p>Analysis: Logistic regression, adjusting for variables shown in results column and sampling weights; separate analyses for children (ages 6–17 yrs) and adults (>17 yrs)</p>			Percentile			Median	75 th	95 th	Children	8.93	16.38	45.97	Adults	5.42	10.53	28.98	<p>Prevalence and OR (95% CI) for asthma symptoms per unit change in log-transformed urinary MIBP level (adjusted for age, race/ethnicity, gender, BMI, creatinine, and cotinine)</p> <p>Children (n = 779)</p> <table border="0"> <tr> <td>Asthma (n = 65)</td> <td align="center">8.4%</td> <td align="center">0.92 (0.26, 3.29)</td> </tr> <tr> <td>Wheeze (n = 80)</td> <td align="center">10.7%</td> <td align="center">1.08 (0.49, 2.35)</td> </tr> </table> <p>Adults (n = 1,546)</p> <table border="0"> <tr> <td>Asthma (n = 116)</td> <td align="center">7.4%</td> <td align="center">1.39 (0.77, 2.50)</td> </tr> <tr> <td>Wheeze (n = 219)</td> <td align="center">16.6%</td> <td align="center">0.92 (0.57, 1.48)</td> </tr> </table> <p>Authors reported that adjustment for poverty income ratio did not alter ORs.</p>	Asthma (n = 65)	8.4%	0.92 (0.26, 3.29)	Wheeze (n = 80)	10.7%	1.08 (0.49, 2.35)	Asthma (n = 116)	7.4%	1.39 (0.77, 2.50)	Wheeze (n = 219)	16.6%	0.92 (0.57, 1.48)
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<p>Sun et al. (2009)^a (China) Population: 92 cases asthma/wheezing, cases and 346 controls, all students of Tianjin University who had participated in a cross-sectional study of allergic symptoms and environmental factors; 2006–2007 Outcome: Self-reported symptoms from questionnaire; asthma/wheezing = in past 12 mos, have you had wheezing or whistling the in the chest; have you had dry cough at night for more than 2 wks, apart from a cough associated with a cold or chest infection; controls responded no to questions on asthma/wheeze, rhinitis, and eczema Exposure: Surface dust sample in dorm rooms DIBP in dust (µg/g):</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> <td align="center">75th percentile</td> </tr> <tr> <td></td> <td align="center">20.24</td> <td align="center">34.77</td> </tr> </table> <p>Analysis: Logistic regression for OR, considering age, gender, passive smoking, smoking, pet raising, atopy, and building age as potential covariates; Mann-Whitney U-test (nonparametric) for comparison between DIBP concentrations of cases and controls; t-test for comparisons between log transformed concentrations</p>		Median	75 th percentile		20.24	34.77	<p>OR for asthma comparing DIBP in dust (µg/g dust) above and below the median (adjusted to age, gender, smoking, atopy, and building age) reportedly did not reach statistical significance (quantitative results not reported)</p> <p>Median concentration DIBP in dust (µg/g dust)</p> <table border="0"> <tr> <td></td> <td align="center">Cases</td> <td align="center">Controls</td> </tr> <tr> <td>Wheezing</td> <td align="center">23.13</td> <td align="center">22.73</td> </tr> </table> <p>(<i>p</i> > 0.46 by Mann Whitney or t-test)</p>		Cases	Controls	Wheezing	23.13	22.73																
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- 1
- 2 ^aAdditional results for this study presented in allergy/immune table.
- 3
- 4 IQR = interquartile range

1 3.2.8. Neurodevelopmental Effects in Humans

2 Table 3-11. Evidence pertaining to DIBP and neurodevelopmental effects in
3 humans

Reference and study design	Results																																					
<i>Attention and executive function in school-aged children</i>																																						
<p>Kobrosly et al. (2014) (United States; Minnesota, Missouri, California, Iowa) Population: 153 children (n = 76 girls, n = 77 boys) from birth cohort study (Study for Future Families), born 2000–2005, ages 6–10 yrs in 2010 follow-up Outcome: Child Behavior Checklist (completed by parent) Exposure: Maternal urine sample, 3rd trimester (mean 26.6 wks) Unadjusted MIBP in urine (ng/mL): Geometric mean (95% CI) 2.3 (2.0, 2.8) Analysis: Linear regression, considering sex, age, mother’s education, urinary creatinine, family stress measure, and race/ethnicity, as potential covariates. Related references: Swan et al. (2005) (exposure data)</p>	<p>Regression coefficient (95% CI) for change in raw score on child behavior checklist per unit increase in ln-transformed MIBP (adjusted for sex, age, mother’s education and urinary creatinine, and family stress score)</p> <table border="1"> <thead> <tr> <th></th> <th>Boys</th> <th>Girls</th> </tr> </thead> <tbody> <tr> <td>Anxiety/depression</td> <td>0.11 (–0.13, 0.34)</td> <td>–0.03 (–0.29, 0.22)</td> </tr> <tr> <td>Withdrawn</td> <td>–0.01 (–0.21, 0.18)</td> <td>–0.04 (–0.25, 0.17)</td> </tr> <tr> <td>Somatic complaints</td> <td>–0.03 (–0.23, 0.16)</td> <td>–0.07 (–0.28, 0.13)</td> </tr> <tr> <td>Social problems</td> <td>0.18 (–0.02, 0.37)</td> <td>–0.06 (–0.27, 0.16)</td> </tr> <tr> <td>Thought problems</td> <td>0.15 (–0.05, 0.35)</td> <td>0.07 (–0.15, 0.29)</td> </tr> <tr> <td>Attention problems</td> <td>0.27 (0.04, 0.50)</td> <td>0.12 (–0.12, 0.36)</td> </tr> <tr> <td>Rule-breaking behavior *</td> <td>0.20 (0.01, 0.38)</td> <td>–0.04 (–0.23, 0.16)</td> </tr> <tr> <td>Aggressive behavior</td> <td>0.34 (0.09, 0.59)</td> <td>0.12 (–0.14, 0.39)</td> </tr> <tr> <td>Internalizing behavior</td> <td>0.09 (–0.18, 0.37)</td> <td>–0.07 (–0.37, 0.22)</td> </tr> <tr> <td>Externalizing behavior</td> <td>0.32 (0.06, 0.58)</td> <td>0.06 (–0.22, 0.34)</td> </tr> <tr> <td>Total problems</td> <td>0.42 (0.05, 0.80)</td> <td>0.07 (–0.33, 0.47)</td> </tr> </tbody> </table> <p>*Sex interaction <i>p</i>-value = 0.04; all other interaction <i>p</i>-values > 0.05</p>			Boys	Girls	Anxiety/depression	0.11 (–0.13, 0.34)	–0.03 (–0.29, 0.22)	Withdrawn	–0.01 (–0.21, 0.18)	–0.04 (–0.25, 0.17)	Somatic complaints	–0.03 (–0.23, 0.16)	–0.07 (–0.28, 0.13)	Social problems	0.18 (–0.02, 0.37)	–0.06 (–0.27, 0.16)	Thought problems	0.15 (–0.05, 0.35)	0.07 (–0.15, 0.29)	Attention problems	0.27 (0.04, 0.50)	0.12 (–0.12, 0.36)	Rule-breaking behavior *	0.20 (0.01, 0.38)	–0.04 (–0.23, 0.16)	Aggressive behavior	0.34 (0.09, 0.59)	0.12 (–0.14, 0.39)	Internalizing behavior	0.09 (–0.18, 0.37)	–0.07 (–0.37, 0.22)	Externalizing behavior	0.32 (0.06, 0.58)	0.06 (–0.22, 0.34)	Total problems	0.42 (0.05, 0.80)	0.07 (–0.33, 0.47)
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

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<p>Engel et al. (2010) (United States; New York City) Population: 177 children from original birth cohort studied by Engel et al. (2009), 54% boys, three follow-up exams at ages 4.5–5.5, 6–6.5, 7–9 yrs Outcome: Behavior assessed by maternal reporting on BRIEF and BASC-PRS Exposure: Maternal urine sample, 25–40 wks gestation*</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> <td align="center">75th percentile</td> </tr> <tr> <td>MIBP (µg/L)*</td> <td align="center">2.6</td> <td align="center">12.2</td> </tr> <tr> <td>Sum LMW (µM/L) (sum of MBP, MEP, MIBP, and MMP)</td> <td align="center">1.88</td> <td align="center">4.59</td> </tr> </table> <p>Analysis: Generalized linear regression model, adjusting for variables shown in results column; other variables (not specified) were considered Related references: Engel et al. (2009) (exposure data for n = 295 children in the cohort) *MIBP concentrations not reported in (Engel et al., 2010); values reported here are from an earlier analysis of this cohort described in Engel et al. (2009)</p>		Median	75 th percentile	MIBP (µg/L)*	2.6	12.2	Sum LMW (µM/L) (sum of MBP, MEP, MIBP, and MMP)	1.88	4.59	<p>Regression coefficient for change in behavioral score (BASC-PRS) per unit increase in ln-phthalate level (µM/L) in boys (adjusted for race, educational level and marital status of the primary caretaker, and urinary creatinine)</p> <table border="0"> <tr> <td></td> <td align="center">MIBP</td> <td align="center">Low molecular weight phthalate sum</td> </tr> <tr> <td colspan="3">Clinical scales (higher score = more problem behaviors)</td> </tr> <tr> <td>Aggression</td> <td align="center">-0.12</td> <td align="center">1.24*</td> </tr> <tr> <td>Anxiety</td> <td align="center">-0.25</td> <td align="center">0.78</td> </tr> <tr> <td>Attention problems</td> <td align="center">0.66</td> <td align="center">1.29*</td> </tr> <tr> <td>Atypicality</td> <td align="center">0.53</td> <td align="center">0.95</td> </tr> <tr> <td>Conduct problems</td> <td align="center">0.23</td> <td align="center">2.40*</td> </tr> <tr> <td>Depression</td> <td align="center">0.29</td> <td align="center">1.18*</td> </tr> <tr> <td>Hyperactivity</td> <td align="center">0.85</td> <td align="center">1.03</td> </tr> <tr> <td>Somatization</td> <td align="center">1.04</td> <td align="center">0.36</td> </tr> <tr> <td>Withdrawal</td> <td align="center">-0.01</td> <td align="center">0.46</td> </tr> <tr> <td colspan="3">Adaptive scales (lower score = more problem behaviors)</td> </tr> <tr> <td>Adaptability</td> <td align="center">-1.32*</td> <td align="center">-1.08*</td> </tr> <tr> <td>Leadership</td> <td align="center">-1.30</td> <td align="center">-0.88</td> </tr> <tr> <td>Social skills</td> <td align="center">-0.93</td> <td align="center">-1.04</td> </tr> <tr> <td colspan="3">Composite scales (higher score = more problem behaviors)</td> </tr> <tr> <td>Externalizing problems</td> <td align="center">0.33</td> <td align="center">1.75*</td> </tr> <tr> <td>Internalizing problems</td> <td align="center">0.46</td> <td align="center">0.99</td> </tr> <tr> <td>Adaptive skills</td> <td align="center">-1.17</td> <td align="center">-0.98</td> </tr> <tr> <td>Behavioral Symptoms Index</td> <td align="center">0.47</td> <td align="center">1.55*</td> </tr> </table> <p>Significant sex-phthalate interactions ($p \leq 0.05$) for aggression, conduct problems, hyperactivity, externalizing problems, and behavioral symptoms index, as reported by study authors.</p> <p>Regression coefficient for change in behavioral score (BRIEF scores; higher score = worse executive functioning) per unit increase in ln-phthalate level (µM/L) in boys and girls (adjusted for race, sex, educational level and marital status of the primary caretaker, and urinary creatinine)</p> <table border="0"> <tr> <td>Emotional control</td> <td align="center">0.09</td> <td align="center">1.33*</td> </tr> <tr> <td>Behavioral</td> <td align="center">0.30</td> <td align="center">1.13</td> </tr> </table>			MIBP	Low molecular weight phthalate sum	Clinical scales (higher score = more problem behaviors)			Aggression	-0.12	1.24*	Anxiety	-0.25	0.78	Attention problems	0.66	1.29*	Atypicality	0.53	0.95	Conduct problems	0.23	2.40*	Depression	0.29	1.18*	Hyperactivity	0.85	1.03	Somatization	1.04	0.36	Withdrawal	-0.01	0.46	Adaptive scales (lower score = more problem behaviors)			Adaptability	-1.32*	-1.08*	Leadership	-1.30	-0.88	Social skills	-0.93	-1.04	Composite scales (higher score = more problem behaviors)			Externalizing problems	0.33	1.75*	Internalizing problems	0.46	0.99	Adaptive skills	-1.17	-0.98	Behavioral Symptoms Index	0.47	1.55*	Emotional control	0.09	1.33*	Behavioral	0.30	1.13
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																				
	regulation index																				
	Initiate	0.83	0.81																		
	Working memory	1.11	1.03																		
	Plan/organize	0.76	1.02																		
	Metacognition index	0.70	1.05																		
	Global executive composite	0.56	1.23*																		
	* $p \leq 0.05$																				
	Study authors reported that there were few significant associations between phthalate concentration and behavior among girls (quantitative results not reported).																				
Neurobehavioral outcomes in infants and preschool-aged children																					
<p>Braun et al. (2014) (United States)</p> <p>Population: 175 children from birth cohort in Ohio (HOME cohort, recruited during pregnancy, 2003–2006); follow-up at ages 4–5 yrs</p> <p>Outcome: Autistic behaviors based on Social Responsiveness Scale completed by mother; 65 item scale, higher score = more autistic behaviors</p> <p>Exposure: Maternal urine samples, 16–26 wks of gestation</p> <p>MIBP in urine ($\mu\text{g/g Cr}$):</p> <table border="1"> <thead> <tr> <th></th> <th align="center" colspan="3">Percentile</th> </tr> <tr> <th></th> <th align="center">Median</th> <th align="center">75th</th> <th align="center">95th</th> </tr> </thead> <tbody> <tr> <td>Cr-adjusted</td> <td align="center">5.6</td> <td align="center">8.6</td> <td align="center">17</td> </tr> </tbody> </table> <p>Analysis: Semi-Bayesian hierarchical regression model</p>		Percentile				Median	75 th	95 th	Cr-adjusted	5.6	8.6	17	<p>Regression coefficient (95% CI) for change in total score per unit increase in log-transformed Cr-adjusted MIBP (adjusted for maternal demographic and perinatal factors, depressive symptoms, caregiving environment, and serum cotinine):</p> <p align="center">0.7 (–1.4, 2.8)</p>								
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	Median	75 th	95 th																		
Cr-adjusted	5.6	8.6	17																		
<p>Téllez-Rojo et al. (2013) (Mexico)</p> <p>Population: 135 children from birth cohort (Early Life Exposure in Mexico to Environmental Toxicants cohort; mothers recruited during first trimester, 1997–2003)</p> <p>Outcome: Mental and psychomotor development based on Bayley Scales of Infant Development-II (assessed by trained examiner, videotaped for quality control assessment) tested at 24, 30, and 36 mo of age</p> <p>Exposure: Maternal urine sample, 3rd trimester</p> <p>MIBP in urine (ng/mL):</p> <table border="1"> <thead> <tr> <th></th> <th align="center" colspan="3">Geometric mean (95% CI)</th> </tr> </thead> <tbody> <tr> <td>SG-adjusted</td> <td align="center" colspan="3">2.30 (1.92, 2.76)</td> </tr> </tbody> </table> <p>Analysis: Linear regression for longitudinal data, stratified by sex and adjusted for variables shown in results column</p> <p>Related reference: Ettinger et al. (2009)</p>		Geometric mean (95% CI)			SG-adjusted	2.30 (1.92, 2.76)			<p>Regression coefficient (95% CI) for change in neurodevelopment score per unit increase in maternal In-MIBP (adjusted for birthweight, breastfeeding practices, weight-for-age, child’s age, mother’s age, mother’s education, and laboratory)</p> <table border="1"> <thead> <tr> <th></th> <th align="center">Total sample (n = 135)</th> <th align="center">Boys (n = 64)</th> <th align="center">Girls (n = 71)</th> </tr> </thead> <tbody> <tr> <td>MDI</td> <td align="center">0.53 (–0.85, 1.91)</td> <td align="center">0.32 (–1.62, 2.28)</td> <td align="center">–0.12 (–1.94, 1.69)</td> </tr> <tr> <td>PDI</td> <td align="center">0.57 (–0.67, 1.82)</td> <td align="center">0.63 (–0.68, 1.95)</td> <td align="center">0.37 (–1.67, 2.43)</td> </tr> </tbody> </table>		Total sample (n = 135)	Boys (n = 64)	Girls (n = 71)	MDI	0.53 (–0.85, 1.91)	0.32 (–1.62, 2.28)	–0.12 (–1.94, 1.69)	PDI	0.57 (–0.67, 1.82)	0.63 (–0.68, 1.95)	0.37 (–1.67, 2.43)
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																																				
<p>Whyatt et al. (2012) (United States, New York City) Population: 297 children from birth cohort (Columbia Center for Children’s Environmental Health), born 1999–2006; 3-yr follow-up, mean age 36 mo (range 27–42 mo) Outcome: Mental, psychomotor and behavioral development at 3 yrs based on Bayley Scales of Infant Development-II (assessed by trained examiners) and Child Behavior Checklist (completed by parent) Exposure: Maternal urine sample, 3rd trimester MIBP in urine (ng/mL): Geometric mean Unadjusted 9.3 Analysis: Linear and logistic regression adjusting for variables shown in results column; Wald test used to detect sex differences</p>	<p>Regression coefficient (95% CI) for change in neurodevelopment score per unit increase in maternal ln-MIBP (adjusted for specific gravity, race/ethnicity, maternal marital status and prenatal alcohol consumption, child’s gestational age and sex, and quality of care-taking environment)</p> <table border="0"> <tr> <td></td> <td align="center">Boys (n = 140)</td> <td align="center">Girls (n = 157)</td> </tr> <tr> <td>MDI</td> <td align="center">0.59 (-1.40, 2.58)</td> <td align="center">-1.33 (-3.20, 0.54)</td> </tr> <tr> <td>PDI</td> <td align="center">-2.21 (-4.61, 0.19)</td> <td align="center">-2.33 (-4.59, -0.08)</td> </tr> </table> <p>OR (95% CI) for risk of mental or psychomotor delay (score ≤85) per ln-unit increase in maternal ln-MIBP (each model adjusted for one or more of the following: specific gravity, race/ethnicity, maternal marital status and prenatal alcohol consumption, child’s gestational age and sex, and quality of care-taking environment)</p> <table border="0"> <tr> <td></td> <td align="center">Boys (n = 140)</td> <td align="center">Girls (n = 157)</td> </tr> <tr> <td>MDI</td> <td align="center">0.87 (0.60, 1.28)</td> <td align="center">0.98 (0.62, 1.56)</td> </tr> <tr> <td>PDI</td> <td align="center">1.80 (1.13, 2.87)</td> <td align="center">1.98 (1.02, 3.83)</td> </tr> </table> <p>Regression coefficient (95% CI) for change in neurobehavior per unit increase in maternal ln-MIBP (adjusted for specific gravity; ethnicity; maternal IQ, demoralization, hardship, satisfaction during pregnancy and prenatal exposure to PAH and BPA; and child’s sex and age at testing)</p> <table border="0"> <tr> <td></td> <td align="center">Boys (n = 129)</td> <td align="center">Girls (n = 148)</td> </tr> <tr> <td>Emotionally reactive</td> <td align="center">0.42 (-0.005, 0.85)</td> <td align="center">0.34 (-0.11, 0.78)</td> </tr> <tr> <td>Anxious/depressed</td> <td align="center">0.12 (-0.38, 0.61)</td> <td align="center">0.16 (-0.34, 0.66)</td> </tr> <tr> <td>Somatic complaints</td> <td align="center">0.31 (-0.18, 0.81)</td> <td align="center">0.24 (-0.22, 0.70)</td> </tr> <tr> <td>Withdrawn behavior</td> <td align="center">0.36 (-0.05, 0.77)</td> <td align="center">0.47 (-0.007, 0.94)</td> </tr> <tr> <td>Internalizing behavior</td> <td align="center">1.21 (-0.16, 2.56)</td> <td align="center">1.20 (-0.15, 2.55)</td> </tr> </table> <p>No effect modification by gender was observed (<i>p</i>-values >0.7).</p> <p>OR (95% CI) for child’s score in the borderline or clinical range (compared to normal) per unit increase in maternal ln-MBP (adjusted for specific gravity, maternal</p>		Boys (n = 140)	Girls (n = 157)	MDI	0.59 (-1.40, 2.58)	-1.33 (-3.20, 0.54)	PDI	-2.21 (-4.61, 0.19)	-2.33 (-4.59, -0.08)		Boys (n = 140)	Girls (n = 157)	MDI	0.87 (0.60, 1.28)	0.98 (0.62, 1.56)	PDI	1.80 (1.13, 2.87)	1.98 (1.02, 3.83)		Boys (n = 129)	Girls (n = 148)	Emotionally reactive	0.42 (-0.005, 0.85)	0.34 (-0.11, 0.78)	Anxious/depressed	0.12 (-0.38, 0.61)	0.16 (-0.34, 0.66)	Somatic complaints	0.31 (-0.18, 0.81)	0.24 (-0.22, 0.70)	Withdrawn behavior	0.36 (-0.05, 0.77)	0.47 (-0.007, 0.94)	Internalizing behavior	1.21 (-0.16, 2.56)	1.20 (-0.15, 2.55)
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results		
	demoralization and satisfaction during pregnancy, and child's sex and age at testing)		
		Borderline Clinical	
	Somatic complaints	1.29 (0.84, 1.99)	0.76 (0.42, 1.36)
	Withdrawn behavior	0.81 (0.44, 1.51)	1.62 (0.97, 2.73)
Internalizing behavior	1.98 (1.24, 3.23)	1.41 (0.91, 2.18)	

- 1
- 2 BASC-PRS = Behavior Assessment System for Children—Parent Rating Scales; BPA = bisphenol A; BRIEF = Behavior
- 3 Rating Inventory of Executive Function; HOME = Health Outcomes and Measures of the Environment; LMW = low
- 4 molecular weight; MDI = mental delay index; MMP = monomethyl phthalate; PAH = polycyclic aromatic
- 5 hydrocarbon; PDI = psychomotor delay index

1 3.2.9. Thyroid Hormone Effects in Humans

2 Table 3-12. Evidence pertaining to DIBP and thyroid hormones in humans

Reference and study design	Results																																														
<p>Dirtu et al. (2013) (Belgium)</p> <p>Population: 152 overweight or obese adults from weight loss cohort (ENDORUP) seen at weight management clinic, 43 age- and sex-matched controls from hospital staff and other volunteers, enrolled 2009–2012; among obese/overweight group, 65 received bariatric surgery and 87 received standard diet and lifestyle counseling; follow-up 3, 6, and 12 mo</p> <p>Outcome: Serum thyroid hormone levels (details of blood collection were not reported)</p> <p>Exposure: Urine sample (24-hr) MIBP in urine (ng/mL):</p> <table border="1"> <thead> <tr> <th></th> <th>Median</th> <th colspan="2">Percentile</th> </tr> <tr> <th></th> <th></th> <th>75th</th> <th>90th</th> </tr> </thead> <tbody> <tr> <td>Controls</td> <td>65</td> <td>93</td> <td>133</td> </tr> <tr> <td>Obese (at baseline)</td> <td>58</td> <td>89</td> <td>129</td> </tr> </tbody> </table> <p>Analysis: Linear regression, adjusting for variables shown in results column</p>		Median	Percentile				75 th	90 th	Controls	65	93	133	Obese (at baseline)	58	89	129	<p>Regression coefficient (<i>p</i>-value) for change in hormone level with unit change in ln-MIBP (adjusted for age, weight loss, and sex, or stratified by sex) (0.0 = no effect)</p> <table border="1"> <thead> <tr> <th></th> <th>Full sample</th> <th>Men</th> <th>Women</th> </tr> </thead> <tbody> <tr> <td colspan="4">Overweight/obese group</td> </tr> <tr> <td>Free T4</td> <td>0.07 (0.41)</td> <td>0.11 (0.47)</td> <td>0.05 (0.66)</td> </tr> <tr> <td>TSH</td> <td>-0.01 (0.93)</td> <td>0.09 (0.58)</td> <td>-0.01 (0.94)</td> </tr> <tr> <td colspan="4">Referent group</td> </tr> <tr> <td>Free T4</td> <td>0.24 (0.14)</td> <td>0.49 (0.12)</td> <td>0.16 (0.44)</td> </tr> <tr> <td>TSH</td> <td>0.23 (0.16)</td> <td>-0.43 (0.19)</td> <td>0.32 (0.10)</td> </tr> </tbody> </table>				Full sample	Men	Women	Overweight/obese group				Free T4	0.07 (0.41)	0.11 (0.47)	0.05 (0.66)	TSH	-0.01 (0.93)	0.09 (0.58)	-0.01 (0.94)	Referent group				Free T4	0.24 (0.14)	0.49 (0.12)	0.16 (0.44)	TSH	0.23 (0.16)	-0.43 (0.19)	0.32 (0.10)
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<p>Meeker and Ferguson (2011) (United States)</p> <p>Population: Participants in population-based survey (NHANES), 2007–2008; 1,346 ages ≥20 yrs and 329 adolescents ages 12–19 yrs</p> <p>Outcome: Serum thyroid hormone levels</p> <p>Exposure: Urine sample collected same day as serum sample Cr-adjusted MIBP in urine (µg/g Cr):</p> <table border="1"> <thead> <tr> <th></th> <th>Median</th> <th colspan="2">Percentile</th> </tr> <tr> <th></th> <th></th> <th>75th</th> <th>95th</th> </tr> </thead> <tbody> <tr> <td>Adults</td> <td>6.67</td> <td>11.1</td> <td>24.1</td> </tr> <tr> <td>Adolescents</td> <td>8.24</td> <td>13.73</td> <td>28.78</td> </tr> </tbody> </table> <p>Analysis: Linear regression adjusting for variables shown in results column.</p>		Median	Percentile				75 th	95 th	Adults	6.67	11.1	24.1	Adolescents	8.24	13.73	28.78	<p>Regression coefficient (95% CI) for change in hormone level with unit increase in ln-MIBP (adjusted for age, sex, race, BMI, ln-serum cotinine, ln-urinary creatinine, and ln-urinary iodine, and weighted for sampling strategy)</p> <table border="1"> <thead> <tr> <th></th> <th>Adults</th> <th>Adolescents</th> </tr> </thead> <tbody> <tr> <td>Total T3 (ng/dL)</td> <td>0.77 (-0.59, 2.12)</td> <td>2.30 (-0.81, 0.52)</td> </tr> <tr> <td>Ln (Free T3) (pg/mL)</td> <td>-0.0012 (-0.0074, 0.0051)</td> <td>0.0083 (-0.0062, 0.023)</td> </tr> <tr> <td>Total T4 (µg/mL)</td> <td>0.020 (-0.075, 0.11)</td> <td>-0.034 (-0.25, 0.19)</td> </tr> <tr> <td>Ln (Free T4) (ng/dL)</td> <td>0.0010 (-0.0094, 0.011)</td> <td>-0.0001 (-0.021, 0.021)</td> </tr> <tr> <td>Ln (TSH) (µIU/mL)</td> <td>-0.013 (-0.054, 0.028)</td> <td>0.003 (-0.076, 0.081)</td> </tr> <tr> <td>Ln (Tg) (ng/mL)</td> <td>-0.018 (-0.081, 0.045)</td> <td>-0.047 (-0.12, 0.074)</td> </tr> </tbody> </table>				Adults	Adolescents	Total T3 (ng/dL)	0.77 (-0.59, 2.12)	2.30 (-0.81, 0.52)	Ln (Free T3) (pg/mL)	-0.0012 (-0.0074, 0.0051)	0.0083 (-0.0062, 0.023)	Total T4 (µg/mL)	0.020 (-0.075, 0.11)	-0.034 (-0.25, 0.19)	Ln (Free T4) (ng/dL)	0.0010 (-0.0094, 0.011)	-0.0001 (-0.021, 0.021)	Ln (TSH) (µIU/mL)	-0.013 (-0.054, 0.028)	0.003 (-0.076, 0.081)	Ln (Tg) (ng/mL)	-0.018 (-0.081, 0.045)	-0.047 (-0.12, 0.074)							
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3
4
5 T3 = triiodothyronine; T4 = thyroxine; TSH = thyroid stimulating hormone

1 3.2.10. Obesity and Metabolic Effects in Humans

2 Table 3-13. Evidence pertaining to DIBP and obesity in humans

Reference and study design	Results																								
<p>Buser et al. (2014) (United States, NHANES) Population: Participants in population-based survey (NHANES), 2007–2010 ages ≥6 yrs [sample size not reported] Outcome: BMI measured at exam; divided into obese (BMI z-score ≥95th percentile in children, BMI ≥30 in adults) and overweight (BMI z-score 85th–95th percentiles in children, BMI 25–29.9 in adults). Exposure: Urine sample, collected at same time as exam Unadjusted MIBP in urine (ng/mL) Geometric mean (SE) Ages 6–19 yrs 10.43 (0.39) Ages ≥20 yrs 6.75 (0.23) Analysis: Logistic regression, considering age, race/ethnicity, sex, urinary creatinine, poverty income ratio, calorie intake, and serum cotinine as potential covariates in analyses of ages 6–19 yrs; or age, race/ethnicity, sex, education, diabetes, alcohol consumption, cigarette smoking, calorie intake, vigorous recreational activities, urinary creatinine, and serum cotinine as potential covariates in analysis of ages ≥20 yrs</p>	<p>OR (95% CI) in children (6–19 yrs of age) for obesity or overweight comparing highest quartile urinary MIBP (>20.84 ng/mL) with lowest quartile (≤5.38 ng/mL) (adjusted for age, race/ethnicity, calorie intake, serum cotinine, urinary creatinine, income level)</p> <table border="1" data-bbox="675 499 1427 688"> <thead> <tr> <th></th> <th>Obese</th> <th>Overweight</th> </tr> </thead> <tbody> <tr> <td>All</td> <td>1.82 (0.73, 4.57)</td> <td>1.85 (0.78, 4.40)</td> </tr> <tr> <td>Boys</td> <td>4.26 (1.32, 13.74)</td> <td>2.22 (0.78, 6.28)</td> </tr> <tr> <td>Girls</td> <td>0.57 (0.18, 1.83)</td> <td>1.57 (0.58, 4.25)</td> </tr> </tbody> </table> <p>OR (95% CI) in adults (≥20 yrs of age) for obesity or overweight comparing highest quartile urinary MIBP (>14.40 ng/mL) with lowest quartile (≤3.49 ng/mL) (adjusted for age, gender, race/ethnicity, calorie intake, recreational activity, serum cotinine, education level, smoking status, alcohol intake, diabetes)</p> <table border="1" data-bbox="675 877 1427 1066"> <thead> <tr> <th></th> <th>Obese</th> <th>Overweight</th> </tr> </thead> <tbody> <tr> <td>All</td> <td>1.40 (0.90, 2.16)</td> <td>1.18 (0.79, 1.78)</td> </tr> <tr> <td>Men</td> <td>0.98 (0.57, 1.67)</td> <td>1.06 (0.60, 1.89)</td> </tr> <tr> <td>Women</td> <td>1.81 (0.94, 3.48)</td> <td>1.25 (0.66, 2.36)</td> </tr> </tbody> </table>		Obese	Overweight	All	1.82 (0.73, 4.57)	1.85 (0.78, 4.40)	Boys	4.26 (1.32, 13.74)	2.22 (0.78, 6.28)	Girls	0.57 (0.18, 1.83)	1.57 (0.58, 4.25)		Obese	Overweight	All	1.40 (0.90, 2.16)	1.18 (0.79, 1.78)	Men	0.98 (0.57, 1.67)	1.06 (0.60, 1.89)	Women	1.81 (0.94, 3.48)	1.25 (0.66, 2.36)
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<p>Hart et al. (2013) (Australia) Population: 121 girls from birth cohort study (Western Australian Pregnancy Cohort), whose mothers were recruited at 18 wks of gestation between 1989 and 1991; follow-up at ages 14–16 yrs Outcome: Offspring BMI (height and weight measured at clinic visit on d 2–5 of menstrual cycle) Exposure: Maternal serum samples (n = 123) collected at 18 and 34–36 wks of gestation (combined aliquot from both time periods) MIBP in serum (ng/mL): Median 90th percentile Unadjusted 1.77 6.16 Analysis: Correlation between log-transformed MIBP and BMI</p>	<p>Authors reported no association between adolescent BMI (either as absolute value or as age- and gender-adjusted z-score) and any phthalate metabolite in maternal serum (r = -0.10–0.04, p = 0.345–0.931)</p>																								

Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

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<p>Trasande et al. (2013a) (United States, NHANES) Population: 2,884 participants in population-based survey (NHANES), 2003–2008; 6–19 yrs old Outcome: BMI z-score, obesity (BMI z-score $\geq 95^{\text{th}}$ percentile), and overweight (BMI z-score $\geq 85^{\text{th}}$ percentile) (measured) Exposure: Urine sample, collected at same time as BMI measurement ΣLMW phthalates in urine (μM): Geometric mean Not obese 0.701 Obese 0.855 ΣLMW phthalates = sum of MEP, MBP, MIBP, and MCP Analysis: Logistic regression for overweight and obese classification; linear regression of BMI z-score as continuous variable; adjusted for variables shown in results column</p>	<p>Full sample results, no association with In-LMW phthalates: OR or regression coefficient (95% CI) per one unit increase in ΣLMW phthalates (μM) (adjusted for urinary creatinine, sex, poverty-income ratio, parental education, serum cotinine, age, and race/ethnicity, caloric intake, and television watching)</p> <table> <tr> <td>Overweight</td> <td>OR (95% CI)</td> <td>1.01 (0.90, 1.13)</td> </tr> <tr> <td>Obese</td> <td>OR (95% CI)</td> <td>1.02 (0.90, 1.17)</td> </tr> <tr> <td>BMI z-score</td> <td>β (95% CI)</td> <td>0.03 (–0.03, 0.09)</td> </tr> </table> <p>Interaction by ethnicity seen, with associations seen between In-LMW phthalates and each of the obesity measures in blacks, but not in whites or Hispanics. The patterns seen with ΣLMW phthalates were also seen in analyses for MIBP. Using same adjustment factors as above, the associations with In-MIBP are:</p> <table> <thead> <tr> <th rowspan="2"></th> <th colspan="2">ΣLMW phthalates</th> <th colspan="2">MIBP</th> </tr> <tr> <th>Hispanic</th> <th>White</th> <th>Black</th> <th>Black</th> </tr> </thead> <tbody> <tr> <td>Over-weight OR (95% CI)</td> <td>0.88 (0.72, 1.08)</td> <td>0.97 (0.78, 1.22)</td> <td>1.21 (1.05, 1.39)</td> <td>1.16 (0.99, 1.37)</td> </tr> <tr> <td>Obese OR (95% CI)</td> <td>0.97 (0.83, 1.14)</td> <td>0.94 (0.69, 1.29)</td> <td>1.22 (1.07, 1.39)</td> <td>1.17 (0.97, 1.41)</td> </tr> <tr> <td>BMI z-score β (95% CI)</td> <td>–0.04 (–0.15, 0.06)</td> <td>0.02 (–0.08, 0.12)</td> <td>0.09 (0.003, 0.18)</td> <td>0.08 (–0.01, 0.17)</td> </tr> </tbody> </table>	Overweight	OR (95% CI)	1.01 (0.90, 1.13)	Obese	OR (95% CI)	1.02 (0.90, 1.17)	BMI z-score	β (95% CI)	0.03 (–0.03, 0.09)		Σ LMW phthalates		MIBP		Hispanic	White	Black	Black	Over-weight OR (95% CI)	0.88 (0.72, 1.08)	0.97 (0.78, 1.22)	1.21 (1.05, 1.39)	1.16 (0.99, 1.37)	Obese OR (95% CI)	0.97 (0.83, 1.14)	0.94 (0.69, 1.29)	1.22 (1.07, 1.39)	1.17 (0.97, 1.41)	BMI z-score β (95% CI)	–0.04 (–0.15, 0.06)	0.02 (–0.08, 0.12)	0.09 (0.003, 0.18)	0.08 (–0.01, 0.17)
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<p>Wang et al. (2013) (China) Population: 259 primary and middle school students, 8–15 yrs old, stratified sample from six schools, selected based on sex and BMI Outcome: BMI, waist circumference (measured) Exposure: First morning urine sample, collected at same time as BMI measurement MIBP in urine (ng/mL): Geometric mean (SD) 38.9 (1.1) Low molecular weight phthalate metabolites included MMP, MEP, MBP, MIBP, and MHBP Analysis: Linear regression, sampling weights applied to adjust for sampling strategy; adjusted for variables shown in the results column</p>	<p>Regression coefficient (95% CI) for change in BMI or waist circumference per unit increase in SG-adjusted InMIBP (adjusted for age and sex in Model 1; plus sum of DBP, MMP, and MEP in Model 2)</p> <table> <thead> <tr> <th></th> <th>Model 1</th> <th>Model 2</th> </tr> </thead> <tbody> <tr> <td>BMI</td> <td>0.027 (0.006, 0.048)</td> <td>0.020 (–0.005, 0.045)</td> </tr> <tr> <td>Waist circumference</td> <td>0.022(0.005, 0.038)</td> <td>0.019 (–0.001, 0.038)</td> </tr> </tbody> </table>		Model 1	Model 2	BMI	0.027 (0.006, 0.048)	0.020 (–0.005, 0.045)	Waist circumference	0.022(0.005, 0.038)	0.019 (–0.001, 0.038)																								
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Waist circumference	0.022(0.005, 0.038)	0.019 (–0.001, 0.038)																																

Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																												
<p>Dirtu et al. (2013) (Belgium) Population: 152 overweight or obese adults from weight loss cohort (ENDORUP) seen at weight management clinic, 43 age- and sex-matched controls from hospital staff and other volunteers, enrolled 2009–2012; among obese/overweight group, 65 received bariatric surgery and 87 received standard diet and lifestyle counseling; follow-up 3, 6, and 12 mo Outcome: Waist circumference measured at each follow-up visit Exposure: Urine sample (24-hr sample) MIBP, in urine (ng/mL):</p> <table border="0"> <tr> <td></td> <td></td> <td align="center" colspan="2">Percentile</td> </tr> <tr> <td></td> <td align="center">Median</td> <td align="center">75th</td> <td align="center">90th</td> </tr> <tr> <td>Controls</td> <td align="center">65</td> <td align="center">93</td> <td align="center">133</td> </tr> <tr> <td>Obese</td> <td align="center">58</td> <td align="center">89</td> <td align="center">129</td> </tr> </table> <p>(at baseline) Analysis: Linear regression, adjusting for variables shown in results column; treatment of repeated urinary phthalate measures was not specified</p>			Percentile			Median	75 th	90 th	Controls	65	93	133	Obese	58	89	129	<p>Regression coefficient (<i>p</i>-value) for change in waist circumference with unit change in ln-MIBP (adjusted for age, weight loss, and sex, or stratified by sex) (0.0 = no effect)</p> <table border="0"> <tr> <td></td> <td align="center">Full sample</td> <td align="center">Men</td> <td align="center">Women</td> </tr> <tr> <td>Overweight/ obese group</td> <td align="center">0.07 (0.40)</td> <td align="center">-0.16 (0.30)</td> <td align="center">0.03 (0.76)</td> </tr> <tr> <td>Referent group</td> <td align="center">-0.16 (0.30)</td> <td align="center">0.07 (0.81)</td> <td align="center">-0.01 (0.98)</td> </tr> </table>		Full sample	Men	Women	Overweight/ obese group	0.07 (0.40)	-0.16 (0.30)	0.03 (0.76)	Referent group	-0.16 (0.30)	0.07 (0.81)	-0.01 (0.98)
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<p>Lind et al. (2012b) (Sweden) Population: 1,016 (507 men, 509 women), from population-based cohort (Prospective Investigation of Vasculature in Uppsala Seniors study), 2001–2003; age 70 yrs at enrollment Outcome: BMI, waist circumference measured at enrollment; DXA (n = 890 participated) and MRI of abdominal region (n = 287 randomly selected) 2 yrs later Exposure: Serum sample (fasting), collected at baseline MIBP in serum (ng/mL):</p> <table border="0"> <tr> <td></td> <td align="center">Median</td> <td align="center">75th percentile</td> </tr> <tr> <td>Women</td> <td align="center">13.4</td> <td align="center">24.5</td> </tr> <tr> <td>Men</td> <td align="center">13.5</td> <td align="center">33.3</td> </tr> </table> <p>Analysis: Linear regression, adjusted for variables shown in results column Related reference: Olsén et al. (2012) reports cross-sectional analysis of BMI from this study population, see Table 14</p>		Median	75 th percentile	Women	13.4	24.5	Men	13.5	33.3	<p>Regression coefficient (95% CI) for change in body metric per unit increase in ln-MIBP (ng/mL) (adjusted for serum cholesterol and triglycerides, education, exercise, and smoking)</p> <table border="0"> <tr> <td></td> <td align="center">Males</td> <td align="center">Females</td> </tr> <tr> <td align="center">Outcome</td> <td align="center">β (95% CI)</td> <td align="center">β (95% CI)</td> </tr> <tr> <td>BMI (kg/m²)</td> <td align="center">-0.083 (-0.35, 0.19)</td> <td align="center">0.39 (0.002, 0.79)</td> </tr> <tr> <td>Waist circumference (cm)</td> <td align="center">-0.025 (-0.80, 0.75)</td> <td align="center">1.3 (0.425, 2.3)</td> </tr> <tr> <td>DXA total fat (kg)</td> <td align="center">-73 (-754, 608)</td> <td align="center">1,079 (283, 1875)</td> </tr> <tr> <td>MRI visceral adipose tissue (cm²)</td> <td align="center">-5.9 (-24, 13)</td> <td align="center">14 (1.4, 26)</td> </tr> </table>		Males	Females	Outcome	β (95% CI)	β (95% CI)	BMI (kg/m ²)	-0.083 (-0.35, 0.19)	0.39 (0.002, 0.79)	Waist circumference (cm)	-0.025 (-0.80, 0.75)	1.3 (0.425, 2.3)	DXA total fat (kg)	-73 (-754, 608)	1,079 (283, 1875)	MRI visceral adipose tissue (cm ²)	-5.9 (-24, 13)	14 (1.4, 26)	
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Reference and study design	Results													
<p>Teitelbaum et al. (2012) (United States, New York City) Population: 387 children (80 boys, 307 girls) in child development cohort (Growing Up Healthy Study), 2004–2008; Hispanic and black), 6–8 yrs at enrollment Outcome: BMI and waist circumference measured 1 yr after enrollment; normal weight = BMI <85th percentile (n = 2,284); overweight = BMI ≥85th percentile (n = 578) Exposure: Urine sample, collected at enrollment Cr-adjusted phthalates in urine (µg/g Cr), median:</p> <table border="0"> <tr> <td></td> <td align="center">MIBP</td> <td align="center">ΣLMW phthalates</td> </tr> <tr> <td>Boys</td> <td align="center">22.7</td> <td align="center">253.2</td> </tr> <tr> <td>Girls</td> <td align="center">22.2</td> <td align="center">294.0</td> </tr> </table> <p>Low molecular weight phthalate metabolites included MEP, MBP, MIBP, and MCP. Analysis: Linear regression, considering sex, age at baseline, sedentary hrs, metabolic equivalent hrs, caloric intake, race, ethnicity, season of urine collection, family income, and parent education as potential covariates; restricted to children with creatinine ≥10 mg/dL</p>		MIBP	ΣLMW phthalates	Boys	22.7	253.2	Girls	22.2	294.0	<p>Full sample results, regression coefficient (95% CI) for change in body metric per unit change in ln-MIBP (µg/g Cr) (adjusted for creatinine, age, sex, sedentary hrs, metabolic equivalent hrs, Hispanic ethnicity, caloric intake, season, and parental education level)</p> <table border="0"> <tr> <td>BMI (kg/m²)</td> <td align="right">-0.27 (-0.73, -0.18)</td> </tr> <tr> <td>Waist circumference (cm)</td> <td align="right">-0.62 (-1.84, -0.61)</td> </tr> </table>	BMI (kg/m ²)	-0.27 (-0.73, -0.18)	Waist circumference (cm)	-0.62 (-1.84, -0.61)
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<p>Olsén et al. (2012) (Sweden) Population: 1,016 (507 men, 509 women), from population-based cohort (Prospective Investigation of Vasculature in Uppsala Seniors study), 2001–2003; age 70 yrs at enrollment Outcome: BMI measured at study visit Exposure: Serum sample, collected at time of examination; results not shown Analysis: Linear regression, adjusted for the variables shown in results column</p>	<p>Regression coefficient for change in outcome per unit increase in ln-MIBP (adjusted for sex, smoking, diabetes (except for glucose) and the other variables in the table; model for Framingham Risk Score only adjusted for sex)</p> <table border="0"> <tr> <td>BMI</td> <td align="right">0.094 (-0.13, 0.32)</td> </tr> </table>	BMI	0.094 (-0.13, 0.32)											
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Reference and study design	Results																
<p>Kasper-Sonnenberg et al. (2012) (Germany) Population: 104 mothers (and children) enrolled in birth cohort study, children born between 2000 and 2002, follow-up in 2007–2009; mean age 39.2 yrs (mothers), 6.8 yrs (children) Outcome: BMI based on questionnaire (mothers) and measurements (children) Exposure: Urine sample (first morning), collected on same day as exam Cr-adjusted MIBP and OH-MIBP in urine (µg/g Cr):</p> <p align="center">Geometric mean (95% CI)</p> <p>Children</p> <table border="0"> <tr><td>MIBP</td><td>64.6 (55.2, 75.7)</td></tr> <tr><td>OH-MIBP</td><td>34.2 (28.5, 41.0)</td></tr> <tr><td>ΣDIBP</td><td>101 (87.2, 118)</td></tr> </table> <p>Adults</p> <table border="0"> <tr><td>MIBP</td><td>37.2 (31.8, 43.5)</td></tr> <tr><td>OH-MIBP</td><td>17.4 (15.1, 20.0)</td></tr> <tr><td>ΣDIBP</td><td>55.9 (48.4, 64.5)</td></tr> </table> <p>Analysis: Spearman’s rank correlation analysis</p>	MIBP	64.6 (55.2, 75.7)	OH-MIBP	34.2 (28.5, 41.0)	ΣDIBP	101 (87.2, 118)	MIBP	37.2 (31.8, 43.5)	OH-MIBP	17.4 (15.1, 20.0)	ΣDIBP	55.9 (48.4, 64.5)	<p>Spearman correlation coefficient between ΣDIBP and BMI in</p> <table border="0"> <tr><td>Children</td><td align="right">-0.035 ($p > 0.05$)</td></tr> <tr><td>Mothers</td><td align="right">-0.137 ($p > 0.05$)</td></tr> </table>	Children	-0.035 ($p > 0.05$)	Mothers	-0.137 ($p > 0.05$)
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<p>Svensson et al. (2011) (Mexico) Population: 182 women; healthy controls without diabetes from case-control study of breast cancer, 2007–2008; mean age 54 yrs Outcome: BMI, waist circumference, and waist:height ratio Exposure: First morning urine sample collected at time of clinical evaluation Cr-adjusted MIBP in urine (µg/g Cr):</p> <p align="center">Geometric mean (SD)</p> <table border="0"> <tr><td>No diabetes</td><td>9.1 (2.3)</td></tr> </table> <p>Analysis: Spearman correlation coefficient Related references: Lopez-Carrillo et al. (2010)</p>	No diabetes	9.1 (2.3)	<p>Spearman correlation coefficient between anthropometric measure and ln-MIBP in urine (µg/g Cr)</p> <table border="0"> <tr><td>BMI (kg/m²)</td><td align="right">0.0457</td></tr> <tr><td>Waist circumference (cm)</td><td align="right">0.0151</td></tr> <tr><td>Waist/height ratio</td><td align="right">-0.0156</td></tr> </table> <p>($p > 0.05$ for all parameters)</p>	BMI (kg/m ²)	0.0457	Waist circumference (cm)	0.0151	Waist/height ratio	-0.0156								
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1
2
3
4

DXA = dual energy x-ray absorptiometry; MCPPE = mono-(3-carboxypropyl) phthalate; MHBP = mono-(3-hydroxybutyl)phthalate; MRI= magnetic resonance imaging; SE = standard error

1
2

Table 3-14. Evidence pertaining to DIBP and diabetes/insulin resistance in humans

Reference and study design	Results										
<i>Diabetes diagnosis</i>											
<p>James-Todd et al. (2012) (United States, NHANES) Population: 215 cases, 1,235 controls from population-based survey (NHANES), 2001–2008; women age 20–79 yrs Outcome: Positive response to, “Other than during pregnancy, have you ever been told by a doctor or health professional that you have diabetes or sugar diabetes?” Exposure: Urine sample, collected at time of survey MIBP in urine (units not reported): Geometric mean Unadjusted 3.7 (based on larger sample of 2,350 women) Analysis: Logistic regression, adjusting for variables shown in the results column</p>	<p>OR (95% CI) for diabetes by quartile of MIBP (adjusted for urinary creatinine, age, race/ethnicity, education, poverty status, fasting time, total caloric intake, total fat intake, smoking status, and physical activity; little change with additional adjustment for BMI and waist circumference)</p> <p>MIBP quartile</p> <table border="0"> <tr> <td>1 (low)</td> <td>1.0 (referent)</td> </tr> <tr> <td>2</td> <td>1.04 (0.66–1.67)</td> </tr> <tr> <td>3</td> <td>1.69 (0.93–3.06)</td> </tr> <tr> <td>4 (high)</td> <td>1.95 (0.99–3.85)</td> </tr> </table>	1 (low)	1.0 (referent)	2	1.04 (0.66–1.67)	3	1.69 (0.93–3.06)	4 (high)	1.95 (0.99–3.85)		
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<p>Lind et al. (2012a) (Sweden) Population: 1,003 (501 men, 502 women), from population-based cohort (Prospective Investigation of Vasculature in Uppsala Seniors study), 2001–2003; age 70 yrs at enrollment Outcome: Diabetes (n = 88; history of diabetes or fasting glucose >7.0 mmol/L, mean duration 8.9 yrs); Exposure: Serum sample (fasting), collected at time of clinical assessment MIBP in serum (ng/mL): Median 75th percentile Women 13.4 24.5 Men 13.5 33.3 Analysis: Logistic regression for diabetes classification, adjusting for variables shown in results column</p>	<p>OR (95% CI) per unit increase in serum ln-MIBP (adjusted for sex, serum cholesterol and triglycerides, BMI, smoking, exercise, and education)</p> <p>1.30 (1.10, 1.55)</p> <p>OR (95% CI) by quintile of ln-MIBP (adjusted for sex, serum cholesterol and triglycerides, BMI, smoking, exercise, and education)</p> <p>MIBP quintile</p> <table border="0"> <tr> <td>1 (low)</td> <td>1.0 (referent)</td> </tr> <tr> <td>2</td> <td>1.19 (0.59, 2.38)</td> </tr> <tr> <td>3</td> <td>0.84 (0.41, 1.76)</td> </tr> <tr> <td>4</td> <td>1.37 (0.7, 2.66)</td> </tr> <tr> <td>5 (high)</td> <td>2.00 (1.03, 3.99)</td> </tr> </table> <p>(trend <i>p</i>) (0.038)</p>	1 (low)	1.0 (referent)	2	1.19 (0.59, 2.38)	3	0.84 (0.41, 1.76)	4	1.37 (0.7, 2.66)	5 (high)	2.00 (1.03, 3.99)
1 (low)	1.0 (referent)										
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Reference and study design	Results																								
<p>Svensson et al. (2011) (Mexico) Population: 221 women with diabetes, 182 healthy without diabetes from case-control study of breast cancer, 2007–2008; mean age 54 yrs Outcome: Self-reported diabetes Exposure: First morning urine samples MIBP in urine (µg/g creatinine): Geometric mean (SD) No diabetes 9.1 (2.3) Diabetes 7.9 (2.1) Analysis: Logistic regression, adjusted for variables shown in the results column (age and waist-height ratio not found to be potential confounders)</p>	<p>OR (95% CI) per unit increase in ln-MIBP (adjusted for creatinine and education)</p> <p align="center">1.01 (0.65, 1.55)</p>																								
<i>Markers of insulin resistance</i>																									
<p>Huang et al. (2014a) (United States, NHANES) Population: 3,083 participants in population-based survey (NHANES), 2001–2008; ages 12–<80 yrs; self-reported non-diabetic, non-pregnant participants Outcome: Fasting blood glucose; fasting insulin; HOMA-IR Exposure: Urine sample at time of clinical exam Cr-adjusted MIBP in urine (µg/g Cr): Median 75th percentile Men 3.8 6.6 Women 4.9 8.9 Analysis: Logistic regression, adjusting for variables shown in the results column</p>	<p>Median change (95% CI) in biomarkers for diabetes by quartile of MIBP (adjusted for age, gender, race/ethnicity, fasting time, urinary creatinine, total caloric intake, triglycerides, education, and poverty and smoking status)</p> <table border="1"> <thead> <tr> <th>MIBP quartile</th> <th>Fasting glucose</th> <th>Fasting insulin</th> <th>HOMA-IR</th> </tr> </thead> <tbody> <tr> <td>1 (low)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> <td>1.0 (referent)</td> </tr> <tr> <td>2</td> <td>1.87 (0.83, 2.92)</td> <td>1.45 (0.85, 2.04)</td> <td>0.38 (0.23, 0.52)</td> </tr> <tr> <td>3</td> <td>2.77 (1.75, 3.80)</td> <td>1.23 (0.57, 1.89)</td> <td>0.35 (0.19, 0.51)</td> </tr> <tr> <td>4 (high)</td> <td>3.69 (2.60, 4.78)</td> <td>1.73 (0.92, 2.54)</td> <td>0.53 (0.33, 0.72)</td> </tr> <tr> <td>(p for trend)</td> <td>(<0.0001)</td> <td>(0.0028)</td> <td>(0.0002)</td> </tr> </tbody> </table>	MIBP quartile	Fasting glucose	Fasting insulin	HOMA-IR	1 (low)	1.0 (referent)	1.0 (referent)	1.0 (referent)	2	1.87 (0.83, 2.92)	1.45 (0.85, 2.04)	0.38 (0.23, 0.52)	3	2.77 (1.75, 3.80)	1.23 (0.57, 1.89)	0.35 (0.19, 0.51)	4 (high)	3.69 (2.60, 4.78)	1.73 (0.92, 2.54)	0.53 (0.33, 0.72)	(p for trend)	(<0.0001)	(0.0028)	(0.0002)
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<p>Trasande et al. (2013b) (United States, NHANES) Population: 766 participants in the 2003–2008 NHANES, 12–19 yrs old Outcome: HOMA, calculated as fasting glucose (mmol/L) multiplied by fasting insulin (µU/mL divided by 22.5). Exposure: Urine sample, collected at same time as insulin resistance measurements. ΣLMW phthalates in urine (µM): Median 75th percentile Unadjusted 0.83 1.89 ΣLMW phthalates = sum of MEP, MBP, and MIBP Urinary concentration of MIBP alone not reported. Analysis: HOMA-IR assessed as continuous or categorical variable; categorical analysis</p>	<p>OR (95% CI) for insulin resistance and ln-urinary metabolite concentration (µM), adjusted for urinary creatinine, BMI category, continuous age, race/ethnicity, caregiver education, poverty-income ratio, gender, serum cotinine, and caloric intake</p> <table border="1"> <tbody> <tr> <td>Ln-MIBP</td> <td>1.57 (1.18, 2.09)</td> </tr> <tr> <td>Ln-ΣLMW</td> <td>0.92 (0.71, 1.19)</td> </tr> </tbody> </table> <p>Regression coefficient (95% CI) for increase in ln-HOMA-IR per unit increase in ln-urinary metabolite concentration (µM), adjusted for urinary creatinine, BMI category, continuous age, race/ethnicity, caregiver education, poverty-income ratio, gender, serum cotinine, and caloric intake.</p> <table border="1"> <tbody> <tr> <td>Ln-MIBP</td> <td>0.15 (0.04, 0.26)</td> </tr> <tr> <td>Ln-ΣLMW</td> <td>-0.07 (-0.18, 0.04)</td> </tr> </tbody> </table>	Ln-MIBP	1.57 (1.18, 2.09)	Ln-ΣLMW	0.92 (0.71, 1.19)	Ln-MIBP	0.15 (0.04, 0.26)	Ln-ΣLMW	-0.07 (-0.18, 0.04)																
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Reference and study design	Results																																																		
<p>used cut point of 4.39, reflecting >2 SD above the mean HOMA-IR for normal weight adolescents with normal fasting glucose in NHANES 1999–2002. Linear and logistic regression analyses, adjusting for variables shown in results column. HOMA-IR and urinary phthalate measures natural-log transformed for analysis.</p>																																																			
<p>James-Todd et al. (2012) (United States, NHANES) Population: 2,092 women without history of diabetes with various measures of insulin resistance from population-based survey (NHANES), 2001–2008; women age 20–79 yrs Outcome: Among women without history of diabetes, FBG (n = 985), HOMA-IR (n = 971), glycosolated hemoglobin A1c (n = 2,092) Exposure: Urine sample, collected at time of survey MIBP in urine (units not reported): Geometric mean Unadjusted 3.7 Analysis: Logistic regression, adjusting for variables shown in the results column</p>	<p>Among women without diabetes, difference (from first quartile) in median value (95% CI) of glucose and insulin parameters by quartile of MIBP (Model 1 adjusted for urine creatinine, age, race/ethnicity, education level, poverty status, fasting time, total caloric intake, total fat intake, smoking status, and physical activity; Model 2 also adjusted for BMI and waist circumference)</p> <table border="1" data-bbox="675 709 1421 1457"> <thead> <tr> <th data-bbox="675 709 837 741">MIBP Quartile</th> <th data-bbox="1036 709 1127 741">Model 1</th> <th data-bbox="1279 709 1370 741">Model 2</th> </tr> </thead> <tbody> <tr> <td data-bbox="675 741 837 793">FBG (mg/dL)</td> <td colspan="2"></td> </tr> <tr> <td data-bbox="675 793 837 825">1 (low)</td> <td data-bbox="1036 793 1127 825">(referent)</td> <td data-bbox="1279 793 1370 825">(referent)</td> </tr> <tr> <td data-bbox="675 825 837 856">2</td> <td data-bbox="997 825 1166 856">3.08 (1.22, 4.93)</td> <td data-bbox="1240 825 1409 856">3.03 (1.05, 5.00)</td> </tr> <tr> <td data-bbox="675 856 837 888">3</td> <td data-bbox="997 856 1166 888">3.50 (1.45, 5.54)</td> <td data-bbox="1240 856 1409 888">3.17 (1.17, 5.17)</td> </tr> <tr> <td data-bbox="675 888 837 919">4 (high)</td> <td data-bbox="997 888 1166 919">5.86 (3.55, 8.17)</td> <td data-bbox="1240 888 1409 919">6.04 (3.81, 8.28)</td> </tr> <tr> <td data-bbox="675 919 837 972">Ln (HOMA)</td> <td colspan="2"></td> </tr> <tr> <td data-bbox="675 972 837 1003">1 (low)</td> <td data-bbox="1036 972 1127 1003">(referent)</td> <td data-bbox="1279 972 1370 1003">(referent)</td> </tr> <tr> <td data-bbox="675 1003 837 1035">2</td> <td data-bbox="997 1003 1166 1035">0.13 (-0.02, 0.28)</td> <td data-bbox="1240 1003 1409 1035">0.13 (0.01, 0.25)</td> </tr> <tr> <td data-bbox="675 1035 837 1066">3</td> <td data-bbox="997 1035 1166 1066">0.08 (-0.08, 0.25)</td> <td data-bbox="1240 1035 1409 1066">0.10 (-0.01, 0.21)</td> </tr> <tr> <td data-bbox="675 1066 837 1098">4 (high)</td> <td data-bbox="997 1066 1166 1098">0.22 (0.06, 0.38)</td> <td data-bbox="1240 1066 1409 1098">0.18 (0.06, 0.31)</td> </tr> <tr> <td data-bbox="675 1098 837 1150">A1c (%)</td> <td colspan="2"></td> </tr> <tr> <td data-bbox="675 1150 837 1182">1 (low)</td> <td data-bbox="1036 1150 1127 1182">(referent)</td> <td data-bbox="1279 1150 1370 1182">(referent)</td> </tr> <tr> <td data-bbox="675 1182 837 1213">2</td> <td data-bbox="997 1182 1166 1213">0.03 (-0.01, 0.08)</td> <td data-bbox="1240 1182 1409 1213">0.03 (-0.01, 0.08)</td> </tr> <tr> <td data-bbox="675 1213 837 1245">3</td> <td data-bbox="997 1213 1166 1245">0.03 (-0.02, 0.09)</td> <td data-bbox="1240 1213 1409 1245">0.04 (0.00, 0.09)</td> </tr> <tr> <td data-bbox="675 1245 837 1276">4 (high)</td> <td data-bbox="997 1245 1166 1276">0.01 (-0.05, 0.07)</td> <td data-bbox="1240 1245 1409 1276">0.01 (-0.04, 0.07)</td> </tr> </tbody> </table>			MIBP Quartile	Model 1	Model 2	FBG (mg/dL)			1 (low)	(referent)	(referent)	2	3.08 (1.22, 4.93)	3.03 (1.05, 5.00)	3	3.50 (1.45, 5.54)	3.17 (1.17, 5.17)	4 (high)	5.86 (3.55, 8.17)	6.04 (3.81, 8.28)	Ln (HOMA)			1 (low)	(referent)	(referent)	2	0.13 (-0.02, 0.28)	0.13 (0.01, 0.25)	3	0.08 (-0.08, 0.25)	0.10 (-0.01, 0.21)	4 (high)	0.22 (0.06, 0.38)	0.18 (0.06, 0.31)	A1c (%)			1 (low)	(referent)	(referent)	2	0.03 (-0.01, 0.08)	0.03 (-0.01, 0.08)	3	0.03 (-0.02, 0.09)	0.04 (0.00, 0.09)	4 (high)	0.01 (-0.05, 0.07)	0.01 (-0.04, 0.07)
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Table 3-15. Evidence pertaining to DIBP and cardiovascular disease risk factors in humans

Reference and study design	Results																												
<p>Shiue (2014) (United States, NHANES) Population: 2,489 participants in population-based survey (NHANES), 2011–2012; ages ≥20 yrs Outcome: High blood pressure (systolic blood pressure ≥140 mmHg and diastolic blood pressure ≥90 mmHg) Exposure: Urine sample collected at time of clinical exam MIBP in urine (units not given) Mean ± SD Normal BP 13.13 ± 22.17 High BP 15.71 ± 25.15 Analysis: Survey-weighted logistic regression, adjusting for variables shown in results column; t-test for comparison between concentrations</p>	<p>OR (95% CI) for high blood pressure with increased log-transformed MIBP (adjusted for urinary creatinine, age, sex, ethnicity, BMI, and sampling weights)</p> <p style="text-align: center;">1.14 (0.92, 1.41)</p> <p>Mean ± SD MIBP in urine (units not given) in participants with normal and high BP</p> <table border="0" style="width: 100%;"> <tr> <td style="width: 60%;">Normal BP (n = 2,180)</td> <td style="width: 40%; text-align: right;">13.13 ± 22.17</td> </tr> <tr> <td>High BP (n = 309)</td> <td style="text-align: right;">15.71 ± 25.15</td> </tr> </table>		Normal BP (n = 2,180)	13.13 ± 22.17	High BP (n = 309)	15.71 ± 25.15																							
Normal BP (n = 2,180)	13.13 ± 22.17																												
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<p>Trasande et al. (2013c) (United States, NHANES) Population: 2,447 children in population-based survey (NHANES), 2003–2008; ages 8–19 yrs old Outcome: Systolic BP and diastolic BP z-score (based on height-, sex-, and age-normalized values); prehypertension (BP ≥90th percentile for age/height/sex); fasting serum triglycerides (n = 906; high = ≥100 mg/dL); nonfasting high density cholesterol (HDL; n = 2,555; low = <40 mg/dL) Exposure: Urine sample, collected at time of BMI measurement ΣLMW phthalates in urine (µM): Geometric mean BP <90th percentile 0.817 BP ≥90th percentile 1.002 ΣLMW phthalate = sum of MEP, MBP, and MIBP Analysis: Logistic regression for prehypertension (BP ≥90th percentile) classification; linear regression for systolic BP and diastolic BP z-score and triglycerides and HDL as continuous variable; all models adjusted for variables shown in results column</p>	<p>Changes in z-score (95% CI) per unit increase in ln-phthalates (adjusted for sex, caloric intake, television watching, poverty:income, parental education, serum cotinine, urinary creatinine, BMI, race/ethnicity, and age)</p> <table border="0" style="width: 100%; text-align: center;"> <tr> <td></td> <td>ΣLMW phthalates</td> <td>MIBP</td> </tr> <tr> <td>Systolic BP</td> <td>0.03 (–0.02, 0.07)</td> <td>0.03 (–0.02, 0.08)</td> </tr> <tr> <td>Diastolic BP</td> <td>0.02 (–0.04, 0.07)</td> <td>–0.02 (–0.09, 0.04)</td> </tr> <tr> <td>Triglycerides</td> <td>–0.22 (–4.40, 0.07)</td> <td>not reported</td> </tr> <tr> <td>HDL</td> <td>0.13 (–0.60, 0.85)</td> <td>not reported</td> </tr> </table> <p>OR (95% CI) for BP ≥90th percentile per unit increase in ln-phthalates</p> <table border="0" style="width: 100%; text-align: center;"> <tr> <td></td> <td>ΣLMW phthalates</td> <td>MIBP</td> </tr> <tr> <td>BP ≥90th percentile</td> <td>1.19 (0.96, 1.47)</td> <td>1.00 (0.74, 1.35)</td> </tr> <tr> <td>High triglycerides</td> <td>0.85 (0.71, 1.01)</td> <td>not reported</td> </tr> <tr> <td>Low HDL</td> <td>1.00 (0.87, 1.15)</td> <td>not reported</td> </tr> </table> <p>Interactions with covariates examined in supplemental analyses; stratified analyses showed no statistically significant associations between ΣLMW phthalates and systolic BP for gender, age, race/ethnicity, cotinine level, or BMI</p>			ΣLMW phthalates	MIBP	Systolic BP	0.03 (–0.02, 0.07)	0.03 (–0.02, 0.08)	Diastolic BP	0.02 (–0.04, 0.07)	–0.02 (–0.09, 0.04)	Triglycerides	–0.22 (–4.40, 0.07)	not reported	HDL	0.13 (–0.60, 0.85)	not reported		ΣLMW phthalates	MIBP	BP ≥90 th percentile	1.19 (0.96, 1.47)	1.00 (0.74, 1.35)	High triglycerides	0.85 (0.71, 1.01)	not reported	Low HDL	1.00 (0.87, 1.15)	not reported
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results																																																																			
<p>Olsén et al. (2012) (Sweden) Population: 1,016 (507 men, 509 women), from population-based cohort (Prospective Investigation of Vasculature in Uppsala Seniors study), 2001–2003; age 70 yrs at enrollment Outcome: Blood pressure measured at study visit; fasting serum sample for LDL and HDL cholesterol, and triglycerides; Framingham risk score Exposure: Serum sample, collected at time of examination; results not shown Analysis: Linear regression, adjusted for the variables shown in results column</p>	<p>Regression coefficient for change in outcome per unit increase in ln-MIBP (adjusted for sex, smoking, diabetes and the other variables in the table; model for Framingham Risk Score only adjusted for sex)</p> <p align="right">(β [SE])</p> <table> <tr> <td>LDL</td> <td align="right">0.044 (–0.01, 0.09)</td> </tr> <tr> <td>HDL</td> <td align="right">0.017 (–0.01, 0.09)</td> </tr> <tr> <td>Triglycerides</td> <td align="right">–0.009 (–0.03, 0.01)</td> </tr> <tr> <td>Systolic BP</td> <td align="right">–0.05 (–1.28, 1.18)</td> </tr> <tr> <td>Diastolic BP</td> <td align="right">0.35 (–0.20, 0.90)</td> </tr> <tr> <td>Framingham risk score</td> <td align="right">0.13 (–0.05, 0.31)</td> </tr> </table>	LDL	0.044 (–0.01, 0.09)	HDL	0.017 (–0.01, 0.09)	Triglycerides	–0.009 (–0.03, 0.01)	Systolic BP	–0.05 (–1.28, 1.18)	Diastolic BP	0.35 (–0.20, 0.90)	Framingham risk score	0.13 (–0.05, 0.31)																																																							
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<p>Lind and Lind (2011) (Sweden) Population: 1,016 (507 men, 509 women), from population-based cohort (Prospective Investigation of Vasculature in Uppsala Seniors study), 2001–2003; age 70 yrs at enrollment Outcome: Carotid artery intima media thickness (IMT); grey scale media of the intima media complex (IM-GSM); plaque in carotid artery Exposure: Serum sample (fasting), collected at time of clinical assessment MIBP in serum (ng/mL): Median 75th percentile 13.5 29.3 Analysis: Linear regression for continuous outcomes (IMT, IM-GSM) and ordinal logistic regression for number of carotid arteries with plaques (0, 1, 2), adjusted for variables shown in results column</p>	<p>Median IMT by quintile of MIBP (adjusted for sex, BMI, fasting blood glucose, systolic BP, diastolic BP, HDL and LDL cholesterol, triglycerides, smoking, antihypertensive treatment, statin use)</p> <table> <thead> <tr> <th rowspan="2">MIBP quintile</th> <th colspan="2">IMT</th> <th colspan="2">IM-GSM</th> </tr> <tr> <th>Median IMT</th> <th>(p-value)</th> <th>Median IM-GSM</th> <th>(p-value)</th> </tr> </thead> <tbody> <tr> <td>1 (low)</td> <td>0.87</td> <td>(referent)</td> <td>80</td> <td>Referent</td> </tr> <tr> <td>2</td> <td>0.89</td> <td>(0.91)</td> <td>72</td> <td>(0.0001)</td> </tr> <tr> <td>3</td> <td>0.86</td> <td>(0.13)</td> <td>68</td> <td>(0.0001)</td> </tr> <tr> <td>4</td> <td>0.89</td> <td>(0.91)</td> <td>69</td> <td>(0.0001)</td> </tr> <tr> <td>5 (high)</td> <td>0.85</td> <td>(0.074)</td> <td>102</td> <td>(0.0001)</td> </tr> </tbody> </table> <p>Regression coefficient (β [p-value]) per unit increase in serum MIBP (adjusted for sex, BMI, fasting blood glucose, systolic BP, diastolic BP, HDL and LDL cholesterol, triglycerides, smoking, antihypertensive treatment, statin use)</p> <table> <tr> <td>IMT</td> <td align="right">–0.0045 (0.14)</td> </tr> <tr> <td>IM-GSM</td> <td align="right">5.5 (0.0001)</td> </tr> </table> <p>OR for presence of plaques and median value of plaque GSM by quintile of MIBP (adjusted for sex, BMI, fasting blood glucose, systolic BP, diastolic BP, HDL and LDL cholesterol, triglycerides, smoking, antihypertensive treatment, statin use)</p> <table> <thead> <tr> <th rowspan="2">MIBP quintile</th> <th colspan="2">Plaque prevalence</th> <th colspan="2">Plaque GSM</th> </tr> <tr> <th>OR</th> <th>(p-value)</th> <th>Median</th> <th>(p-value)</th> </tr> </thead> <tbody> <tr> <td>1 (low)</td> <td>1.0</td> <td>(referent)</td> <td>65</td> <td>(referent)</td> </tr> <tr> <td>2</td> <td>0.70</td> <td>(0.059)</td> <td>69</td> <td>(0.37)</td> </tr> <tr> <td>3</td> <td>0.74</td> <td>(0.17)</td> <td>59</td> <td>(0.11)</td> </tr> <tr> <td>4</td> <td>1.00</td> <td>(0.78)</td> <td>62</td> <td>(0.074)</td> </tr> </tbody> </table>	MIBP quintile	IMT		IM-GSM		Median IMT	(p-value)	Median IM-GSM	(p-value)	1 (low)	0.87	(referent)	80	Referent	2	0.89	(0.91)	72	(0.0001)	3	0.86	(0.13)	68	(0.0001)	4	0.89	(0.91)	69	(0.0001)	5 (high)	0.85	(0.074)	102	(0.0001)	IMT	–0.0045 (0.14)	IM-GSM	5.5 (0.0001)	MIBP quintile	Plaque prevalence		Plaque GSM		OR	(p-value)	Median	(p-value)	1 (low)	1.0	(referent)	65	(referent)	2	0.70	(0.059)	69	(0.37)	3	0.74	(0.17)	59	(0.11)	4	1.00	(0.78)	62	(0.074)
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Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results				
	5 (high)	0.64	(0.011)	99	(0.0001)
	OR or regression coefficient per unit increase in serum MIBP				
	Plaque prevalence	OR (95% CI)		0.88 (0.79, 0.98)	
	Plaque GSM	β (<i>p</i> -value)		8.0 (0.0001)	
	The regression models did not show evidence of interaction by gender, except for IMT (interaction term <i>p</i> -value = 0.030).				

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BP = blood pressure; HDL = high-density lipoprotein; IM-GSM = grey scale media of the intima media complex;
IMT = intima media thickness; LDL = low-density lipoprotein

1 **3.3. EXPERIMENTAL STUDIES**

2 **3.3.1. Developmental Effects**

3 **Table 3-17. Evidence pertaining to developmental effects in animals following**
 4 **oral exposure to DIBP**

Reference and study design	Results ^a					
<i>Fetal survival</i>						
See Table 3-19						
<i>Fetal growth</i>						
BASF (2007)	Fetal body weight (percent change compared to control)					
Rat (Wistar); 22–23 dams/group	Doses	0	88	363	942	
0, 88, 363, 942 mg/kg-day	<i>M BW</i>	0%	–3%	–3%	–5%**	
Diet	<i>F BW</i>	0%	–3%	–3%	–6%**	
GDs 6–20 (GD 20 c-section)						
Borch et al. (2006)	Fetal body weight (percent change compared to control)					
Rat (Wistar); 11–12 dams/group	Doses (M)	0				600
0, 600 mg/kg-day	<i>BW (GD 19)</i> <i>(data presented in graph^b)</i>	0%				–27%*
Gavage	<i>BW (GD 20/21)</i> <i>(data presented in graph^b)</i>	0%				–12%
GDs 7–19 (GD 19 c-section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time point	Doses (F)	0				600
	<i>BW (GD 19)</i> <i>(data presented in graph^b)</i>	0%				–28%*
	<i>BW (GD 20/21)</i> <i>(data presented in graph^b)</i>	0%				12%
Saillenfait et al. (2006)	Fetal body weight (mean percent change compared to control)					
Rat (Sprague-Dawley); 20–22 dams/group	Doses	0	250	500	750	1,000
0, 250, 500, 750, 1,000 mg/kg-day	<i>M and F (all fetuses) BW</i>	0%	0%	–7%**	–17%**	–24%**
Gavage	<i>M BW</i>	0%	0%	–6%*	–17%**	–25%**
GDs 6–20 (GD 21 c-section)	<i>F BW</i>	0%	–1%	–8%**	–18%**	–26%**

Preliminary Materials for the IRIS Toxicological Review of Diisobutyl Phthalate

Reference and study design	Results ^a					
<i>Postnatal survival</i>						
Saillenfait et al. (2008)	Pup survival (percent change compared to control [litter means])					
Rat (Sprague-Dawley); 11–14 dams/group	Doses	0	125	250	500	625
0, 125, 250, 500, 625 mg/kg-day	<i>percentage pup survival PNDs 1–4</i>	0%	–1%	–1%	–1%	–7%
GDs 12–21 (dams allowed to deliver)	<i>percentage pup survival PNDs 4–21</i>	0%	2%	5%	3%	5%
<i>Postnatal and adult growth</i>						
Eastman Kodak (1954)	Body weight gain (percent change compared to control)					
Rat (no strain designation); 5 male and 5 females/group	Doses (M)	0	97	1,000	7,800	
	<i>BW gain (weaning to 4 weeks post-weaning)</i>	0%	3%	–2%	–61%	
0, 0.1, 1, 5% DIBP (0, 97, 1,000, 7,800 mg/kg-day for males; 0, 110, 1,100, 6,400 mg/kg-day for females) ^c	Doses (F)	0	110	1,100	6,400	
	<i>BW gain (weaning to 4 weeks post-weaning)</i>	0%	–9%	1%	–34%	
	Body weight (percent change compared to control)					
Diet	Doses (M)	0	97	1,000	7,800	
Weaning to 8 weeks post-weaning	<i>4 weeks post-weaning BW</i>	0%	2%	–1%	–41%	
	Doses (F)	0	110	1,100	6,400	
	<i>4 weeks post-weaning BW</i>	0%	–5%	1%	–19%	
	Body weight gain (percent change compared to control)					
	Doses (M)	0	97	1,000	7,800	
	<i>BW gain (weaning to 8 weeks post-weaning)</i>	0%	3%	–3%	–58%	
	Doses (F)	0	110	1,100	6,400	
	<i>BW gain (weaning to 8 weeks post-weaning)</i>	0%	–11%	0%	–34%	
	Body weight (percent change compared to control)					
	Doses (M)	0	97	1,000	7,800	
	<i>8 weeks post-weaning BW</i>	0%	2%	–3%	–44%	
	Doses (F)	0	110	1,100	6,400	
	<i>8 weeks post-weaning BW</i>	0%	–7%	0%	–22%	
	Note: Statistical analysis not reported in study.					

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Reference and study design	Results ^a						
<p>(Hazleton Laboratories (1992), 1987); NIOSH (1983)</p> <p>Mouse (CD-1); 50 control females; 10 females/treated group (6.5–9 weeks old)</p> <p>0, 1,000, 1,795, 3,225, 5,790, 10,400 mg/kg-day</p> <p>Gavage</p> <p>8 days</p>	Body weight change (g)						
	Doses (F)	0	1,000	1,795	3,225	5,790	10,400
	<i>BW change (days 1–8 of study)</i>	0	0	1	0	1	1
	Body weight change (g)						
	Doses (F)	0	1,000	1,795	3,225	5,790	10,400
	<i>BW change (days 1–12 of study)</i>	1	1	1	0	1	1
	Body weight change (g)						
	Doses (F)	0	1,000	1,795	3,225	5,790	10,400
	<i>BW change (days 1–16 of study)</i>	1	1	1	1	1	1
	Note: Statistical analysis not reported in study.						
<p>Oishi and Hiraga (1980d)</p> <p>MIBP</p> <p>Rat (Wistar) (JCL); 10 males/group</p> <p>0, 2% in diet (0, 1,100 mg/kg-day)^d</p> <p>Diet</p> <p>1 week</p>	Body weight and weight gain (percent change compared to control)						
	Doses (M)	0				1,100	
	<i>BW at 6 weeks</i>	0%				–10%*	
	<i>BW gain (5–6 weeks^e)</i>	0%				–31%	
	Note: Statistical analysis was not performed on BW gain.						
<p>Oishi and Hiraga (1980a)</p> <p>Mouse (JCL:ICR); 10 males/group</p> <p>0, 2% (0, 2,100 mg/kg-day)^d</p> <p>Diet</p> <p>1 week</p>	Body weight and weight gain (percent change compared to control)						
	Doses (M)	0				2,100	
	<i>BW at 6 weeks</i>	0%				–13%*	
	<i>BW gain (5–6 weeks^e)</i>	0%				–54%	
	Note: Statistical analysis was not performed on BW gain.						
<p>Saillenfait et al. (2008)</p> <p>Rat (Sprague-Dawley); 11–14 dams/group</p> <p>0, 125, 250, 500, 625 mg/kg-day</p> <p>Gavage</p>	Body weight (percent change compared to control [litter means])						
	Doses	0	125	250	500	625	
	<i>M postnatal (PND 1) BW</i>	0%	–1%	–2%	–2%	–10%**	
	<i>M postnatal (PND 21) BW</i>	0%	–1%	–3%	–6%	–10%*	
	<i>F postnatal (PND 21) BW</i>	0%	–3%	–5%	–3%	–10%	

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Reference and study design	Results^a						
GDs 12–21 (dams allowed to deliver)	<i>M BW at day of PPS</i>	0%	-8%*	-5%*	7%	2%	
	<i>M adult (PNDs 77–84) BW</i>	0%	-6%	-4%	-7%*	-9%**	
<u>University of Rochester (1953)</u>	Body weight gain (percent change compared to control)						
Rat (Albino; no other strain designation); 5 males/group	Doses (M)	0	15	140	1,400	3,000	8,900
	<i>BW gain^e (weaning to 1 month post-weaning)</i>	0%	-22%	-19%	-22%	-27%	-51%
	Note: Statistical analysis was not performed on BW gain.						
0, 0.01, 0.1, 1, 2, 5% (0, 15, 140, 1,400, 3,000, 8,900 mg/kg-day) ^f	Body weight (percent change compared to control)						
	Doses (M)	0	15	140	1,400	3,000	8,900
Diet	<i>1 month post-weaning BW</i>	0%	-16%	-14%	-16%	-20%	-38%
Weaning to 1 month post-weaning	Note: Statistical analysis not reported in study.						
<u>University of Rochester (1953)</u>	Body weight gain (percent change compared to control)						
University of Rochester (1953) University of Rochester (1953) University of Rochester (1953)	Doses (M)	0	15	140	1,400	3,000	8,900
	<i>BW gain (weaning to PND 49 [~after PPS^d])</i>	0%	-20%	-18%	-21%	-27%	-49%
	Note: Statistical analysis was not performed on BW gain.						
	Body weight (percent change compared to control)						
	Doses (M)	0	15	140	1,400	3,000	8,900
	<i>PND 49 (~after PPS^d) BW</i>	0%	-14%	-13%	-15%	-19%	-35%
	Note: Statistical analysis not reported in study.						
<u>University of Rochester (1954)</u>	Body weight gain (percent change compared to control)						
Rat (Albino; no other strain designation); 5 males and 5 females/group	Doses (M)	0	65	710	5,800		
	<i>BW gain (weaning to 1 month post-weaning)^e</i>	0%	5%	-6%	-60%		
0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^h	Doses (F)	0	82	770	4,700		
	<i>BW gain (weaning to 1 month post-weaning)^e</i>	0%	-7%	3%	-27%		
Diet	Note: Statistical analysis was not performed on BW gain.						
Weaning to 4 months post-weaning	Body weight (percent change compared to control)						

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Reference and study design	Results ^a				
	Doses (M)	0	65	710	5,800
	<i>BW at 1 month post-weaning</i>	0%	4%	-4%	-42%
	Doses (F)	0	82	770	4,700
	<i>BW at 1 month post-weaning</i>	0%	-4%	2%	-16%
	Note: Statistical analysis not reported in study.				
	Body weight gain (percent change compared to control)				
	Doses (M)	0	65	710	5,800
	<i>BW gain (weaning to PND 49 [~after PPS^g])^e</i>	0%	3%	-2%	-61%
	Doses (F)	0 ^y	82	770	4,700
	<i>BW gain (weaning to PND 49 [~after VO^g])^e</i>	0%	-8%	3%	-33%
	Note: Statistical analysis was not performed on BW gain.				
	Body weight (percent change compared to control)				
	Doses (M)	0	65	710	5,800
	<i>BW at PND 49 (~after PPS^g)</i>	0%	2%	-1%	-41%
	Doses (F)	0	82	770	4,700
	<i>BW at PND 49 (~after VO^g)</i>	0%	-4%	1%	-18%
	Note: Statistical analysis not reported in study.				
	Body weight gain (percent change compared to control)				
	Doses (M)	0	65	710	5,800
	<i>BW gain (weaning to 4 months post-weaning)</i>	0%	5%	-11%	-53%
	Doses (F)	0 ^y	82	770	4,700
	<i>BW gain (weaning to 4 months post-weaning)</i>	0%	-1%	10%	-19%
	Note: Statistical analysis was not performed on BW gain.				
	Body weight (percent change compared to control)				
	Doses (M)	0	65	710	5,800
	<i>BW at 4 months post-weaning</i>	0%	4%	-9%	-43%
	Doses (F)	0	82	770	4,700

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Reference and study design	Results ^a					
	<i>BW at 4 months post-weaning</i>	0%	-1%	7%	-13%	
	Note: Statistical analysis not reported in study.					
Fetal morphological development						
Saillenfait et al. (2006) Rat (Sprague-Dawley) rats; 20–22 dams/group 0, 250, 500, 750, 1,000 mg/kg-day Gavage GDs 6–20 (GD 21 c-section)	Malformations					
	External malformations (incidence; number of affected fetuses [litters])					
	Doses	0	250	500	750	1,000
	total fetuses (litters) examined for external malformations	281 (22)	276 (21)	237 (21)	212 (21)	111 (18)
	<i>anasarca</i>	0	0	0	0	1 (1)
	<i>exophthalmos (unilateral) and absence of eyelids (bilateral)</i>	0	0	0	1 (1)	0
	<i>exencephaly</i>	0	0	0	2 (2)	0
	<i>meningoencephalocele</i>	0	0	0	3 (3)	3 (2)
	<i>microstomia</i>	0	0	0	0	1 (1)
	<i>ectopia cordis</i>	0	0	0	0	1 (1)
	<i>omphalocele</i>	0	0	0	0	1 (1)
	Combined total with external malformations (incidence [percent])					
	Doses	0	250	500	750	1,000
	<i>total number (%) fetuses with external malformations</i>	0	0	0	5 (2%)*	6 (5%)**
	<i>total number (%) litters with external malformations</i>	0	0	0	4 (19%)	4 (22%)
	<i>mean % fetuses with external malformations/litter</i>	0%	0%	0%	2%	4%
	Visceral malformations (incidence; number of affected fetuses [litters])					
	Doses	0	250	500	750	1,000
	total fetuses (litters) examined for visceral malformations	141 (22)	138 (21)	119 (21)	106 (21)	56 (18)
	<i>anophthalmia, uni- or bilateral</i>	0	0	0	6 (4)	4 (3)
<i>aorta and/or pulmonary artery transposed</i>	0	0	0	6 (5)	3 (3)	

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Reference and study design	Results ^a				
<i>diaphragmatic hernia</i>	0	2 (1)	2 (2)	2 (2)	1 (1)
<i>kidney and ureter, absent, uni- or bilateral</i>	0	0	0	1 (1)	3 (3)
<i>kidney, small, uni- or bilateral</i>	0	0	0	1 (1)	1 (1)
Combined total with visceral malformations (incidence [percent])					
Doses	0	250	500	750	1,000
<i>total number (%) fetuses with visceral malformations</i>	0	2 (1%)	2 (2%)	13 (12%)**	10 (18%)**
<i>total number (%) litters with visceral malformations</i>	0	1 (5%)	2 (10%)	8 (38%)**	8 (44%)**
<i>mean % fetuses with visceral malformations/litter</i>	0%	1%	2%	13%*	16%*
Skeletal malformations (incidence; number of affected fetuses [litters])					
Doses	0	250	500	750	1,000
<i>total number of fetuses (litters) examined for skeletal malformations</i>	140 (22)	138 (21)	118 (21)	106 (21)	55 (18)
<i>mandible, small</i>	0	0	0	0	1 (1)
<i>sternebrae, fused</i>	0	0	0	7 (6)	14 (9)**
<i>sternebrae, fused and scrambled</i>	0	0	0	5 (3)	12 (7)
<i>sternebrae, total</i>	0	0	0	12 (7)*	26 (13)**
<i>cleft sternum</i>	0	0	1 (1)	1 (1)	2 (2)
<i>sternebrae, checkerboard</i>	0	0	0	2 (2)	0
<i>ribs, fused</i>	0	0	0	0	2 (2)
<i>cervical arches, fused</i>	0	0	0	3 (3)	3 (3)
<i>thoracic or lumbar vertebral arches, fused</i>	0	0	1 (1)	2 (2)	2 (2)
<i>thoracic or lumbar vertebral centra, fused</i>	0	0	1 (1)	0	4 (3)
<i>thoracic or lumbar centrum, hemicentric</i>	0	0	1 (1)	4 (3)	3 (3)
<i>thoracic or lumbar vertebral centra, misaligned</i>	0	0	2 (2)	3 (2)	5 (4)

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Reference and study design	Results ^a				
	Combined total with skeletal malformations (incidence [percent])				
Doses	0	250	500	750	1,000
<i>total number (%) fetuses with skeletal malformations</i>	0	0	4 (3%)	18 (17%)**	34 (62%)**
<i>total number (%) litters with skeletal malformations</i>	0	0	4 (19%)	11 (52%)**	15 (83%)**
<i>mean % fetuses with skeletal malformations/litter</i>	0%	0%	3%	18%**	67%**
	Variations				
	External variations (incidence; number of affected fetuses [litters])				
Doses	0	250	500	750	1,000
total fetuses (litters) examined for external variations	281 (22)	276 (21)	237 (21)	212 (21)	111 (18)
<i>clubfoot</i>	0	2 (1)	0	0	0
<i>tail, curly</i>	0	0	0	1 (1)	0
<i>tail tip, haemorrhage</i>	0	1 (1)	0	0	0
	Visceral variations (incidence; number of affected fetuses [litters])				
Doses	0	250	500	750	1,000
total fetuses (litters) examined for visceral variations	141 (22)	138 (21)	119 (21)	106 (21)	56 (18)
<i>dilated cerebral ventricle, slight</i>	0	0	0	1 (1)	0
<i>dilated renal pelvis</i>	1 (1)	0	0	2 (2)	5(4)
<i>ureter (all)</i>	3 (3)	0	2 (2)	10 (8)	12 (8)*
<i>hydroureter</i>	0	0	0	4 (4)	6 (5)
<i>distended ureter</i>	3 (3)	0	2 (2)	6 (5)	6 (4)
<i>ovaries, displaced</i>	0	0	0	5 (4)	2 (2)
<i>testis, ectopic</i>	0	0	3 (2)	30 (16)**	30 (16)**
<i>degree of trans-abdominal testicular migration (mean)</i>	2.6	3.8	13.6**	42.2**	58.1**

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Reference and study design	Results ^a					
	Skeletal variations (<i>incidence; number of affected fetuses [litters]</i>)					
	Doses	0	250	500	750	1,000
	total fetuses (litters) examined for skeletal variations	140 (22)	138 (21)	118 (21)	106 (21)	55 (18)
	<i>parietals or supraoccipital, incomplete ossification</i>	0	0	0	3 (2)	1 (1)
	<i>hyoid, absent or incomplete ossification</i>	0	0	0	1 (1)	8 (7)
	<i>sternbrae, fused, 1st and 2nd only</i>	1 (1)	0	8 (4)	29 (11)**	5 (4)
	<i>sternbrae, bipartite</i>	0	1 (1)	2 (2)	7 (5)	4 (4)
	<i>sternbrae, incomplete ossification</i>	0	1 (1)	5 (5)	9 (6)	6 (5)
	<i>ribs, cervical, rudimentary</i>	0	0	2 (2)	12 (9)*	9 (6)
	<i>ribs, 14th, any supernumerary</i>	23 (11)	32 (14)	42 (18)	72 (20)**	52 (18)**
	<i>ribs, 14th, long supernumerary</i>	1 (1)	1 (1)	2 (2)	15 (9)*	9 (9)*
	<i>ribs, short or reduced ossification (unilateral)</i>	0	0	0	1 (1)	1 (1)
	<i>thoracic or lumbar vertebral centra, incomplete ossification</i>	3 (2)	8 (6)	7 (7)	18 (14)**	16 (8)*
	<i>vertebrae, 27 presacral</i>	0	0	0	0	2 (2)
	Note: A single fetus may be represented more than once in the individual variations.					
<u>BASF (2007)</u> ⁱ	Malformations					
Rat (Wistar); 25 dams/group	External malformations (<i>incidence; number of affected fetuses [litters]</i>)					
0, 88, 363, 942 mg/kg-day	Doses	0	88	363	942	
Diet	total fetuses (litters) examined for external malformations and variations	208 (23)	197 (22)	182 (22)	211 (23)	
GDs 6–20 (GD 20 c-section)	<i>malformed head</i>	0	1 (1)	0	0	
	<i>anophthalmia</i>	0	1 (1)	0	0	
	Combined total with external malformations (<i>incidence [percent]</i>)					
	Doses	0	88	363	942	
	<i>fetuses</i>	0	1 (1%)	0	0	

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Reference and study design	Results ^a			
<i>litters</i>	0	1 (5%)	0	0
Combined total with soft tissue malformations				
<i>No fetuses affected at any dose</i>				
Skeletal malformations (incidence; number of affected fetuses [litters])				
Doses	0	88	363	942
<i>total fetuses (litters) examined for skeletal malformations/ variations</i>	109 (23)	101 (22)	97 (22)	110 (23)
<i>severely malformed skull bones</i>	0	1 (1)	0	0
<i>shortened scapula (cartilage present)</i>	0	0	1 (1)	0
<i>malpositioned and bipartite sternebral (unchanged cartilage)</i>	1 (1)	1 (1)	0	0
<i>branched rib</i>	1 (1)	0	0	0
<i>misshapen humerus</i>	0	2 (2)	0	2 (2)
<i>shortened humerus</i>	0	0	2 (1)	0
Combined total with skeletal malformations (incidence [percent])				
<i>fetuses</i>	1 (1%)	4 (4%)	2 (2%)	2 (2%)
<i>litters</i>	1 (4%)	4 (18%)	1 (5%)	2 (9%)
Variations				
Combined total with external variations				
<i>No fetuses affected at any dose</i>				
Soft tissue variations (incidence; number of affected fetuses [litters])				
Doses	0	88	363	942
<i>total fetuses (litters) examined for external soft tissue malformations and variations</i>	99 (23)	96 (22)	85 (22)	101 (23)
<i>dilated renal pelvis</i>	10 (7)	7 (5)	9 (8)	7 (5)
<i>dilated ureter</i>	2 (2)	2 (1)	1 (1)	1 (1)
Combined total with soft tissue variations (incidence (percent))				
Doses	0	88	363	942
<i>fetuses</i>	10 (10%)	7 (7%)	9 (11%)	7 (7%)
<i>litters</i>	7 (30%)	5 (23%)	8 (36%)	5 (22%)

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Reference and study design	Results ^a			
	Skeletal variations (<i>incidence; number of affected fetuses [litters]</i>)			
Doses	0	88	363	942
<i>supraoccipital hole(s)</i>	31 (15)	23 (13)	15 (12)	38 (19)
<i>incomplete ossification of basisphenoid</i>	7 (3)	2 (2)	6 (5)	8 (4)
<i>incomplete ossification of interparietal (unchanged cartilage)</i>	24 (15)	11 (7)	22 (12)	13 (9)
<i>incomplete ossification of parietal (unchanged cartilage)</i>	16 (10)	13 (9)	22 (13)	13 (7)
<i>incomplete ossification of supraoccipital (unchanged cartilage)</i>	9 (7)	14 (11)	14 (12)	13 (6)
<i>incomplete ossification of skull (unchanged cartilage)</i>	5 (4)	2(1)	7 (4)	2 (2)
<i>incomplete ossification of hyoid (cartilage present)</i>	1 (1)	0	1 (1)	1 (1)
<i>incomplete ossification of cervical arch (cartilage present)</i>	1 (1)	0	0	0
<i>incomplete ossification of thoracic centrum (unchanged cartilage)</i>	0	3 (3)	3 (3)	0
<i>dumbbell ossification of thoracic centrum (unchanged cartilage)</i>	4 (3)	3 (3)	2 (2)	9 (7)
<i>dumbbell ossification of thoracic centrum (dumbbell-shaped cartilage of centrum)</i>	14 (9)	13 (9)	13 (13)	17 (14)
<i>bipartite ossification of thoracic centrum (dumbbell-shaped cartilage of centrum)</i>	2 (2)	2 (2)	0	1 (1)
<i>supernumerary thoracic vertebra</i>	1 (1)	2 (1)	1 (1)	3 (2)
<i>unossified thoracic centrum (dumbbell-shaped cartilage of centrum)</i>	1 (1)	0	0	0

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Reference and study design	Results ^a			
<i>dumbbell ossification of lumbar centrum (dumbbell-shaped cartilage of centrum)</i>	1 (1)	0	0	0
<i>incomplete ossification of lumbar arch (cartilage present)</i>	0	0	1 (1)	0
<i>misshapen sacral vertebra</i>	1 (1)	1 (1)	1 (1)	4 (4)
<i>fused sacral centrum and arch (unchanged cartilage)</i>	3 (2)	7 (3)	5 (4)	5 (3)
<i>incomplete ossification of sacral arch (cartilage present)</i>	5 (4)	2 (2)	0	0
<i>unossified sternebra (unchanged cartilage)</i>	4 (4)	11 (7)	1 (1)	6 (4)
<i>incomplete ossification of sternebra (unchanged cartilage)</i>	42 (17)	44 (17)	44 (19)	75 (22*)
<i>misshapen sternebra (unchanged cartilage)</i>	32 (19)	26 (17)	20 (12)	28 (16)
<i>unilateral ossification of sternebra (unchanged cartilage)</i>	0	0	0	4 (4)
<i>extra sternebra ossification site (unchanged cartilage)</i>	0	0	1 (1)	0
<i>bipartite ossification of sternebra (unchanged cartilage)</i>	1 (1)	0	0	2 (2)
<i>supernumerary rib (14th) (cartilage present)</i>	6 (5)	1 (1)	6 (6)	6 (5)
<i>supernumerary rib (14th) (cartilage not present)</i>	50 (15)	33 (15)	40 (17)	62 (22*)
<i>cervical rib (cartilage present)</i>	0	1 (1)	0	0
<i>cervical rib (cartilage not present)</i>	5 (5)	5 (4)	5 (3)	4 (4)
<i>wavy rib</i>	6 (5)	2 (2)	10 (4)	9 (9)
<i>incomplete ossification of pubis (cartilage present)</i>	0	1 (1)	0	0

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Reference and study design	Results ^a			
	Combined total with skeletal variations (incidence (percent))			
Doses	0	88	363	942
<i>fetuses</i>	101 (93%)	87 (86%)	85 (88%)	107 (97%)
<i>litters</i>	23 (100%)	22 (100%)	22 (100%)	23 (100%)
	Unclassified observations			
	Unclassified external observations (incidence; number of affected fetuses [litters])			
Doses	0	88	363	942
total fetuses (litters) examined for unclassified observations	208 (23)	197 (22)	182 (22)	211 (23)
<i>discolored amniotic fluid</i>	0	0	0	1 (1)
	Combined total with external unclassified observations (incidence [percent])			
Doses	0	88	363	942
<i>fetuses</i>	0	0	0	1 (1%)
<i>litters</i>	0	0	0	1 (4%)
	Combined total with unclassified soft tissue observations			
	<i>No fetuses affected at any dose</i>			
	Skeletal unclassified cartilage observations (incidence; number of affected fetuses [litters])			
Doses	0	88	363	942
total fetuses (litters) examined for skeletal unclassified observations	109 (23)	101 (22)	97 (22)	110 (23)
<i>notched cartilage between basipheneoid and basioccipital</i>	2 (2)	0	0	1 (1)
<i>fused cervical arch cartilage</i>	1 (1)	0	0	0
<i>dumbbell-shaped cartilage of cervical centrum</i>	0	1 (1)	0	0
<i>hole in processus coracoideus</i>	0	1 (1)	0	0
<i>bipartite processus xiphoideus</i>	36 (14)	30 (15)	30 (15)	40 (14)

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Reference and study design	Results ^a				
	<i>notched manubrium</i>	5 (5)	7 (5)	3 (3)	1 (1)
	<i>fused rib cartilage</i>	0	0	1 (1)	0
	<i>branched rib cartilage</i>	0	0	1 (1)	0
	Combined total with skeletal unclassified cartilage observations (incidence [percent])				
	Doses	0	88	363	942
	<i>fetuses</i>	39 (36%)	35 (35%)	32 (33%)	41 (37%)
	<i>litters</i>	16 (70%)	17 (77%)	16 (73%)	15 (65%)
Borch et al. (2006)	AGD change in females (percent change compared to control)				
Rat (Wistar); 11–12 dams/group	Doses	0		600	
0, 600 mg/kg-day	<i>AGD at GD 19 (data shown in graph^b)</i>	0%		16%	
Gavage	<i>AGD at GD 20/21 (data shown in graph^b)</i>	0%		26%*	
GDs 7–19 (GD 19 c-section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time point	AGD/cubic root of BW change in females (percent change compared to control)				
	Doses	0		600	
	<i>at GD 19 (data shown in graph^b)</i>	0%		27%**	
	<i>at GD 20/21 (data shown in graph^b)</i>	0%		27%*	

1
2 ^aResponse is % control (indicated by %) or in cases when % control was not possible to present (e.g., if control
3 value was 0), response levels are presented. Equation used to calculate percent change compared to control:
4
$$\frac{\text{treated value} - \text{control value}}{\text{control value}} \times 100$$

5

6 ^bGrabIt Software used to estimate % control from graph.

7 ^cDose conversions were performed using this information: For the [Eastman Kodak \(1954\)](#) study, average BWs were
8 183, 186, 180, and 115 g for males, and 132, 126, 133, and 110 g for females at 0, 0.1, 1.0, and 5.0%, respectively.
9 Reference values for food consumption of 0.018 and 0.014 kg/day for male and female rats of an unspecified
10 species ([U.S. EPA, 1988](#)) were used.

11 ^dDose conversions were performed using this information: In [Oishi and Hiraga \(1980d\)](#), average BWs for rats and
12 mice in these studies were 145 and 25 g, respectively, and the default food consumption rates of 0.008 kg/day for
13 male Wistar rats and 0.0025 kg/day for male B6C3F₁ mice ([U.S. EPA, 1988](#)) were applied. In [Oishi and Hiraga](#)
14 ([1980a](#)), average BWs over the week-long studies were 132 and 24 g for rats and mice, respectively, and the
15 default food consumption rates of 0.008 kg/day for male Wistar rats and 0.0025 kg/day for male B6C3F₁ mice
16 were applied ([U.S. EPA, 1988](#)).

17 ^eChange in body weight was calculated by EPA.

18 ^fDose conversions were performed using this information: For [University of Rochester \(1953\)](#), average BWs were
19 139, 124, 127, 127, 121, and 101 g at 0, 0.01, 0.1, 1.0, 2.0, and 5.0%, respectively). Reference values for food
20 consumption of 0.018 and 0.014 kg/day for male and female rats of an unspecified strain ([U.S. EPA, 1988](#)) were
21 used.

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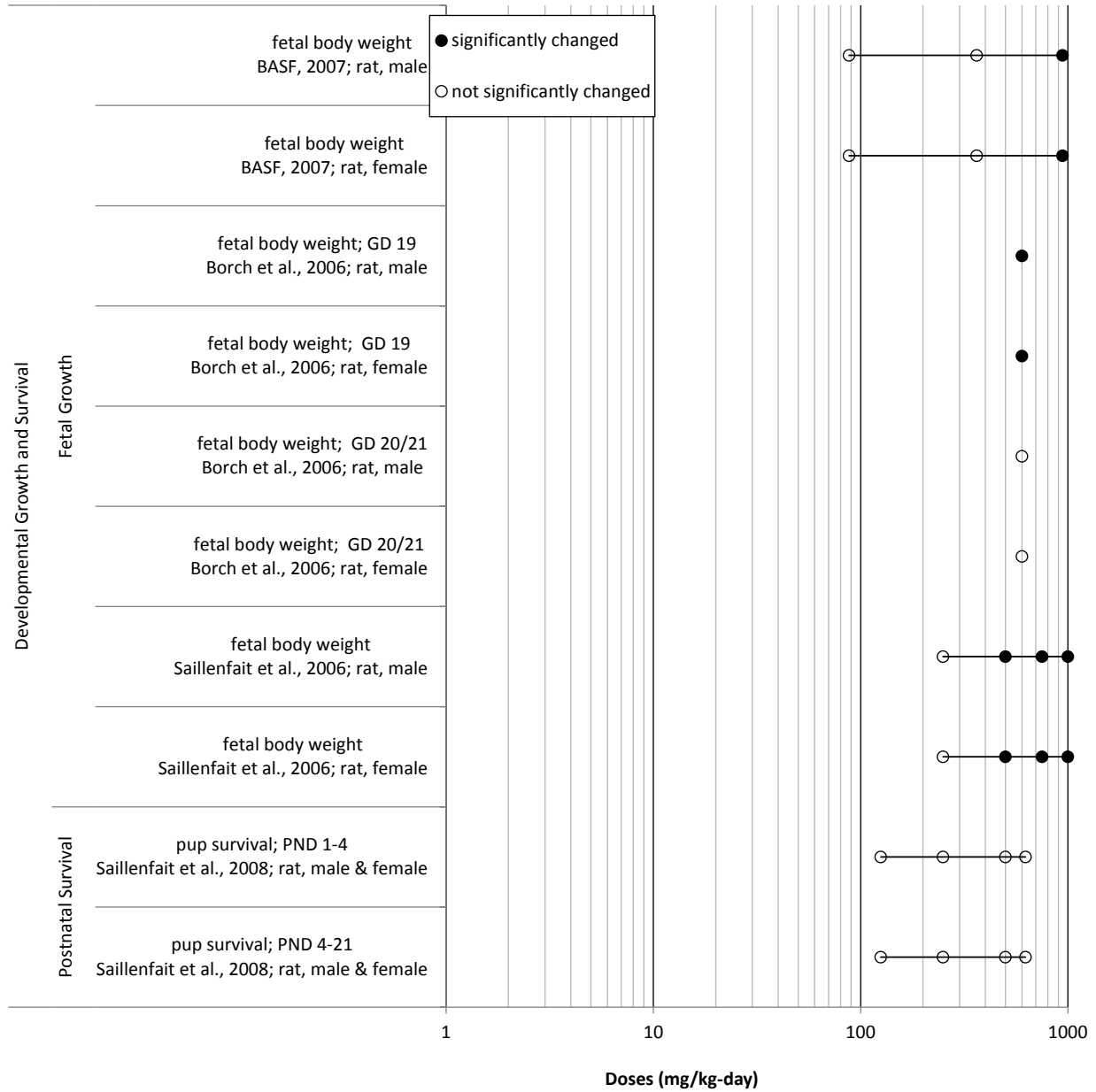
1 ^ePND 49 was selected among the periodic weight measurement ages to present in this table because it
2 corresponds to the age when VO and PPS, the developmental markers of puberty, would be expected to have
3 completed in the male and female rat.

4 ^hDose conversions were performed by EPA using this information: For [University of Rochester \(1954\)](#), average BWs
5 were 269, 277, 252, and 155 g for males, and 178, 170, 182, and 148 g for females at 0, 0.1, 1.0, and 5.0%,
6 respectively. Reference values for food consumption of 0.018 and 0.014 kg/day for male and female rats of an
7 unspecified species ([U.S. EPA, 1988](#)) were used.

8 ⁱMale reproductive organs were not evaluated in the BASF study.

9
10 * = Statistically significant difference at $p < 0.05$ from control value, as reported by study authors; ** = Statistically
11 significant difference at $p < 0.01$ from control value, as reported by study authors; *** = Statistically significant
12 difference at $p < 0.001$ from control value, as reported by study authors; BW = body weight; GD = gestation day;
13 PND = postnatal day; PPS = preputial separation; VO = vaginal opening

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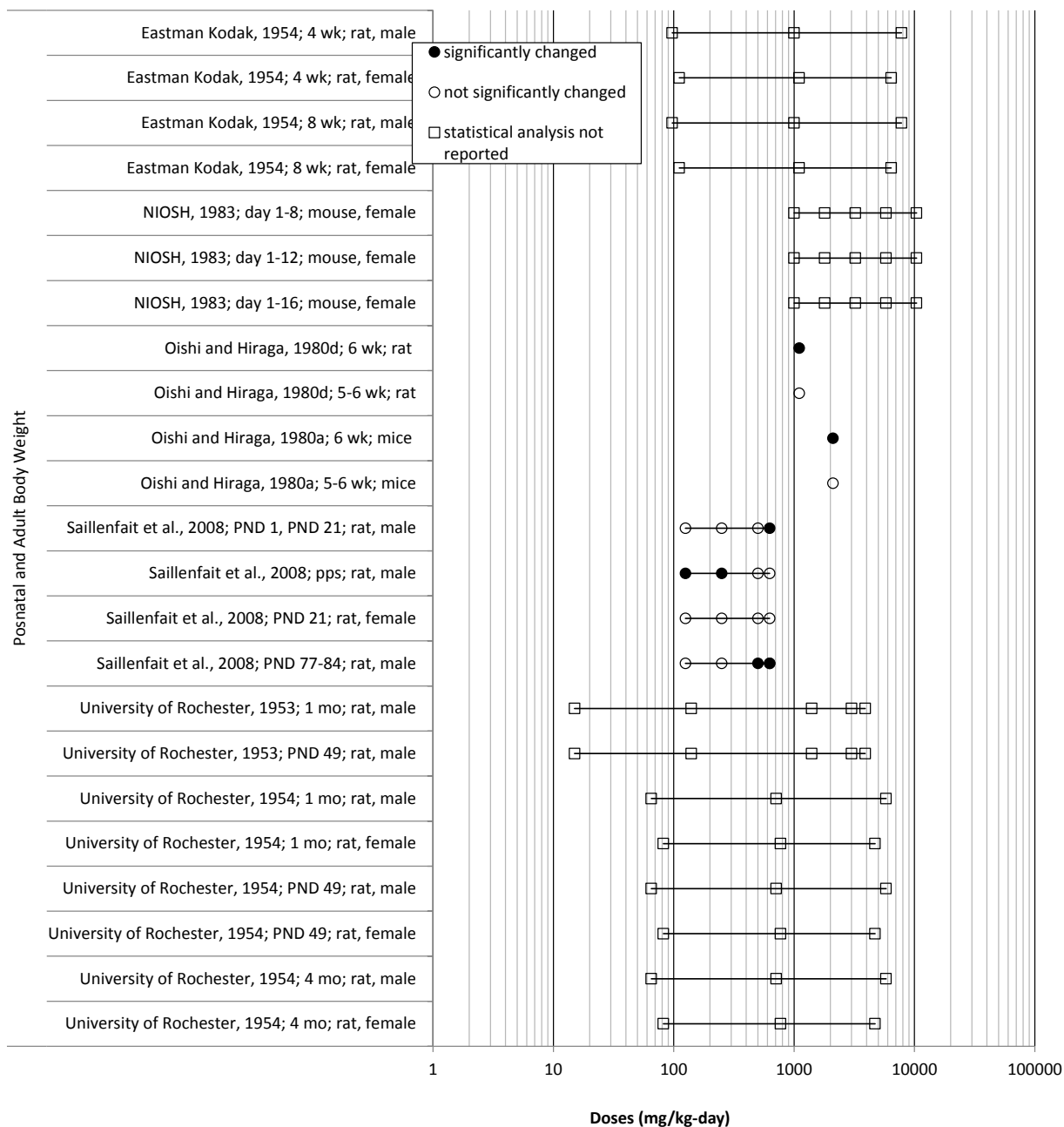


1

2 **Figure 3-1. Exposure-response array of effects on developmental growth and**
 3 **survival following developmental oral exposure to DIBP.**

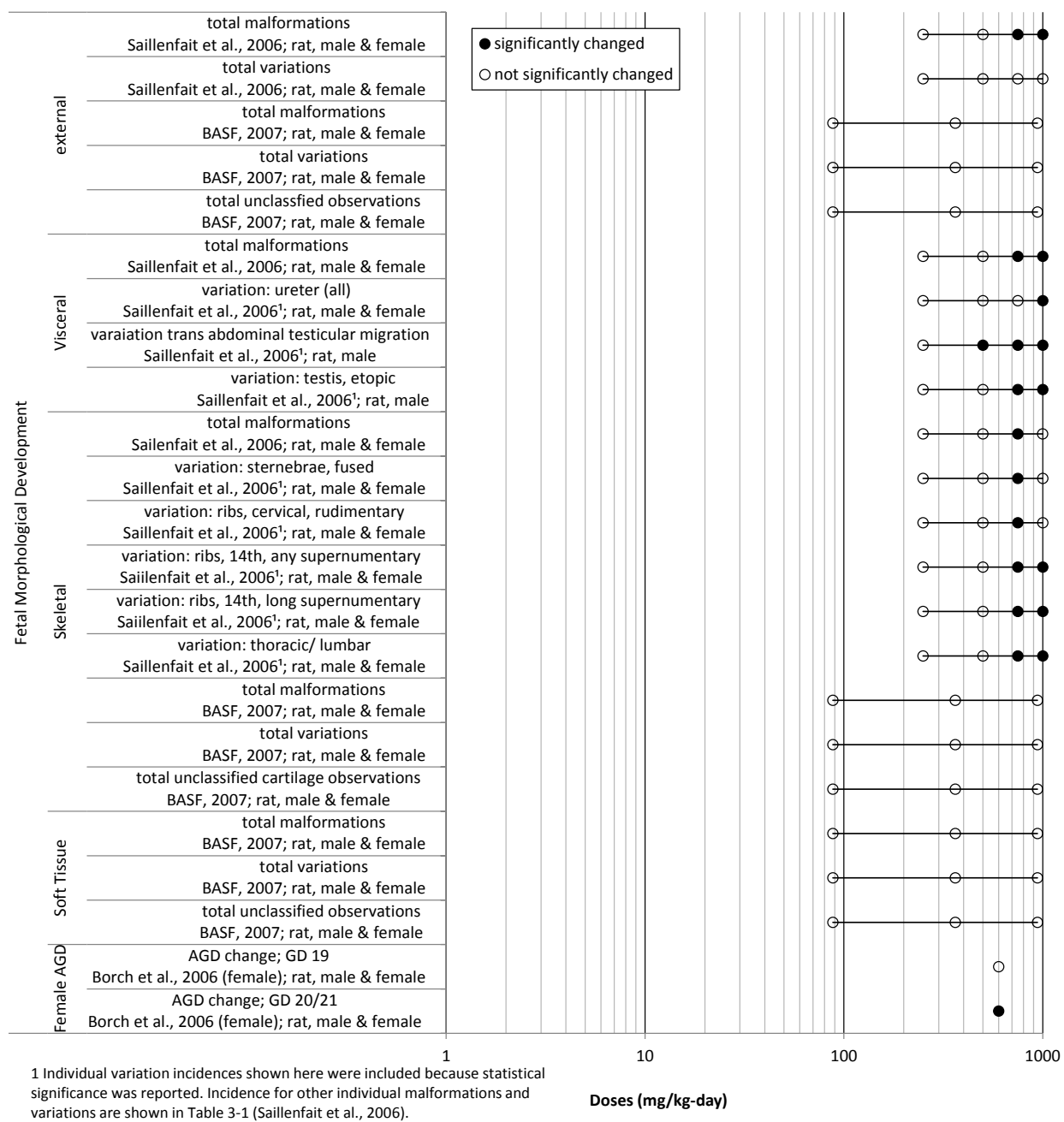
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1
 2 **Figure 3-2. Exposure-response array of effects on postnatal and adult body weight**
 3 **following developmental oral exposure to DIBP.**

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¹ Individual variation incidences shown here were included because statistical significance was reported. Incidence for other individual malformations and variations are shown in Table 3-1 (Saillenfait et al., 2006).

1

2

3

Figure 3-3. Exposure-response array of effects on fetal morphological developmental following developmental oral exposure to DIBP.

4

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1 **3.3.2. Male Reproductive Effects**

2 **Table 3-18. Evidence pertaining to male reproductive effects in animals**
 3 **following oral exposure to DIBP**

Reference and study design	Results ^a	
<i>Morphological development (assessed in fetal or postnatal development or adults)</i>		
Borch et al. (2006) Rat (Wistar); 11–12 dams/group 0, 600 mg/kg-day Gavage GDs 7–19 (GD 19 section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time point	AGD change in fetus (percent change compared to control)	
	Doses	0 600
	At GD 19 (data shown in graph ^b)	0% -15%**
	At GD 20/21 (data shown in graph ^b)	0% -11%**
	AGD/cubic root BW change in fetus (percent change compared to control)	
	Doses (M)	0 600
	GD 19 (data shown in graph ^b)	0% -5%
	GD 20/21 (data shown in graph ^b)	0% -9%**
<i>Histologic lesions in fetal testis</i>		
Borch et al. (2006) Rat (Wistar); 11–12 dams/group 0, 600 mg/kg-day Gavage GDs 7–19 (GD 19 section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time point; 1–3 males/litter	Testicular histological changes (incidence; percentage incidence in fetuses)	
	Doses	0 600
	Fetuses GD 19	
	<i>clustering of small Leydig cells</i>	2/13 9/9***
		15% 100%***
	<i>Sertoli cell vacuolization</i>	0/13 1/9
		0% 11%
	<i>central localization of gonocytes</i>	0/13 2/9
		0% 22%
	<i>multinucleated gonocytes</i>	1/13 0/9
	8% 0%	
Fetuses GD 20/21		

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Reference and study design	Results ^a					
	<i>clustering of small Leydig cells</i>	0/10		13/15***		
		0%		87%***		
	<i>Sertoli cell vacuolization</i>	0/10		14/16***		
		0%		88%***		
	<i>central localization of gonocytes</i>	0/10		14/16***		
		0%		88%***		
	<i>multinucleated gonocytes</i>	1/10		10/16*		
	10%		63%*			
Fetal testicular testosterone production						
Borch et al. (2006)	Testicular testosterone (T) content (percentage change compared to control)					
Rat (Wistar); 11–12 dams/group	Doses	0		600		
0, 600 mg/kg-day	<i>T content (GD 19 M) (data shown in graph^b)</i>	0%		-70%		
Gavage	<i>T content (GD 20/21 M) (data shown in graph^b)</i>	0%		-90%***		
GDs 7–19 (GD 19 c-section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time point	Testicular testosterone (T) production (percentage change compared to control)					
	Doses	0		600		
	<i>testicular T production ex vivo (GD 19 M) (data shown in graph^b)</i>	0%		-21%		
	<i>testicular T production ex vivo (GD 20/21 M) (data shown in graph^b)</i>	0%		-96%***		
Hannas et al. (2011)	Fetal testicular testosterone (T) production (percentage change compared to control)					
Rat (Harlan Sprague-Dawley); 3 dams/group; 3 males/dam	Doses	0	100	300	600	900
0, 100, 300, 600, 900 mg/kg-day	<i>T production</i>	0%	10%	-56%**	-80%**	-87%**
Gavage						
GDs 14–18						

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Reference and study design	Results ^a					
Hannas et al. (2012) Rat (Sprague-Dawley); 3 dams/group; 3 males/dam 0, 500 mg/kg-day Gavage GDs 14–18	Fetal testicular testosterone (T) production (percentage change compared to control)					
	Doses	0		500		
	<i>T</i> production (data shown in graph ^b)	0%		-73%**		
Howdeshell et al. (2008) Rat (Sprague-Dawley); 5–8 dams/group; 3 males/dam 0, 100, 300, 600, 900 mg/kg-day Gavage GDs 8–18; c-section on GD 18	Fetal testicular testosterone (T) production (percentage change compared to control)					
	Doses	0	100	300	600	900
	<i>T</i> production (litter mean)	0%	-5%	-40%**	-59%**	-63%**
<i>Morphological development assessed in postnatal development and adults</i>						
Saillenfait et al. (2008) Rat (Sprague-Dawley); 11–14 dams/group 0, 125, 250, 500, 625 mg/kg-day Gavage GDs 12–21 (dams allowed to deliver)	Postnatal effects (percent change in litter mean compared to control)					
	Doses	0	125	250	500	625
	AGD (PND 1)	0%	-4%	-11%*	-21%**	-22%**
	age at PPS	0%	-4%*	-1%	10%**	6%*
	Postnatal effects (incidence; percentage incidence)					
	Doses	0	125	250	500	625
	<i>retained nipples or areolas at PNDs 12–14</i>	0/76	0/78	8/96	47/79	56/76
		0%	0%	8%	59%	74%
	Note: No statistical analysis was reported by the authors for this endpoint.					
	Male adult effects at necropsy (PNDs 77–84 or 112–119; percentage incidence)					
Doses	0	125	250	500	625	
<i>retained nipples or areolas</i>	0/46	0/40	4/55	24/44	29/38	
	0%	0%	7%	55%	76%	
<i>hypospadias</i>	0/46	0/40	0/55	5/44	22/39	
	0%	0%	0%	11%	56%	

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Reference and study design	Results ^a						
	<i>exposed os penis</i>	0/46	0/40	0/55	4/44	11/39	
		0%	0%	0%	9%	28%	
	<i>cleft prepuce</i>	0/46	0/40	0/55	0/44	10/39	
		0%	0%	0%	0%	26%	
	<i>nonscrotal testis</i>	0/46	0/40	0/55	11/44	30/39	
		0%	0%	0%	25%	77%	
Note: No statistical analysis was reported by the authors for this endpoint.							
<i>Histopathologic lesions in adult testis and epididymis</i>							
Saillenfait et al. (2008) Rat (Sprague-Dawley); 11–14 dams/group 0, 125, 250, 500, 625 mg/kg-day Gavage GDs 12–21 (dams allowed to deliver)	Adult effects^c (PNDs 77–84; incidence)						
	Doses	0	125	250	500	625	
	<i>number of males (litters) examined</i>	24 (12)	20 (10)	28 (14)	22 (11)	20 (10)	
	Epididymides (number of males with effect)						
	Doses	0	125	250	500	625	
	<i>oligospermia</i>	0	1	3	2	1	
	<i>azoospermia</i>	0	1	3	10	18	
	<i>granulomatous inflammation</i>	0	0	0	4	3	
	Testes (number of males with effect)						
	Doses	0	125	250	500	625	
	<i>tubular degeneration-atrophy/hypoplasia</i>	2	2	7	16	20	
	<i>tubular necrosis</i>	0	0	1	3	5	
	<i>interstitial cell hyperplasia</i>	0	0	0	1	9	
	Note: No statistical analysis was reported by the authors for these endpoints.						

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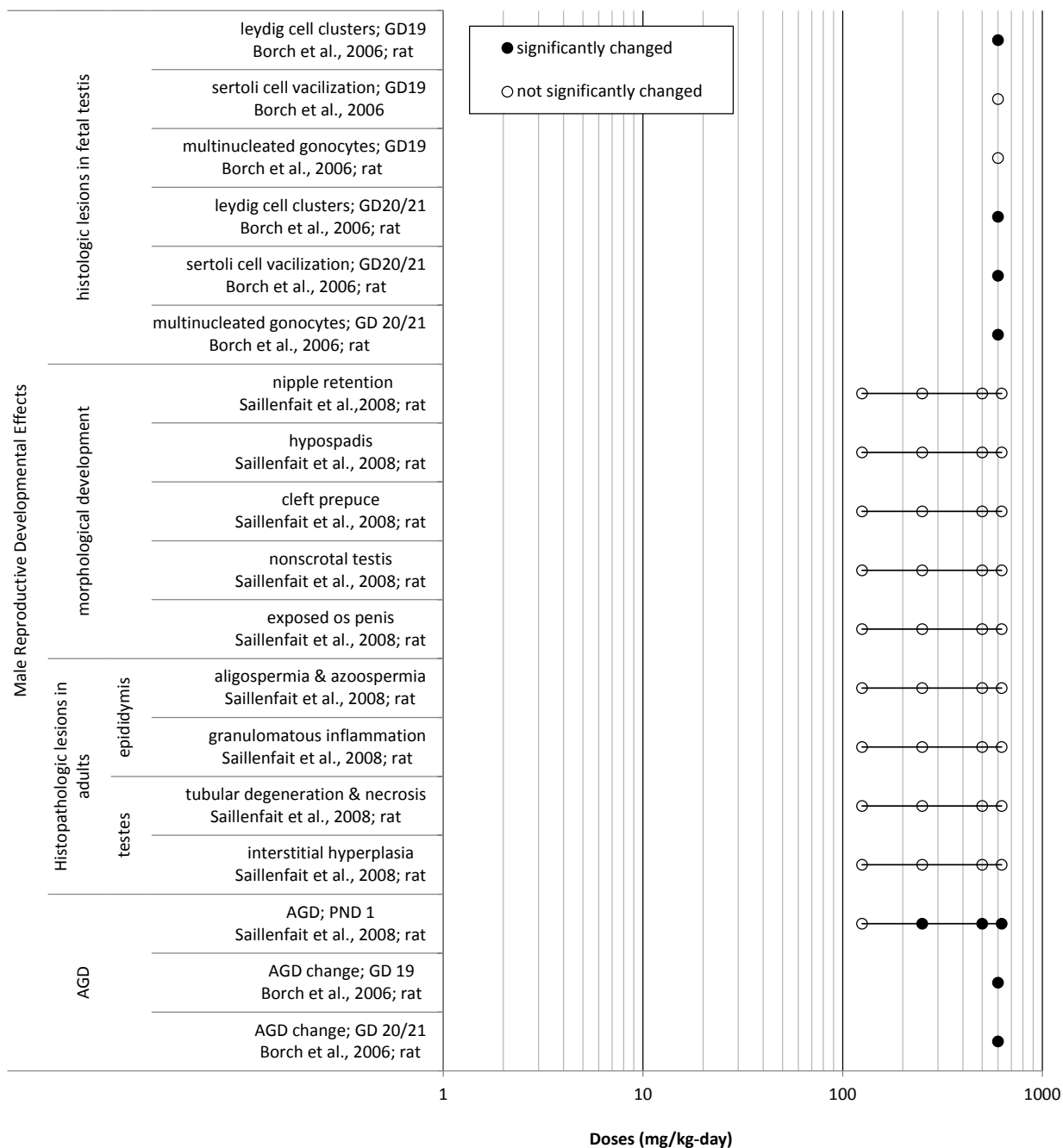
Reference and study design	Results ^a					
Oishi and Hiraga (1980b) MIBP Mouse (JCL:ICR); 10 males/group 0, 2% (0, 2,000 mg/kg-day) ^d Diet 1 week	Testes weight (percent change compared to control)					
	Doses			0		2,000
	<i>relative weight</i>			0%		45%*
Saillenfait et al. (2008) Sprague-Dawley rats; 11–14 dams/group 0, 125, 250, 500, 625 mg/kg-day Gavage GDs 12–21 (dams allowed to deliver) Assessed PNDs 77–84 (adults) after in utero exposure	Male reproductive organ weights (percent change compared to control)					
	Doses	0	125	250	500	625
	<i>right testis weight</i>	0%	1%	0%	–22%	–52%**
	<i>right epididymal weight</i>	0%	–2%	–6%	–22%**	–49%**
	<i>left testis weight</i>	0%	–2%	–1%	–13%	–59%**
	<i>left epididymal weight</i>	0%	–4%	–8%	–16%**	–49%**
	<i>seminal vesicles</i>	0%	1%	–6%	–18%**	–33%**
<i>prostate</i>	0%	–10%	–11%*	–16%**	–30%**	
University of Rochester (1954) Rat (Albino; no other strain designation); 5 males/group 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day) ^e Diet Weaning to 4 months post-weaning	Testes weight at 4 months (percent change compared to control)					
	Doses		0 ^h	65	710	5,800
	<i>absolute weight</i>		0%	2%	–1%	–70%
	<i>relative weight</i>		0%	1%	12%	–45%
Note: Statistical analysis not reported in study.						
<i>Seminal vesicle weight</i>						
(Foster et al. (1982); Foster et al. (1981)) MIBP Rat (Sprague-Dawley); 6 males/group 0, 800 mg/kg-day Gavage 6 days	Seminal vesicle weight (percent change compared to control)					
	Doses			0		800
	<i>absolute weight</i>			0%		–18%
	<i>relative weight</i>			0%		–11%

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Reference and study design	Results ^a	
<i>Prostate weight</i>		
(Foster et al. (1982); Foster et al. (1981)) MIBP Rat (Sprague-Dawley); 6 males/group 0, 800 mg/kg-day Gavage 6 days	Prostate weight (percent change compared to control)	
	Doses	0 800
	<i>absolute weight</i>	0% -13%
	<i>relative weight</i>	0% 4%
<i>Testosterone concentration in adults</i>		
Oishi and Hiraga (1980c) Rat (JCL:Wistar); 10 treated males; 20 control males 0, 2% (0, 1,200 mg/kg-day) ^d Diet 1 week	Testosterone (T) concentration (percent change compared to control)	
	Doses	0 1,200
	<i>serum T concentration (data shown in graph^b)</i>	0% 19%
	<i>testicular T concentration (data shown in graph^b)</i>	0% 158%*
	Dihydrotestosterone (DHT) concentration (percent change compared to control)	
	Doses	0 1,200
	<i>serum DHT concentration (data shown in graph^b)</i>	0% 40%
Oishi and Hiraga (1980d) MIBP Rat (JCL:Wistar); 10 males/group 0, 2% (0, 1,100 mg/kg-day) ^d Diet 1 week	Testosterone (T) concentration (percent change compared to control)	
	Doses	0 1,100
	<i>serum T concentration (data shown in graph^b)</i>	0% 61%*
	<i>testicular T concentration (data shown in graph^b)</i>	0% 161%*
Oishi and Hiraga (1980a) Mouse (JCL:ICR); 10 males/group 0, 2% (0, 2,100 mg/kg-day) ^d Diet 1 week	Testicular testosterone (T) concentration (percent change compared to control)	
	Doses	0 2,100
	<i>T concentration</i>	0% 7%
Oishi and Hiraga (1980b) MIBP Mouse (JCL:ICR); 10 males/group 0, 2% (0, 2,000 mg/kg-day) ^d	Testicular testosterone (T) concentration (percent change compared to control)	
	Doses	0 2,000
	<i>T concentration</i>	0% -83%*

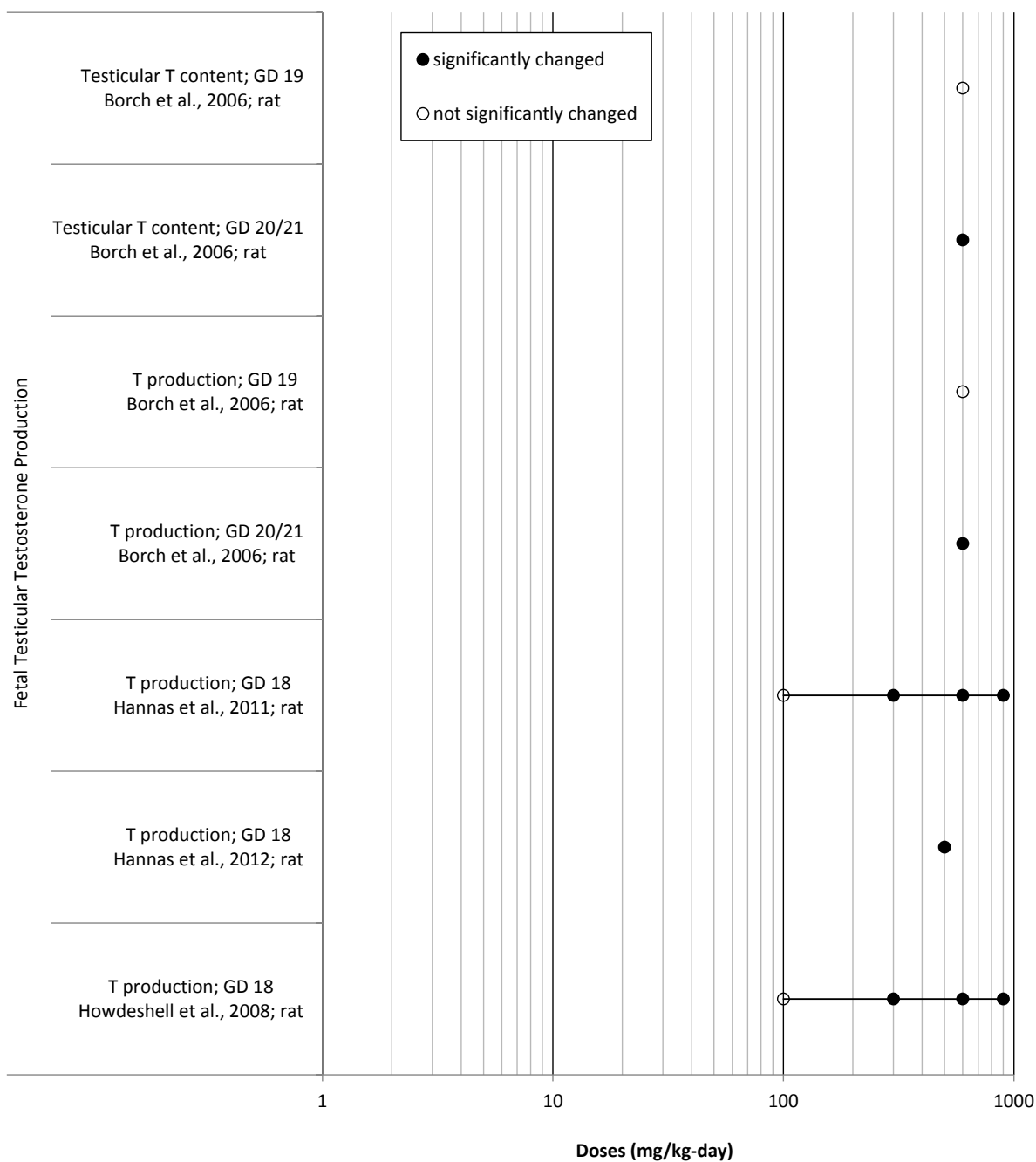
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Figure 3-4. Exposure-response array of effects on male reproductive development following developmental oral exposure to DIBP.



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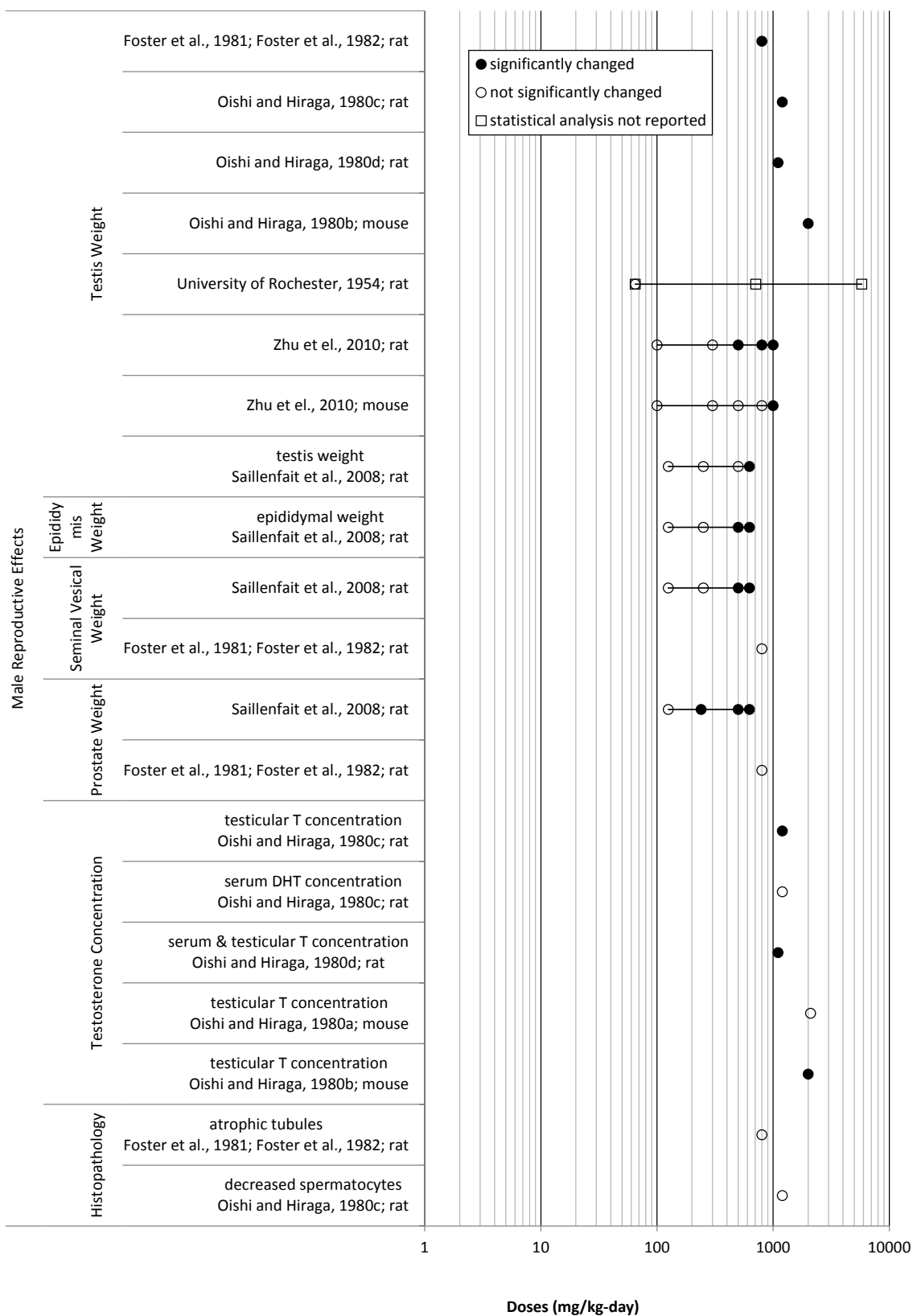
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Figure 3-5. Exposure-response array of effects on fetal testosterone (T) following developmental oral exposure to DIBP.

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Figure 3-6. Exposure-response array of male reproductive effects following oral exposure to DIBP.

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1 3.3.3. Female Reproductive Effects

2 Table 3-19. Evidence pertaining to female reproductive effects in animals
3 following oral exposure to DIBP

Reference and study design	Results					
<i>Maternal weight^a</i>						
Howdeshell et al. (2008) Rat (Sprague-Dawley); 5–8 dams/group 0, 100, 300, 600, 900 mg/kg-day Gavage GDs 8–18 (GD 18 c-section)	Maternal body weight (percent change compared to control^b)					
	Doses	0	100	300	600	900
	<i>maternal BW gain GDs 8–18</i>	0%	9%	4%	–35%	–42%*
Borch et al. (2006) Rat (Wistar); 11–12 dams/group 0, 600 mg/kg-day Gavage GDs 7–19 (GD 19 c-section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time-point	Maternal body weight					
	Doses	0	600			
		—	<i>No significant effect on maternal weight gain during pregnancy (quantitative data not reported by study authors)</i>			
BASF (2007)^c Rat (Wistar); 22–23 dams/group 0, 88, 363, 942 mg/kg-day Diet GDs 6–20 (GD 20 c-section)	Maternal body weight (percent change compared to control^b)					
	Doses	0	88	363	942	
	<i>BW change GDs 6–20</i>	0%	–3%	–6%	–11%*	
	<i>gravid uterine weight</i>	0%	–3%	–8%	–3%	
	<i>corrected BW gain GDs 6–20^d</i>	0%	–2%	–2%	–25%*	
Saillenfait et al. (2008) Rat (Sprague-Dawley); 11–14 dams/group 0, 125, 250, 500, 625 mg/kg-day Gavage GDs 12–21 (dams allowed to deliver)	Maternal body weight (percent change compared to control^b)					
	Doses	0	125	250	500	625
	<i>BW gain GDs 12–21</i>	0%	4%	6%	6%	–3%

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Reference and study design	Results					
Saillenfait et al. (2006) Rat (Sprague-Dawley); 20–22 dams/group 0, 250, 500, 750, 1,000 mg/kg-day Gavage GDs 6–20 (GD 21 c-section)	Maternal body weight (percent change compared to control^b)					
	Doses	0	250	500	750	1,000
	<i>BW gain GDs 6–21</i>	0%	-1%	-14%	-14%	-39%**
	<i>gravid uterine weight</i>	0%	-2%	-19%*	-28%**	-61%**
	<i>corrected BW gain GDs 6–21^e</i>	0%	0%	0%	19%	19%
	Maternal food consumption					
	Doses	0	250	500	750	1,000
<i>food consumption every 4–6 days, GDs 0–21</i>	– No statistically significant change from control					
<i>Maternal toxicity</i>						
BASF (2007)^c Rat (Wistar); 25 females/group 0, 88, 363, 942 mg/kg-day Diet GDs 6–20 (GD 20 c-section)	Abnormalities in dams examined at necropsy					
	Doses	0	88	363	942	
	<i>abnormalities (incidence)</i> <i>Observed: hemorrhagic thymus, diaphragmatic hernia, and dilated renal pelvis</i>	0/25	2/25	0/25	2/25	
	<i>abnormalities (percent incidence)</i>	0%	8%	0%	8%	
	Note: Statistical analysis was not performed on these data					
<i>Fertility/fetal survival</i>						
BASF (2007)^c Rat (Wistar); 25 dams/group 0, 88, 363, 942 mg/kg-day Diet GDs 6–20 (GD 20 c-section) Note: BW and food consumption measured every 1–3 days through GD 20	Fertility (percent change compared to control^b)					
	Doses	0	88	363	942	
	<i>percentage pregnant</i>	23/25	22/25	22/25	23/25	
	Fetal survival (incidence)					
	Doses	0	88	363	942	
	<i>dams with all resorptions</i>	0/23	0/22	0/22	0/23	
	Fetal survival (percent change compared to control^b)					
	Doses	0	88	363	942	
	<i>percentage preimplantation loss/litter</i>	0%	-19%	25%	-30%	
	<i>percentage postimplantation loss/litter</i>	0%	36%	105%	16%	
	<i>percentage resorptions/litter</i>	0%	36%	105%	16%	
<i>number of live fetuses/litter</i>	0%	0%	-8%	2%		
<i>number of live male fetuses/litter</i>	0%	5%	2%	-2%		

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Reference and study design	Results					
Borch et al. (2006) Rat (Wistar); 16 dams/group 0, 600 mg/kg-day Gavage GDs 7–19 (GD 19 c-section) or GDs 7–20/21 (GD 20/21 c-section); 5–6 dams/group per time-point)	Fertility (incidence)					
	Doses	0	600			
	<i>number pregnant/dams mated</i>	11/16	12/16			
	Fetal survival					
	Doses	0	600			
	— No significant effect on litter size, fetal viability, or number of resorptions (quantitative data not reported by study authors)					
Howdeshell et al. (2008) Rat (Sprague-Dawley); 5–8 dams/group 0, 100, 300, 600, 900 mg/kg-day Gavage GDs 8–18 (GD 18 c-section)	Fetal survival (n litters evaluated for endpoint)					
	Doses	0	100	300	600	900
	<i>dams with whole litter loss/total dams</i>	0/5	0/8	0/5	0/5	1/5
	<i>number of implantations/litter</i>	13.7 (3)	14.8 (4)	16.0 (3)	12.7 (3)	13.3 (5)
	<i>number of live fetuses/litter</i>	13.3 (3)	13.5 (4)	15.3 (3)	9.3 (3)	5.0* (3)
	<i>total resorptions/litter</i>	0.2 (5)	1.0 (8)	0.4 (5)	2.0 (5)	7.8* (5)
	<i>percentage fetal mortality per litter</i>	1.3% (3)	4.6% (4)	2.7% (3)	17.2% (5)	59.0%* (5)
Saillenfait et al. (2006) Rat (Sprague-Dawley); 23–24 dams/group 0, 250, 500, 750, 1,000 mg/kg-day Gavage GDs 6–20 (GD 21 c-section)	Fertility					
	Doses	0	250	500	750	1,000
	<i>number pregnant/mated dams (percent)</i>	22/24 (91.7%)	22/24 (91.7%)	22/23 (95.7%)	21/23 (91.3%)	20/24 (83.3%)
	Fetal survival (percent incidence)					
	Doses	0	250	500	750	1,000
	<i>percentage postimplantation loss/litter</i>	6.7%	11.0%	13.9%	28.2%**	59.6%**
	<i>percentage dead fetuses per litter</i>	0%	0%	0.3%	0.7%	0.3%
	<i>percentage resorptions/litter</i>	6.7%	11.0%	13.6%	27.6%**	59.3%**
	Fetal survival (percent change compared to control^b)					
	Doses	0	250	500	750	1,000
	<i>percentage live litters</i>	0%	-5%	-5%	0%	-10%
<i>number of live fetuses/litter</i>	0%	2%	-12%	-21%*	-52%**	
<i>percentage male fetuses/litter</i>	0%	-4%	-5%	0%	17%	

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Reference and study design	Results					
Saillenfait et al. (2008)	Fetal survival (percent change compared to control^b)					
Rat (Sprague-Dawley); 11–14 dams/group	Doses	0	125	250	500	625
	<i>gestation length</i>	0%	1%	0%	0%	1%
0, 125, 250, 500, 625 mg/kg-day	<i>percentage postimplantation loss per litter</i>	0%	41%	–35%	–31%	–13%
Gavage	<i>percentage pups born alive per litter</i>	0%	–1%	–4%	–1%	–8%
GDs 12–21 (dams allowed to deliver)	<i>live pups/litter at PND 1</i>	0%	5%	0%	8%	1%

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^aSome studies measured BW at multiple timepoints/lifestages and not all of these data are presented here. For the sake of comparability of data across the available studies, BW data measures presented are similar across studies and/or measures of BW change over the dosing period or greatest time period.

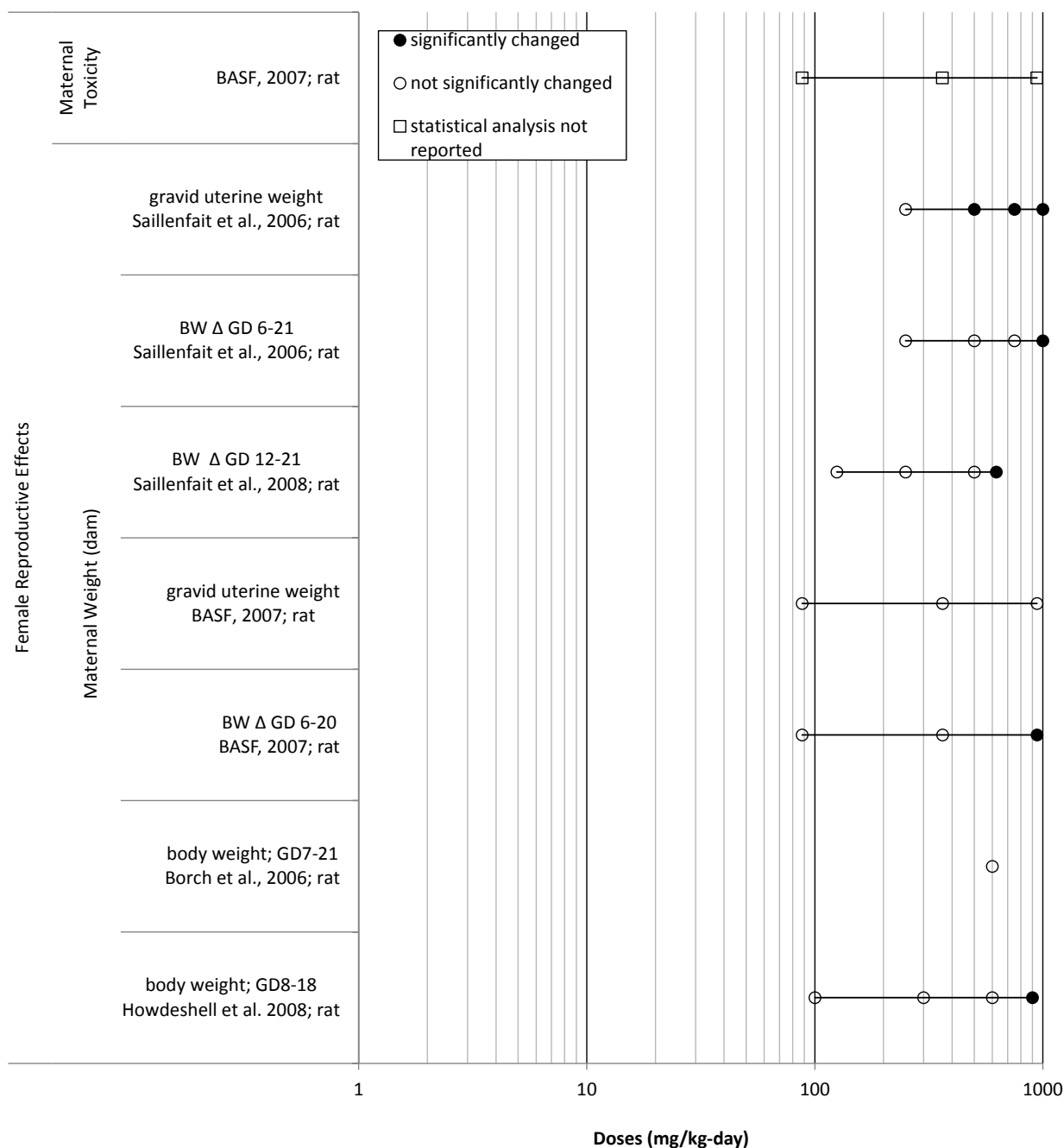
^bPercent change compared to control = $\frac{\text{treated value} - \text{control value}}{\text{control value}} \times 100$

^c[BASF \(2007\)](#): Dams in the 952 mg/kg-day group showed significantly decreased food consumption on days 10–13 and 15–17 (<10% decreased compared to control); however, overall food consumption did not differ between groups.

^dCorrected weight gain = carcass weight (GD 20 body weight – gravid uterine weight) – GD 6 body weight.

^eCorrected weight gain = BW gain GDs 6–21 – gravid uterine weight.

* = Statistically significant difference at $p < 0.05$ from control value, as reported by study authors; ** = Statistically significant difference at $p < 0.01$ from control value, as reported by study authors; *** = Statistically significant difference at $p < 0.001$ from control value, as reported by study authors.

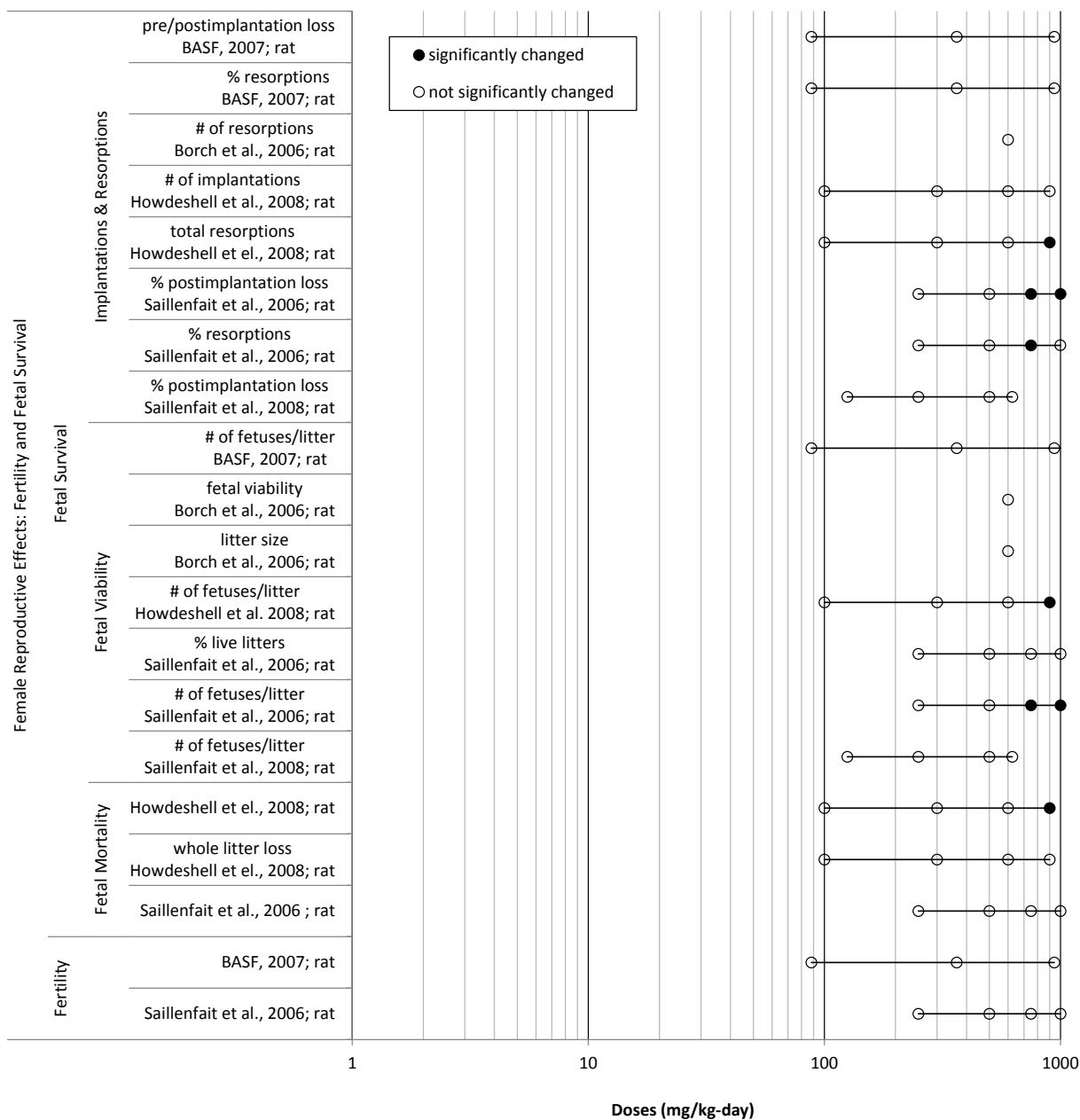


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Figure 3-7. Exposure-response array of female reproductive effects, maternal weight and toxicity, following oral exposure to DIBP.



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2 **Figure 3-8. Exposure-response array of female reproductive effects, fertility**
3 **and fetal survival, following oral exposure to DIBP.**

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1 3.3.4. Liver Effects

2 **Table 3-20. Evidence pertaining to hepatic effects in animals following oral**
 3 **exposure to DIBP**

Reference and study design	Results				
<i>Liver weight</i>					
Foster et al. (1982); Foster et al. (1981) MIBP Rat (Sprague-Dawley); 6 males/group 0, 800 mg/kg-day Gavage 4 days	Liver weight (percent change compared to control^f)				
	Doses	0	800		
	<i>relative weight</i>	0	30%***		
Oishi and Hiraga (1980c) Rat (JCL:Wistar); 10 treated males; 20 control males 0, 2% (0, 1,200 mg/kg-day) ^b Diet 1 week	Liver weight (percent change compared to control^f)				
	Doses	0	1,200		
	<i>absolute weight</i>	0%	27%*		
	<i>relative weight</i>	0%	35%*		
Oishi and Hiraga (1980a) Mouse (JCL:ICR); 10 males/group 0, 2% (0, 2,100 mg/kg-day) ^b Diet 1 week	Liver (with gallbladder) weight (percent change compared to control^f)				
	Doses	0	2,100		
	<i>relative weight</i>	0%	45%*		
Oishi and Hiraga (1980b) MIBP Mouse (JCL:ICR); 10 males/group 0, 2% (0, 2,000 mg/kg-day) ^b Diet 1 week	Liver weight (percent change compared to control^f)				
	Doses	0	2,000		
	<i>relative weight</i>	0%	30%*		
University of Rochester (1954) Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^c Diet Weaning to 4 months post-weaning	Liver weight (percent change compared to control^f)				
	Doses (M)	0	65	710	5,800
	<i>absolute weight</i>	0%	6%	11%	5%
	<i>relative weight</i>	0%	2%	22%	84%
	Doses (F)	0 ^f	82	770	4,700
	<i>absolute weight</i>	0%	0%	16%	41%

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Reference and study design	Results						
	<i>relative weight</i>	0%	0%	8%	62%		
	Note: Statistical analysis not reported in study.						
<u>University of Rochester (1953)</u>	Liver weight (percent change compared to control^a)						
Rat (Albino; no other strain designation); 5 males/dose	Doses	0	15	140	1,400	3,000	8,900
0, 0.01, 0.1, 1, 2, 5% (0, 15, 140, 1,400, 3,000, 8,900 mg/kg-day) ^d	<i>absolute weight</i>	0%	-17%	-12%	5%	15%	13%
Diet	<i>relative weight</i>	0%	-1%	5%	26%	43%	79%
Weaning to 1 month post-weaning	Note: Statistical analysis not reported in study.						
<i>Liver histopathology</i>							
<u>University of Rochester (1954)</u>	Liver histopathology						
Rat (Albino; no other strain designation); 5 males and 5 females/dose	<i>No treatment-related differences from control were observed at any dose group.</i>						
0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^c	Note: 4–5 animals per dose group were assessed. Statistical analysis not reported in study.						
Diet							
Weaning to 4 months post-weaning							
<u>University of Rochester (1953)</u>	Liver histopathology						
Rat (Albino; no other strain designation); 5 males/dose	<i>Histopathological findings only noted in the control group. No treatment-related differences were observed.</i>						
0, 0.01, 0.1, 1, 2, 5% (0, 15, 140, 1,400, 3,000, 8,900 mg/kg-day) ^d	Notes: Number of animals assessed is unclear. Findings limited to “filled with coarse granular cytoplasm.” Statistical analysis not reported in study.						
Diet							
Weaning to 1 month post-weaning							

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^aPercent change compared to control = $\frac{\text{treated value} - \text{control value}}{\text{control value}} \times 100$

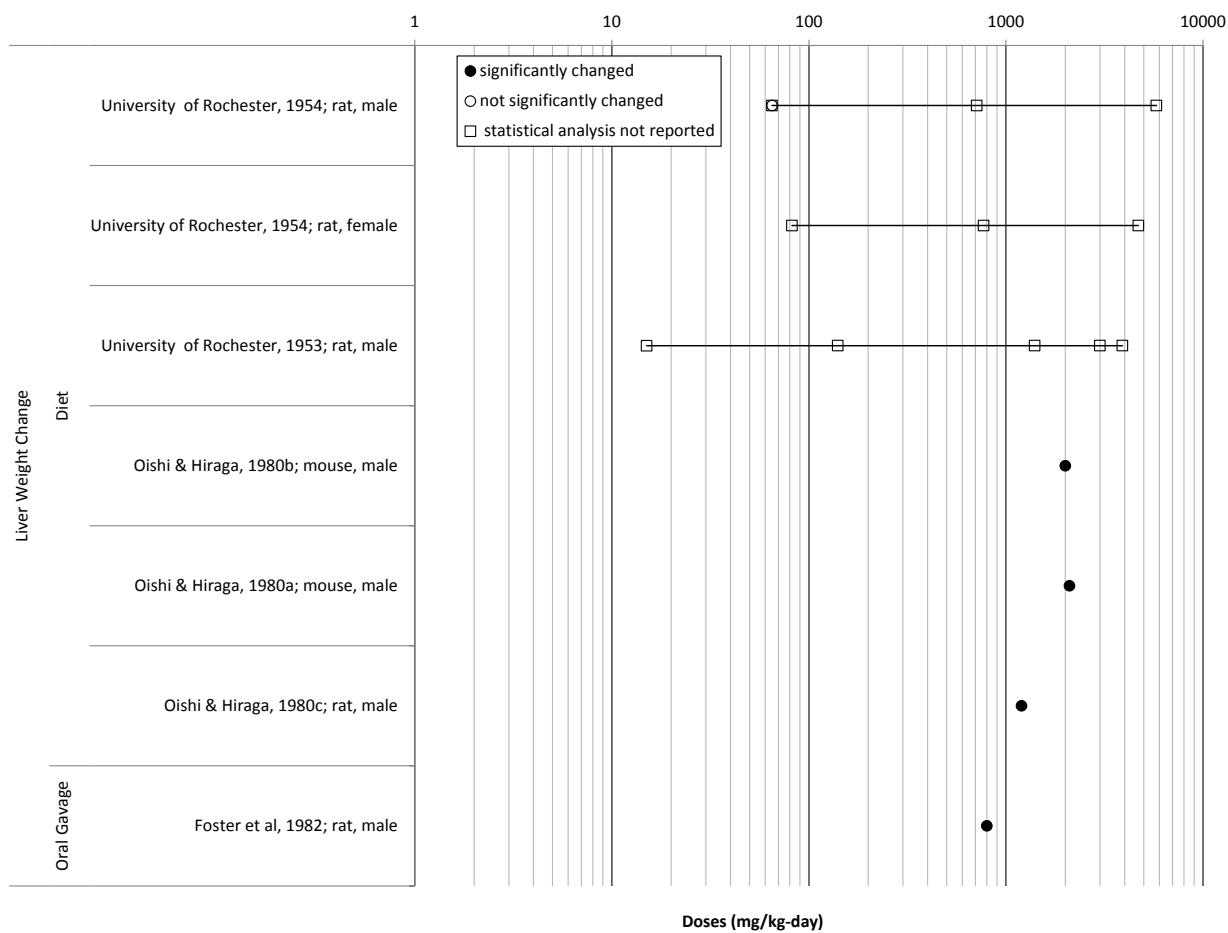
^bDose conversions were performed using this information: (Oishi and Hiraga (1980a), 1980c) average BWs over the week-long studies were 132 and 24 g for rats and mice, respectively, and the default food consumption rates of 0.008 kg/day for male weanling Wistar rats and 0.0025 kg/day for male weanling B6C3F₁ mice were applied (U.S. EPA, 1988). Oishi and Hiraga (1980b) average BWs for rats and mice in these studies were 145 and 25 g, respectively, and the default food consumption rates of 0.008 kg/day for male weanling Wistar rats and 0.0025kg/day for male weanling B6C3F₁ mice (U.S. EPA, 1988) were applied. Note that Table 1-6 of U.S. EPA (1988) listed the default food consumption rate for male weanling Wistar rats as 0.080 kg/day. However, it was later determined using an equation in Table 1-3 of the document that this value was actually supposed to be 0.008 kg/day.

^cDose conversions were performed using this information: University of Rochester (1954) average BWs (measured at least once weekly) were 269, 277, 252, and 155 g for male rats, and 178, 170, 182, and 148 g for female rats at

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1 0, 0.1, 1.0, and 5.0%, respectively; and the default food consumption of 0.018 for male and 0.014 kg/day for
2 female rats ([U.S. EPA, 1988](#)) of an unspecified strain in a subchronic study were applied.
3 ^dDose conversions were performed using this information: [University of Rochester \(1953\)](#) average BWs (measured
4 weekly) of the rats were 139, 124, 127, 127, 121, and 101 g at 0, 0.01, 0.1, 1.0, 2.0, and 5.0%, respectively; and
5 the default food consumption of 0.018 for male rats and 0.014 kg/day for female rats ([U.S. EPA, 1988](#)) of an
6 unspecified strain in a subchronic study were applied.
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8 * = Statistically significant difference at $p < 0.05$ from control value, as reported by study authors; ** = Statistically
9 significant difference at $p < 0.01$ from control value, as reported by study authors; *** = Statistically significant
10 difference at $p < 0.001$ from control value, as reported by study authors.
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Figure 3-9. Exposure-response array of liver effects following oral exposure to DIBP.

1 3.3.5. Kidney Effects

2 Table 3-21. Evidence pertaining to renal effects in animals following oral
3 exposure to DIBP

Reference and study design	Results				
<i>Kidney weight</i>					
Foster et al. (1982); Foster et al. (1981) MIBP Rat (Sprague-Dawley); 6 males/group 0, 800 mg/kg-day Gavage 4 days	Kidney weight (percent change compared to control^a)				
	Doses	0	800		
	<i>relative weight</i>	0	396%***		
Oishi and Hiraga (1980c) Rat (JCL:Wistar); 10 treated males; 20 control males 0, 2% (0, 1,200 mg/kg-day) ^b Diet 1 week	Kidney weight (percent change compared to control^a)				
	Doses	0	1,200		
	<i>absolute weight</i>	0%	-5%		
	<i>relative weight</i>	0%	2%		
Oishi and Hiraga (1980b) MIBP Mouse (JCL:ICR); 10 males/group 0, 2% (0, 2,000 mg/kg-day) ^b Diet 1 week	Kidney weight (percent change compared to control^a)				
	Doses	0	2,000		
	<i>relative weight</i>	0%	-5%		
Oishi and Hiraga (1980a) Mouse (JCL:ICR); 10 males/group Diet 0, 2% (0, 2,100 mg/kg-day) ^b 1 week	Kidney weight (percent change compared to control^a)				
	Doses	0	2,100		
	<i>relative weight</i>	0%	-10%*		
University of Rochester (1954) Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^c Diet	Kidney weight (percent change compared to control^a)				
	Doses (M)	0	65	710	5,800
	<i>absolute weight</i>	0%	10%	9%	-31%
	<i>relative weight</i>	0%	7%	20%	22%

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Reference and study design	Results						
Weaning to 4 months post-weaning	Doses (F)	0	82	770	4,700		
	<i>absolute weight</i>	0%	4%	11%	-2%		
	<i>relative weight</i>	0%	5%	4%	13%		
	Note: Statistical analysis not reported in study.						
<u>University of Rochester (1953)</u>	Kidney weight (percent change compared to control^a)						
Rat (Albino; no other strain designation); 5 males/dose 0, 0.01, 0.1, 1, 2, 5% (0, 15, 142, 1,417, 2,975, 8,911 mg/kg-day) ^d Diet Weaning to 1 month post-weaning	Doses	0	15	140	1,400	3,000	8,900
	<i>absolute weight</i>	0%	-14%	-11%	-11%	-11%	-23%
	<i>relative weight</i>	0%	3%	7%	7%	12%	23%
Note: Statistical analysis not reported in study.							
Kidney histopathology							
<u>University of Rochester (1954)</u>	Kidney histopathology						
Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^c Diet Weaning to 4 months post-weaning	<i>No treatment-related differences in males or females were observed compared with control.</i>						
	Note: 4–5 animals per dose group were assessed. Findings in males limited to pyelitis, granuloma, and pyelonephritis; Findings in females limited to pyelitis and pyelonephritis.						
<u>University of Rochester (1953)</u>	Kidney histopathology						
Rat (Albino; no other strain designation); 5 males/dose 0, 0.01, 0.1, 1, 2, 5% (0, 15, 142, 1,417, 2,975, 8,911 mg/kg-day) ^d Diet Weaning to 1 month post-weaning	<i>No treatment-related differences were observed.</i>						
	Note: Number of animals assessed is unclear. Findings limited to eosinophils and inflammatory cells. Statistical analysis not reported in study.						

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2 ^aPercent change compared to control = $\frac{\text{treated value} - \text{control value}}{\text{control value}} \times 100$
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4 ^bDose conversions were performed using this information: (Oishi and Hiraga (1980a), 1980c)) average BWs over
5 the week-long studies were 132 and 24 g for rats and mice, respectively, and the default food consumption rates
6 of 0.008 kg/day for male weanling Wistar rats and 0.0025 kg/day for male weanling B6C3F₁ mice were applied
7 (U.S. EPA, 1988). Oishi and Hiraga (1980b) average BWs for rats and mice in these studies were 145 and 25 g,
8 respectively, and the default food consumption rates of 0.008 kg/day for male weanling Wistar rats and 0.0025
9 kg/day for male weanling B6C3F₁ mice (U.S. EPA, 1988) were applied. Note that Table 1-6 of U.S. EPA (1988)
10 listed the default food consumption rate for male weanling Wistar rats as 0.080 kg/day. However, it was later

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1 determined using an equation in Table 1-3 of the document that this value was actually supposed to be 0.008
2 kg/day.

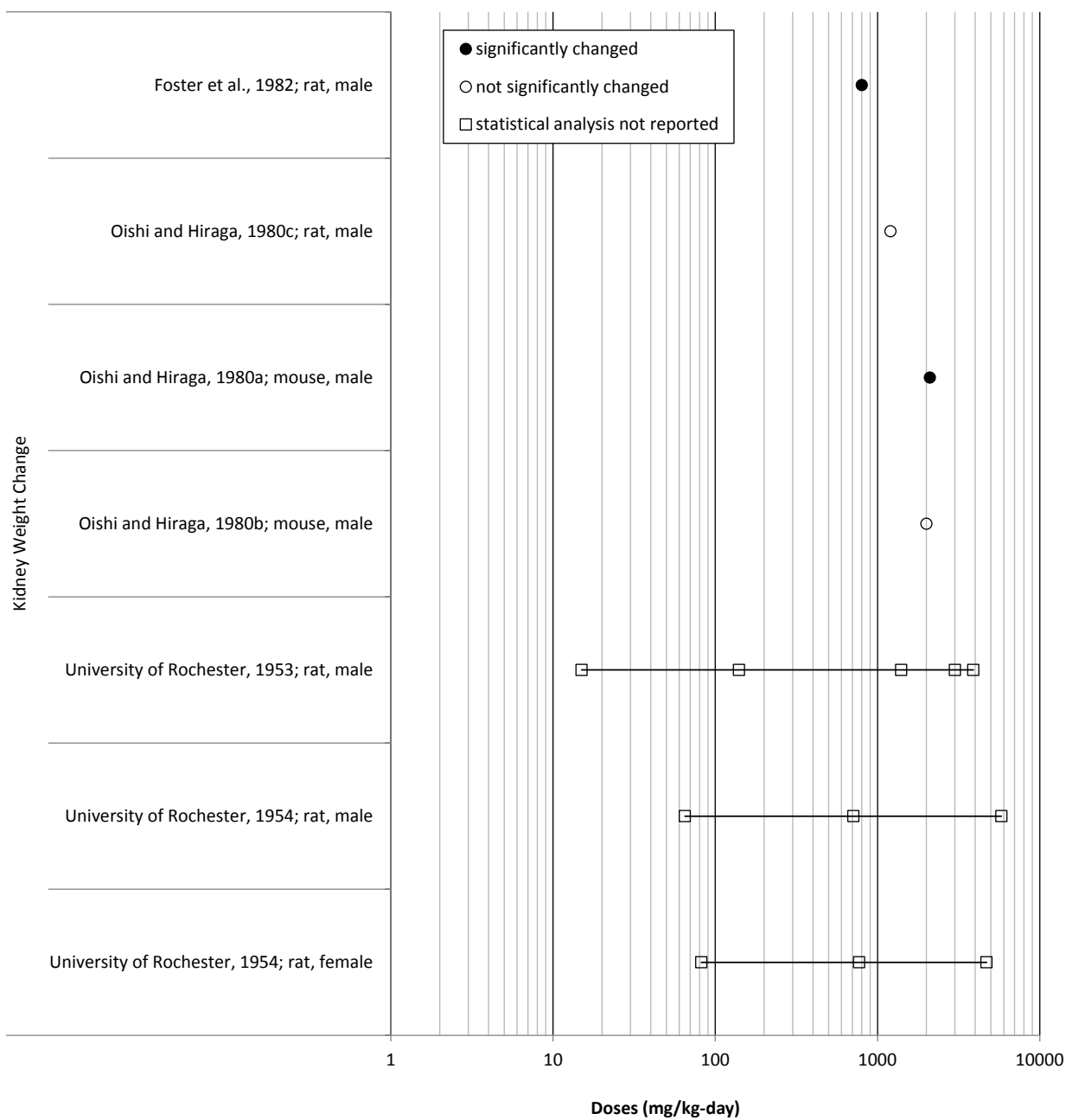
3 ^cDose conversions were performed using this information: [University of Rochester \(1954\)](#) average BWs (measured
4 at least once weekly) were 269, 277, 252, and 155 g for male rats, and 178, 170, 182, and 148 g for female rats at
5 0, 0.1, 1.0, and 5.0%, respectively; and default food consumption of 0.018 kg/day for male rats and 0.014 kg/day
6 for female rats ([U.S. EPA, 1988](#)) of an unspecified strain in a subchronic study were applied.

7 ^dDose conversions were performed using this information: [University of Rochester \(1953\)](#) average BWs (measured
8 weekly) were 139, 124, 127, 127, 121, and 101 g at 0, 0.01, 0.1, 1.0, 2.0, and 5.0%, respectively; and default food
9 consumption of 0.018 for male rats and 0.014 kg/day for female rats ([U.S. EPA, 1988](#)) of an unspecified strain in a
10 subchronic study were applied.

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12 * = Statistically significant difference at $p < 0.05$ from control value, as reported by study authors.

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Figure 3-10. Exposure-response array of kidney effects following oral exposure to DIBP.

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1 3.3.6. Hematopoietic Effects

2 Table 3-22. Evidence pertaining to hematopoietic effects in animals following
3 oral exposure to DIBP

Reference and study design	Results				
<i>Hematology</i>					
<p>University of Rochester (1954)</p> <p>Rat (Albino; no other strain designation); 5 males and 5 females/dose</p> <p>0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females)^b</p> <p>Diet</p> <p>Weaning to 4 months post-weaning</p>	Hematology at 4 months (<i>percent change compared to control^a</i>)				
	Doses (M)	0	65	710	5,800
	RBCs	0%	-1%	-5%	-16%
	WBCs	0%	-37%	-15%	38%
	Hgb	0%	-4%	-5%	-9%
	Doses (F)	0	82	770	4,700
	RBCs	0%	6%	1%	13%
	WBCs	0%	-19%	-8%	29%
	Hgb	0%	0%	3%	-6%
	Note: Statistical analysis not reported in study.				
<p>University of Rochester (1954)</p> <p>Rat (Albino; no other strain designation); 5 males and 5 females/dose</p> <p>0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females)^b</p> <p>Diet</p> <p>Weaning to 4 months post-weaning</p>	Differential counts at 4 months (<i>percent of each type of WBC</i>)				
	Doses (M)	0	65	710	5,800
	<i>neutrophils</i>	20%	18%	19%	17%
	<i>eosinophils</i>	1%	5%	5%	2%
	<i>basophils</i>	0%	0%	1%	1%
	<i>lymphocytes</i>	79%	76%	75%	80%
	<i>monocytes</i>	1%	0%	0%	0%
	<i>myeloids</i>	0%	0%	0%	0%
	<i>blast forms</i>	0%	0%	0%	0%
	<i>plasma cells</i>	0%	0%	0%	0%
	Doses (F)	0	82	770	4,700
	<i>neutrophils</i>	15%	14%	22%	13%
	<i>eosinophils</i>	2%	4%	4%	3%
	<i>basophils</i>	0%	0%	0%	1%
<i>lymphocytes</i>	83%	82%	73%	84%	
<i>monocytes</i>	0%	0%	0%	0%	

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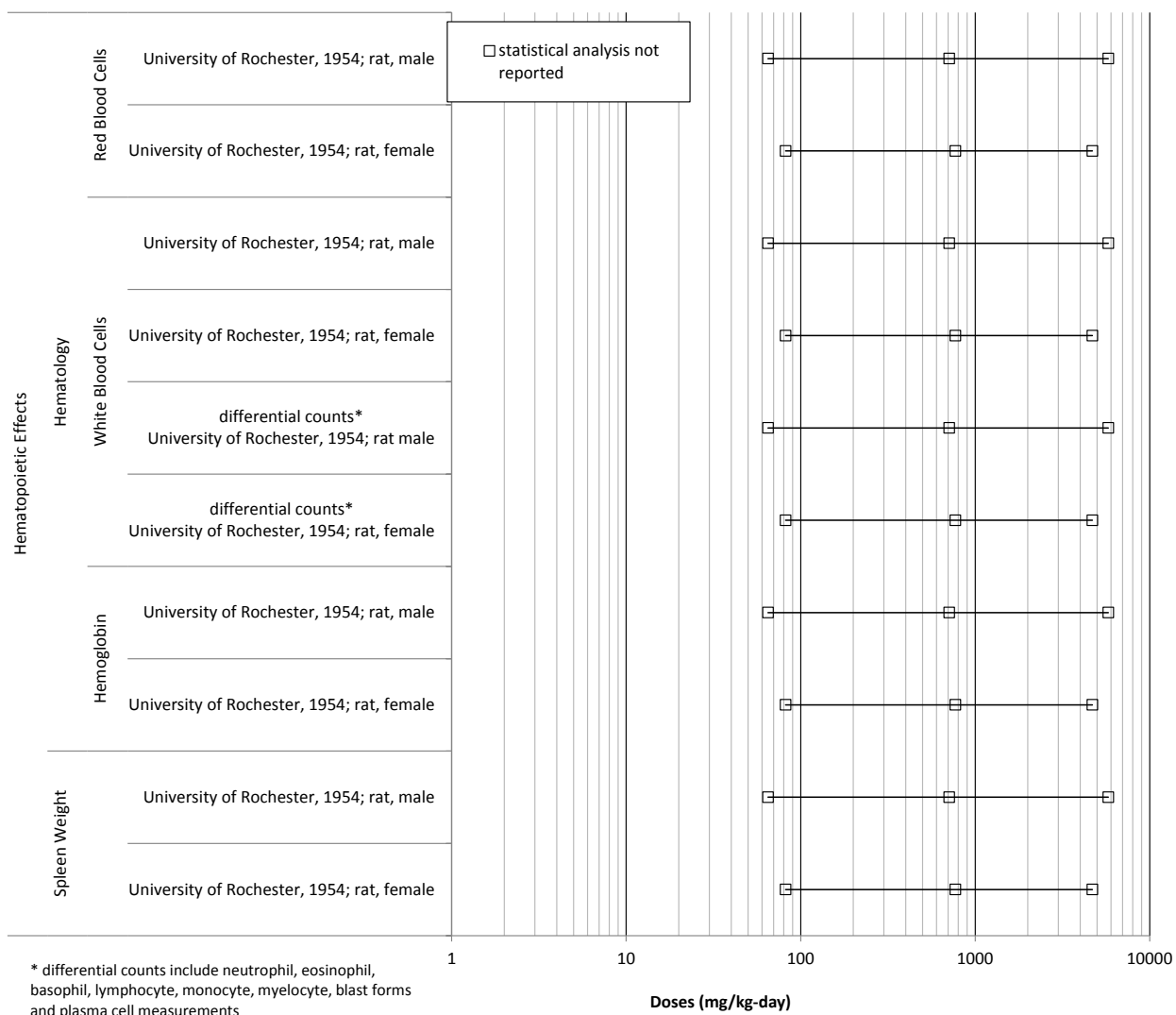
Reference and study design	Results				
	<i>myeloids</i>	0%	0%	0%	0%
	<i>blast forms</i>	0%	0%	0%	0%
Note: Statistical analysis not reported in study.					
<i>Spleen weight</i>					
University of Rochester (1954)	Spleen weight (<i>percent change compared to control^a</i>)				
Rat (Albino; no other strain designation); 5 males and 5 females/dose	Doses (M)	0	65	710	5,800
0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^b	<i>absolute weight</i>	2%	9%	7%	-13%
Diet	<i>relative weight</i>	0%	5%	17%	52%
Weaning to 4 months post-weaning	Doses (F)	0	82	770	4,700
	<i>absolute weight</i>	0%	8%	93%	-6%
	<i>relative weight</i>	0%	11%	80%	9%
Note: Statistical analysis not reported in study.					

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^aPercent change compared to control = $\frac{\text{treated value} - \text{control value}}{\text{control value}} \times 100$

^bDose conversions were performed by EPA using this information: [University of Rochester \(1954\)](#) average BWs (measured at least once weekly) were 269, 277, 252, and 155 g for male rats, and 178, 170, 182, and 148 g for female rats at 0, 0.1, 1.0, and 5.0%, respectively; and the default food consumption of 0.018 kg/day for male rats and 0.014 kg/day for female rats ([U.S. EPA, 1988](#)) of an unspecified strain in a subchronic study were applied.

Hgb = hemoglobin; RBC = red blood cell; WBC = white blood cell



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Figure 3-11. Exposure-response array of hematopoietic effects following oral exposure to DIBP.

1 3.3.7. Other Effects

2 Table 3-23. Evidence pertaining to other toxicity effects in animals following
3 oral exposure to DIBP

Reference and study design	Results				
<i>Neurotoxicity effects</i>					
<u>University of Rochester (1954)</u> Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^b Diet Weaning to 4 months post-weaning	Brain weight (percent change compared to control ^d)				
	Doses (M)	0	65	710	5,800
	<i>absolute weight</i>	0%	1%	0%	-3%
	<i>relative weight</i>	0%	-2%	11%	72%
	Doses (F)	0	82	770	4,700
	<i>absolute weight</i>	0%	3%	1%	2%
	<i>relative weight</i>	0%	4%	-6%	17%
	Note: Statistical analysis not reported in study.				
	<i>Cardiac effects</i>				
<u>University of Rochester (1954)</u> Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^b Diet Weaning to 4 months post-weaning	Heart weight (percent change compared to control ^d)				
	Doses (M)	0	65	710	5,800
	<i>absolute weight</i>	0%	2%	-6%	-28%
	<i>relative weight</i>	0%	-3%	3%	24%
	Doses (F)	0	82	770	4,700
	<i>absolute weight</i>	0%	10%	-4%	11%
	<i>relative weight</i>	0%	10%	-11%	28%
	Note: Statistical analysis not reported in study.				
	<i>Lung effects</i>				
<u>University of Rochester (1954)</u> Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^b Diet	Lung weight (percent change compared to control ^d)				
	Doses (M)	0	65	710	5,800
	<i>absolute weight</i>	0%	10%	4%	-30%
	<i>relative weight</i>	0%	5%	16%	23%

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Reference and study design	Results						
Weaning to 4 months post-weaning	Doses (F)	0	82	770	4,700		
	<i>absolute weight</i>	0%	-20%	-6%	-27%		
	<i>relative weight</i>	0%	-19%	-12%	-16%		
	Note: Statistical analysis not reported in study.						
<i>Stomach effects</i>							
University of Rochester (1954)	Stomach weight (percent change compared to control^d)						
Rat (Albino; no other strain designation); 5 males and 5 females/dose 0, 0.1, 1, 5% (0, 65, 710, 5,800 mg/kg-day for males; 0, 82, 770, 4,700 mg/kg-day for females) ^b Diet Weaning to 4 months post-weaning	Doses (M)	0	65	710	5,800		
	<i>absolute weight</i>	0%	10%	6%	1%		
	<i>relative weight</i>	0%	7%	18%	80%		
	Doses (F)	0	82	770	4,700		
	<i>absolute weight</i>	0%	8%	7%	18%		
	<i>relative weight</i>	0%	9%	0%	35%		
Note: Statistical analysis not reported in study.							
<i>Clinical signs</i>							
(Hazleton Laboratories (1992), 1987); NIOSH (1983)	Clinical signs of toxicity (incidence / total animals)						
Mouse (CD-1); 50 control females, 10 females/treated group 0, 1,000, 1,795, 3,225, 5,790, 10,400 mg/kg-day Gavage 8 days	Doses	0	1,000	1,795	3,225	5,790	10,400
	<i>languid</i>	1/50	0/10	0/10	2/10	5/10	0/10
	<i>prostrate</i>	0/50	0/10	0/10	0/10	4/10	0/10
	<i>ataxia</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>hunched</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>tremors</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>head tilt</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>thin</i>	1/50	0/10	0/10	0/10	0/10	0/10
	<i>wheezing</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>dyspnea</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>urine stains</i>	0/50	0/10	0/10	0/10	0/10	10/10
	<i>alopecia</i>	0/50	0/10	0/10	0/10	0/10	0/10
	<i>rough hair coat</i>	0/50	0/10	0/10	0/10	0/10	10/10
	<i>sores</i>	0/50	0/10	0/10	0/10	0/10	0/10
<i>piloerection</i>	0/50	0/10	0/10	0/10	0/10	0/10	
<i>opaque eyes</i>	N/A	0/10	0/10	0/10	1/10	0/10	

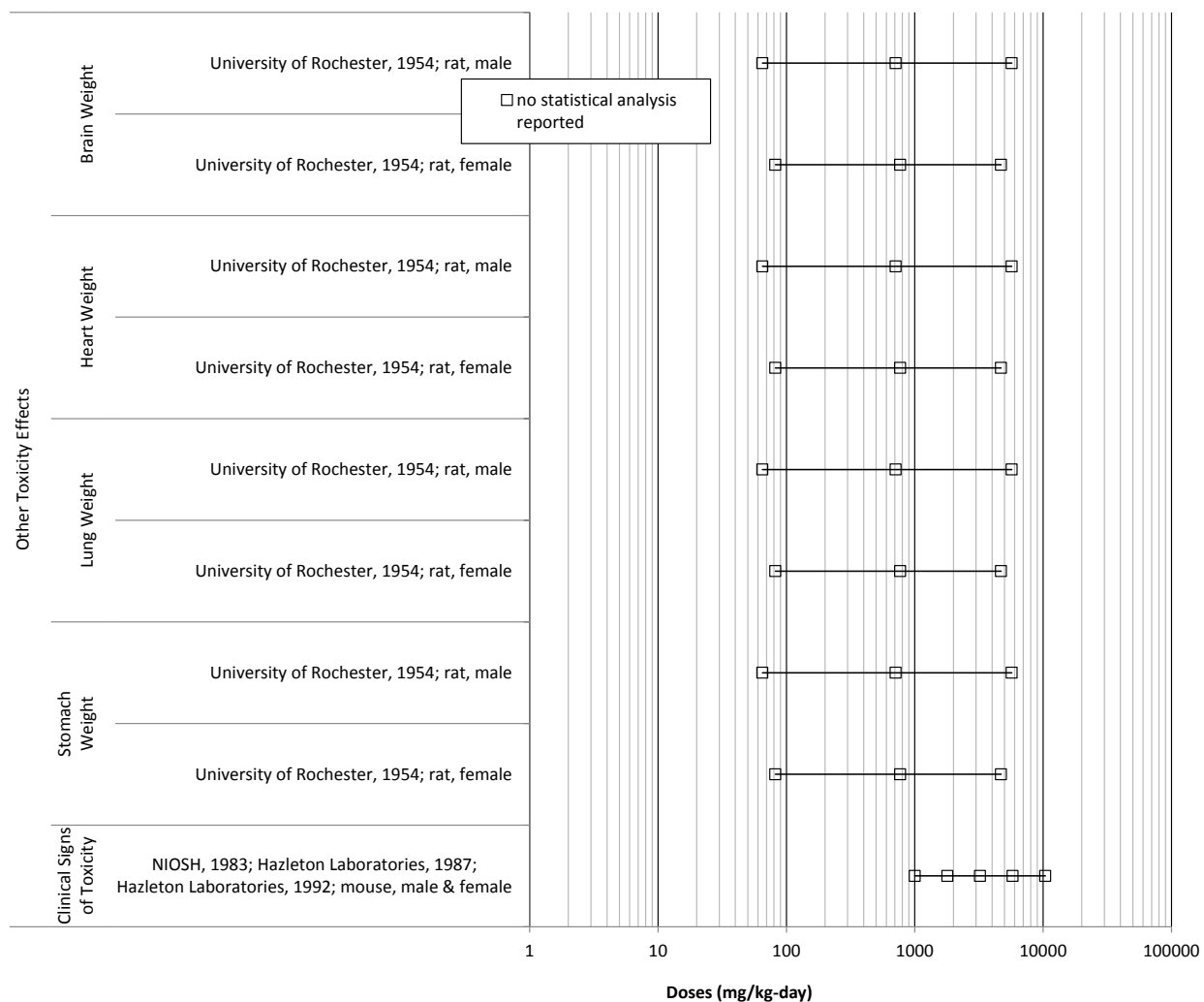
This document is a draft for review purposes only and does not constitute Agency policy.

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Reference and study design	Results						
	<i>discoloration (yellow hair)</i>	N/A	0/10	0/10	0/10	1/10	0/10
Note: Statistical analysis not reported in study for clinical signs data.							

- 1
- 2 ^aPercent change compared to control = $\frac{\text{treated value} - \text{control value}}{\text{control value}} \times 100$
- 3
- 4 ^bDose conversions were performed using this information: [University of Rochester \(1954\)](#) average BWs (measured
- 5 at least once weekly) were 269, 277, 252, and 155 g for male rats, and 178, 170, 182, and 148 g for female rats at
- 6 0, 0.1, 1.0, and 5.0%, respectively; and default food consumption of 0.018 kg/day for male rats and 0.014 kg/day
- 7 for female rats ([U.S. EPA, 1988](#)) of an unspecified strain in a subchronic study were applied.
- 8

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1
2
3
4

Figure 3-12. Exposure-response array of effects on other toxicities following oral exposure to DIBP.

3.4. PRELIMINARY MECHANISTIC INFORMATION FOR DIBP

The systematic literature search for DIBP also identified studies evaluating mechanisms of action considered potentially relevant to effects observed following exposure to DIBP. Studies were included if they evaluated mechanistic events following exposure to DIBP or the metabolite, MIBP, or contained information relevant to the mechanistic understanding of DIBP toxicity. Reviews or analyses that do not contain original data are not included here, but may be considered in later stages of assessment development.

The diverse array of mechanistic studies presented here includes investigations of the cellular, biochemical, and molecular mechanisms underlying toxicological outcomes. For this preliminary evaluation, information reported in each study was extracted into a database (in the form of an Excel spreadsheet) that will facilitate future evaluation of mechanistic information. This information is being made available to provide an opportunity for stakeholder input, including the identification of relevant studies not captured here.

The information extracted from each study and included in the database, corresponds to the column headings in the spreadsheet, and is as follows: link to the HERO record (contained within a URL that links to the study abstract in the HERO database), HERO ID, author(s), year, molecular formulation, in vitro/in vivo, species, cell type, endpoint(s) (i.e., mechanistic outcomes), assay, and mechanistic category. The database supports sorting capabilities, e.g., data can be organized by assay. The database is available through HERO at [http://hero.epa.gov/index.cfm?action=reference.details&reference_id=2508641]. To access the database, click on the link at the top of the web page and select “download” and then “ok” to view the spreadsheet in Excel. This spreadsheet may also be saved to your desktop by downloading and selecting “save.” The resulting inventory of DIBP mechanistic studies consists of 32 mechanistic outcomes from 13 identified in vivo studies, as well as 28 mechanistic outcomes from 23 in vitro assays. Table 3-24 presents a summary of the mechanistic outcomes recording in the database from each study identified.

The mechanistic categories developed here are not mutually exclusive and are designed to facilitate the analysis of similar studies and experimental observations in a systematic manner. This process will allow the identification of mechanistic events that contribute to mode(s) of action (MOAs) and/or adverse outcome pathways (AOPs) following DIBP exposure. The mechanistic categories assigned to each mechanistic outcome reported by an individual study are as follows: (1) mutation, including investigations of gene and chromosomal mutation; (2) DNA damage, including indicator assays of genetic damage; (3) DNA repair; (4) oxidative stress; (5) cell death and division (this captures a broad range of assays, but it is useful to consider them together as observations resulting from cell cycle alterations; (6) pathology, which includes morphological evaluations pertaining to the dysfunction of organs, tissues, and cells; (7) epigenetic effects, which are observations of heritable changes in gene function that cannot be explained by changes in the DNA sequence; (8) receptor-mediated and cell signaling effects; (9) immune system effects; (10) cellular differentiation and transformation; (11) cellular energetics; and (12) “other,” to

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1 capture those mechanistic outcomes not easily assigned to a defined category. Mechanistic
 2 outcomes in the “other” category include sex steroid hormone (e.g., testosterone) production and
 3 gene expression.
 4

5 **Table 3-24. Summary of mechanistic outcomes evaluated following DIBP or**
 6 **MIBP administration**

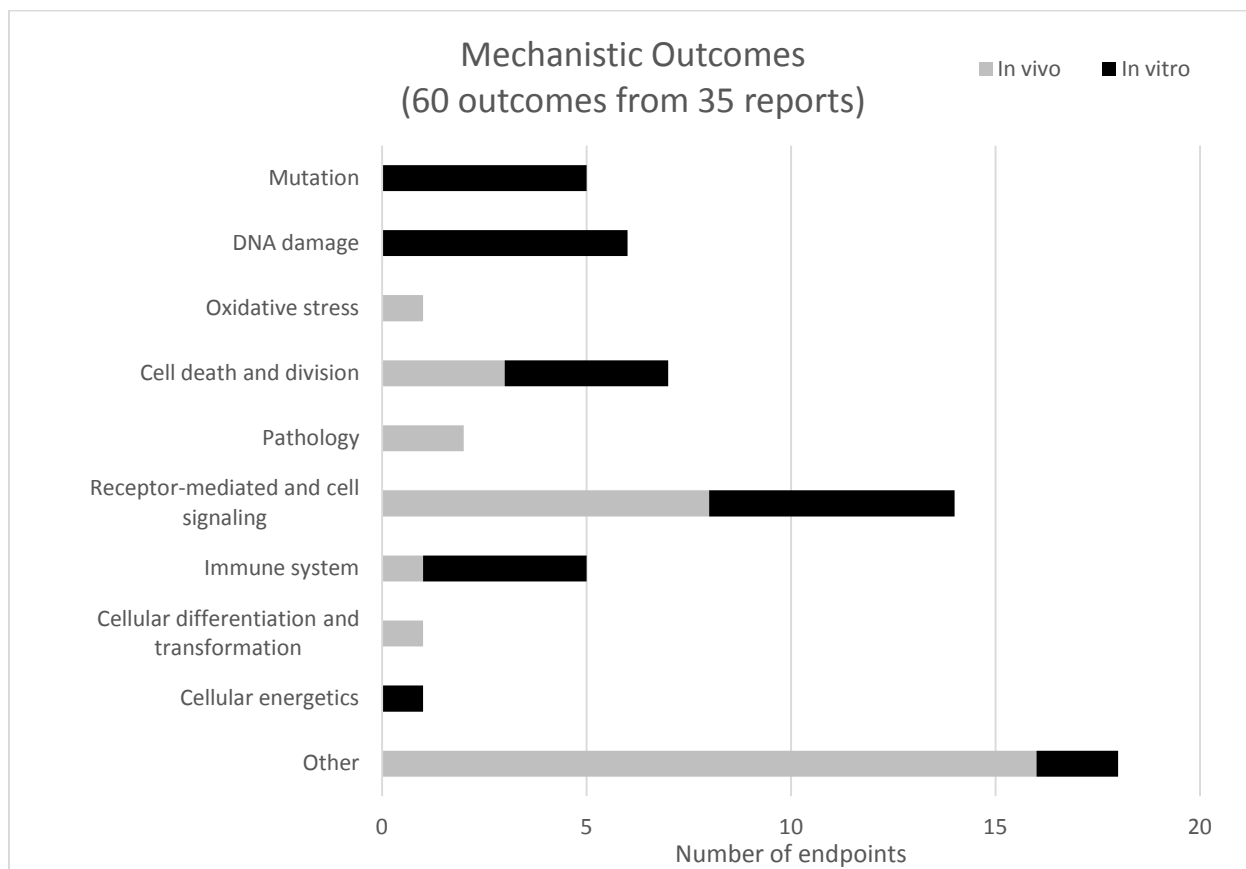
Mechanistic category	Total # outcomes/ # studies	In vivo (# outcomes/ # studies)				In vitro (# outcomes/# studies)				
		Total	Human	Rat	Mouse	Total	Human	Primate	Rat	Mouse
Mutation ^a	5/5	0	0	0	0	5/5	0	0	0	0
DNA damage	6/4	0	0	0	0	6/4	6/4	0	0	0
<i>DNA repair</i>										
Oxidative stress ^b	1/1	1/1	0	0	0	0	0	0	0	0
Cell death and division	7/5	3/2	0	2/2	1/1	4/3	3/2	0	0	1/1
Pathology	2/2	2/2	0	2/2	0	N/A	N/A			
<i>Epigenetics</i>										
Receptor-mediated and cell signaling ^c	14/9	8/4	1/1	7/3	0	6/5	0	1/1	0	0
Immune system	5/3	1/1	0	1/1	0	4/2	1/1	0	2/1	1/1
Cellular differentiation and transformation	1/1	1/1	1/1	0	0	0	0	0	0	0
Cellular energetics	1/1	0	0	0	0	1/1	0	0	1/1	0
Other ^d	18/11	16/10	0	14/8	2/2	2/2	0	0	2/2	0
Total	60/35		32/13			28/23				

7
 8 ^aDatabase included five outcomes in five studies utilizing *Salmonella typhimurium*.
 9 ^bDatabase included one outcome from one study utilizing *Caenorhabditis elegans*.
 10 ^cDatabase included two outcomes from one study utilizing cultured hamster cells, two outcomes from two studies
 11 utilizing yeast, and one cell-free system.
 12 ^dDatabase primarily composed of hormone (testosterone, estradiol) content or production in tissues from rats and
 13 mice.
 14

15 Notes: The number in rows may not sum to “total” amounts as several studies evaluated multiple species or
 16 employed both in vivo and in vitro models. The mechanistic categories in italics and in gray shading had no DIBP-
 17 specific information available.
 18

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1 Information summarized in Table 3-24 and Figure 3-13 and detailed in the mechanistic
2 database can be used to ascertain the breadth and scope of available mechanistic studies. At this
3 preliminary stage, study results are not presented. Additionally, the inclusion of a study in the
4 spreadsheet does not reflect conclusions reached as to mechanistic study quality or relevance.
5 After the epidemiological and experimental studies on each health effect have been synthesized,
6 mechanistic studies will be reviewed and findings synthesized to evaluate potential MOAs and/or
7 AOPs, which can be used to inform hazard identification and dose-response assessment, specifically
8 addressing questions of human relevance, susceptibility, and dose-response relationships.
9
10



11
12 **Figure 3-13. Summary of in vivo and in vitro mechanistic data for DIBP and**
13 **MIBP by mechanistic category.**

14

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