

# Statistical Considerations in Studying Epigenetic Changes

Integrative Analysis of Metabolomics and Epigenetics  
using the ELEMENT Cohort

Peter Song

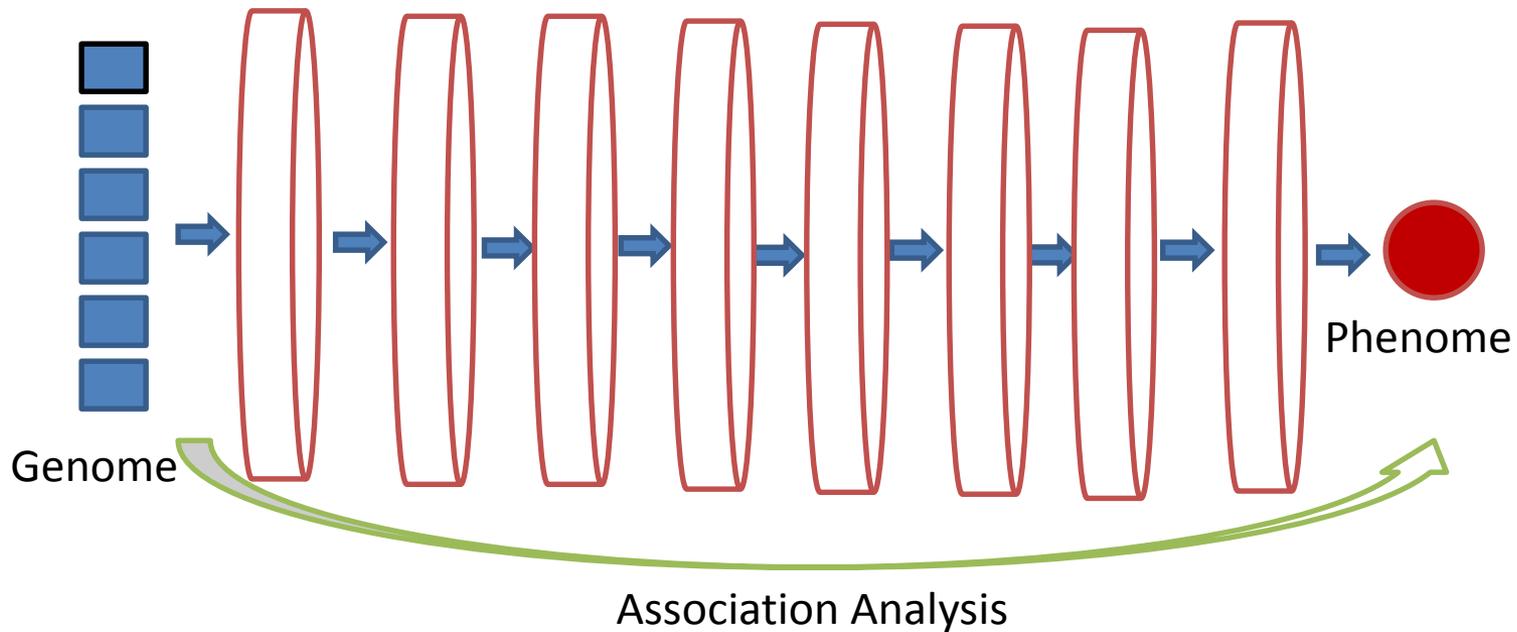
Department of Biostatistics

University of Michigan School of Public Health

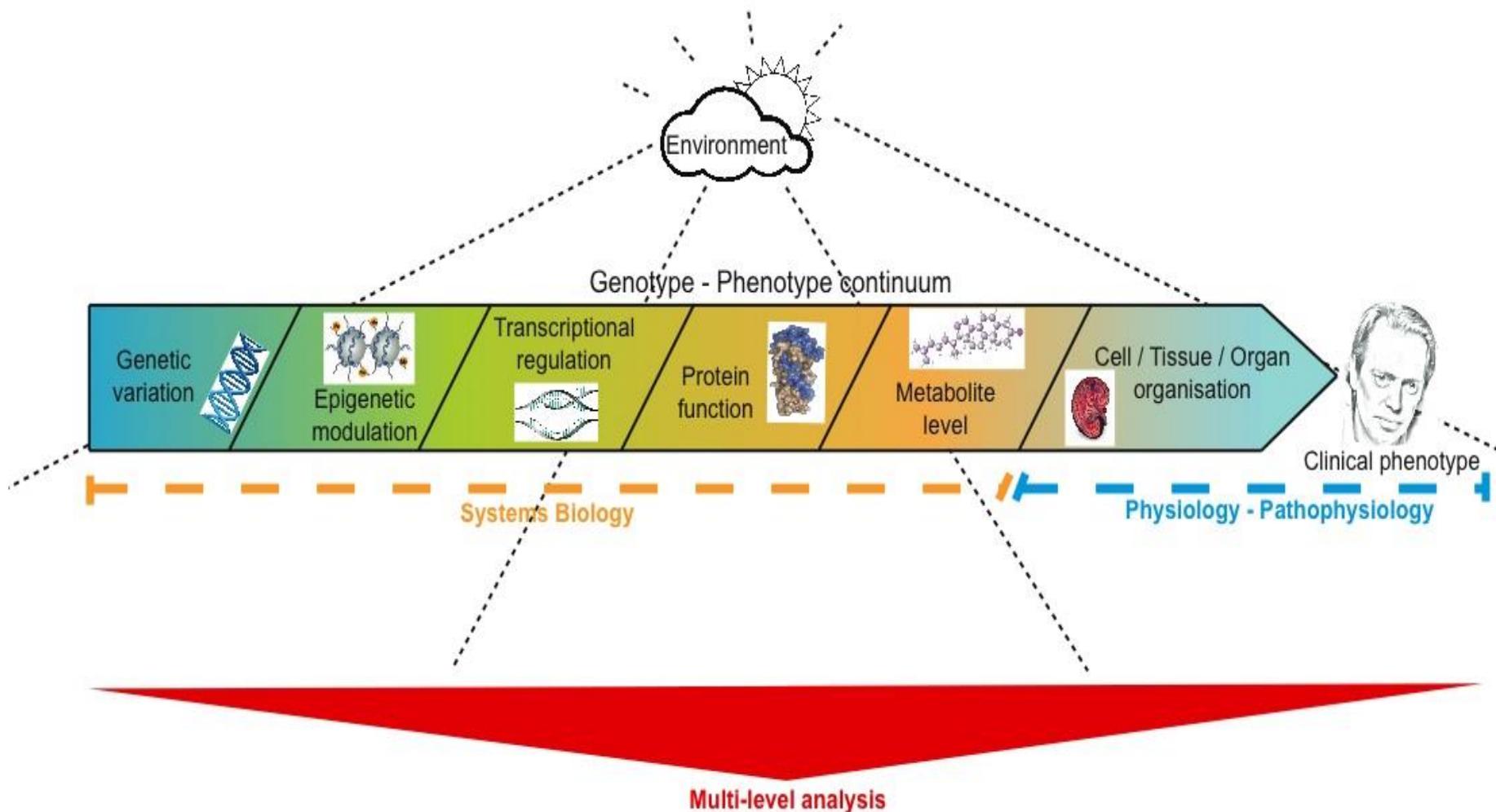
September 2, 2015

**Funding:** P20 ES018171/RD 834800 , P30 ES017885  
P01 ES02284401/RD 83543601, P30 DK089503, RO1 ES024732

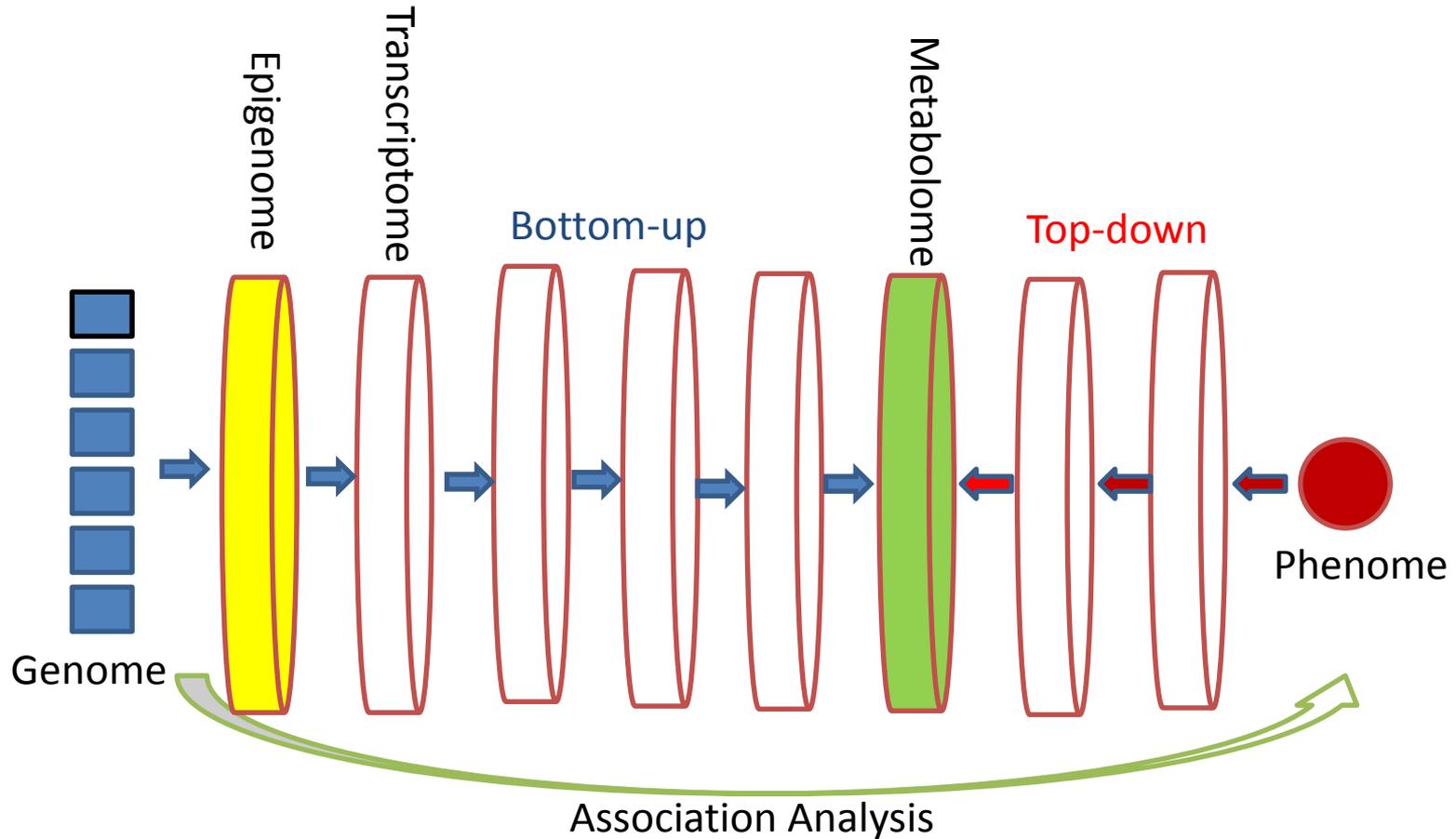
# Pitch I: Biological pathway is the key to success



# Translational Biomedical Sciences



# Epigenome and Metabolome



Both “bottom-up” and “top-down” analyses may involve high throughput (high-dimensional) data, and it is an analytic challenge to overcome in order to minimize false discovery.

# Integrative Analysis of Metabolomics and Epigenetic Data in Environmental Health Research

- Prenatal, early postnatal, and concurrent exposures influence children's growth and development
- Epigenetic perturbations
  - Biomarker of persistent changes from prenatal exposures and/or the cumulative impact of continued exposures
  - Stable yet potentially modifiable (via pharmacological, dietary, and perhaps lifestyle changes)
  - Mechanistic link between exposures and outcomes
- Metabolite profiles
  - Biomarker of phenotypic changes and possible risk for future chronic disease
  - Mechanistic link between exposures, epigenetics, and outcomes
- Cumulative Risk Assessment
  - Children are exposed to multiple environmental agents throughout development
  - Understanding relationships between exposures, the epigenome, metabolome, and growth will enable better understanding of the cumulative impact of common exposures on life-long health

## Pitch II:

Epigenetic change/effect is hard to measure

**Numerical (stochastic) change or  
biological (genetic) change?**



- Design of time points for data collection
- Quality control during data collection
- Data pre-processing and pre-treatment
- Types of changes to extract from cleaned data
- Bring biology in and focus on target genes or not
- Others...

# 250 Subjects from ELEMENT Longitudinal Cohort Study

- Early Life Exposure in Mexico to ENvironmental Toxicants (ELEMENT)
  - 4 longitudinal cohorts: 1994, 1997, 2000, 2005
  - Low-to-moderate income populations in Mexico City
  - Lead exposure is measured in prenatal and concurrent

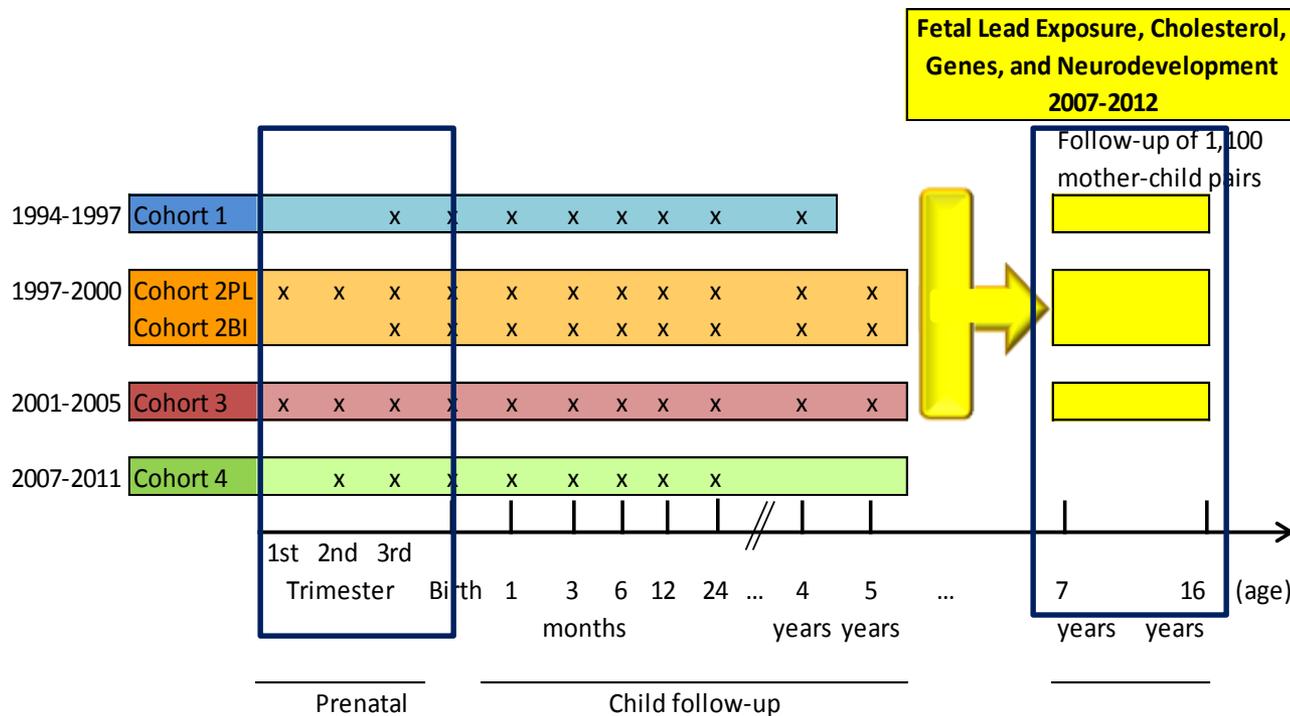
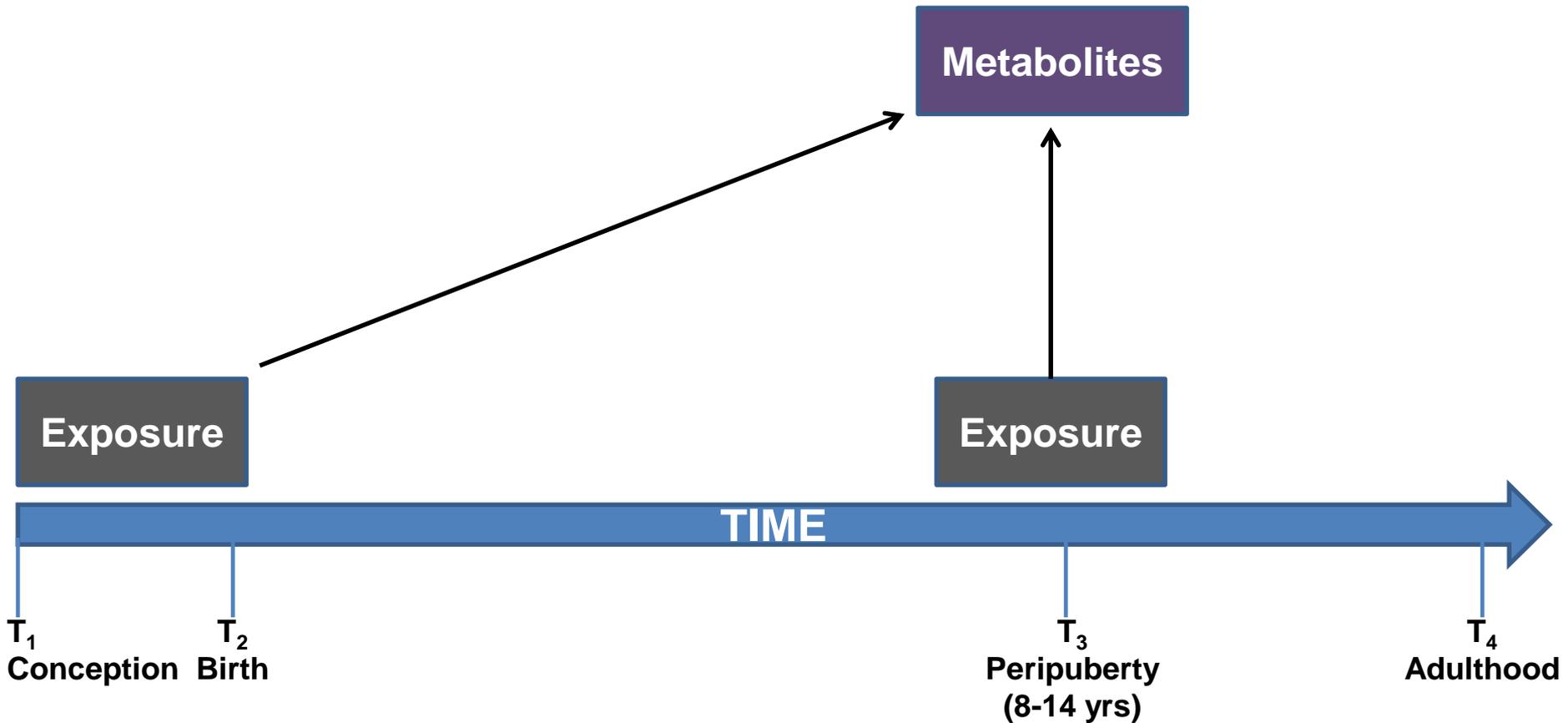


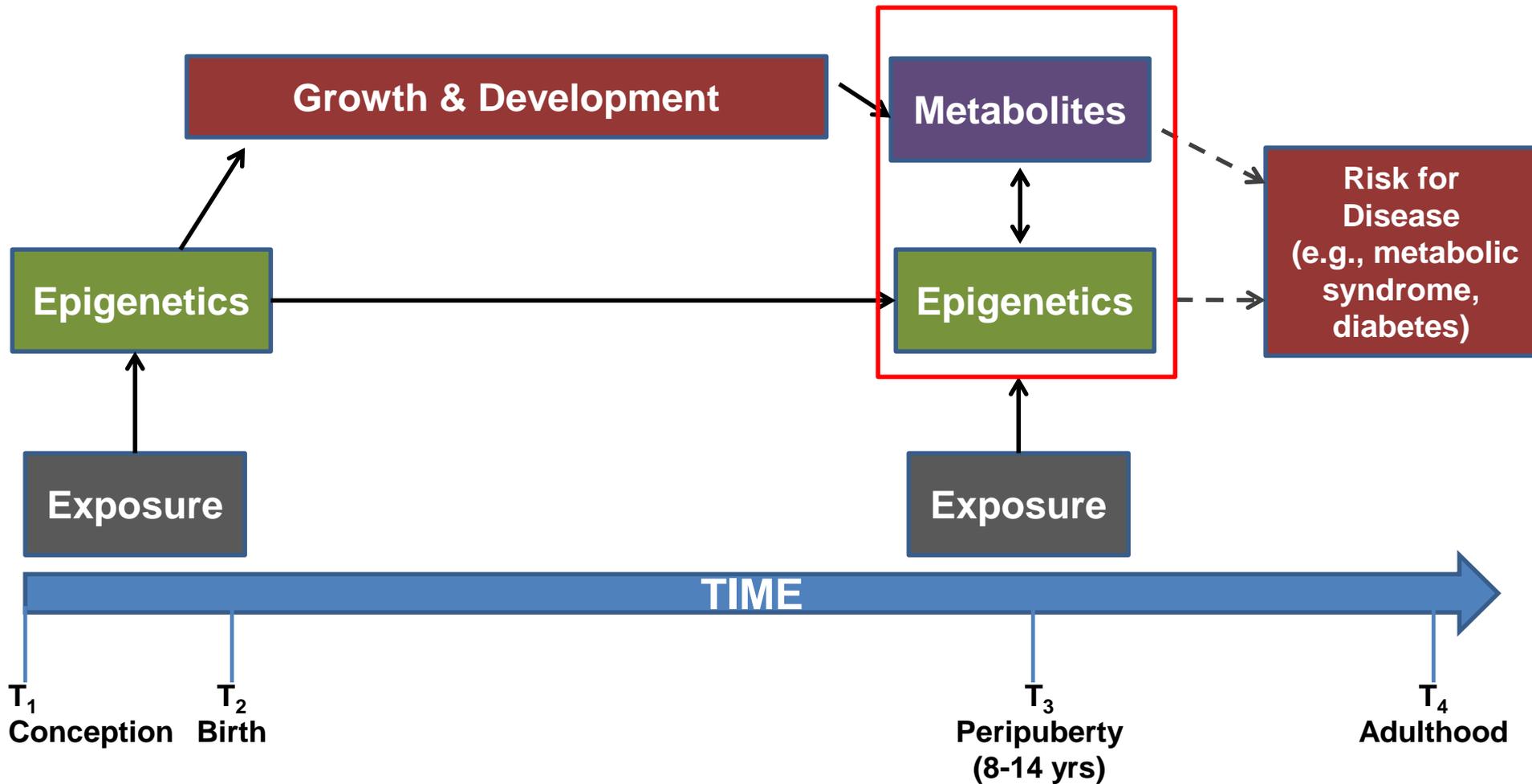
Figure created by Myriam Afeiche

# Metabolites may be affected by Prenatal and Concurrent Exposures

To identify metabolites out of 10K features affected by exposures



# Some Hypothesized Relationships between Exposures, Epigenetics, Metabolites, & Outcomes

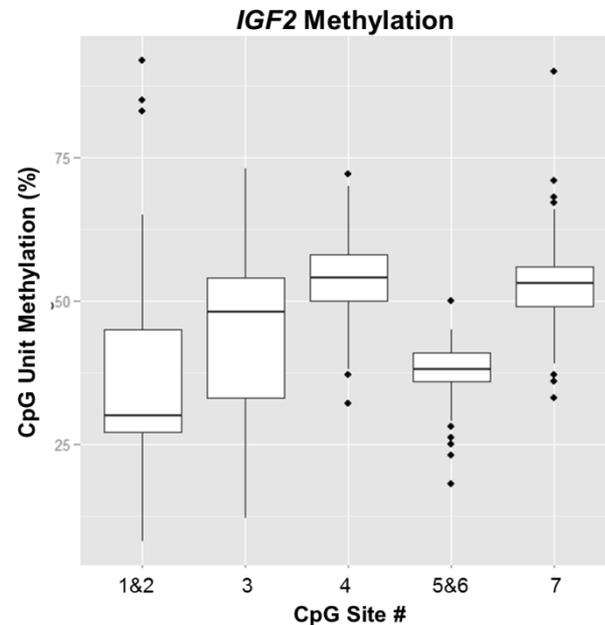
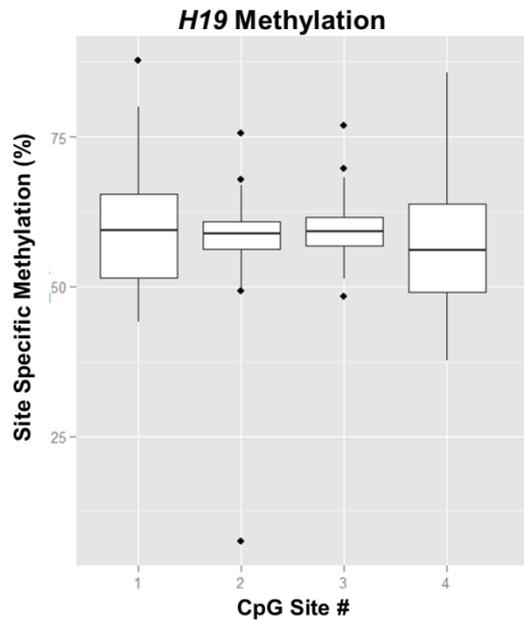
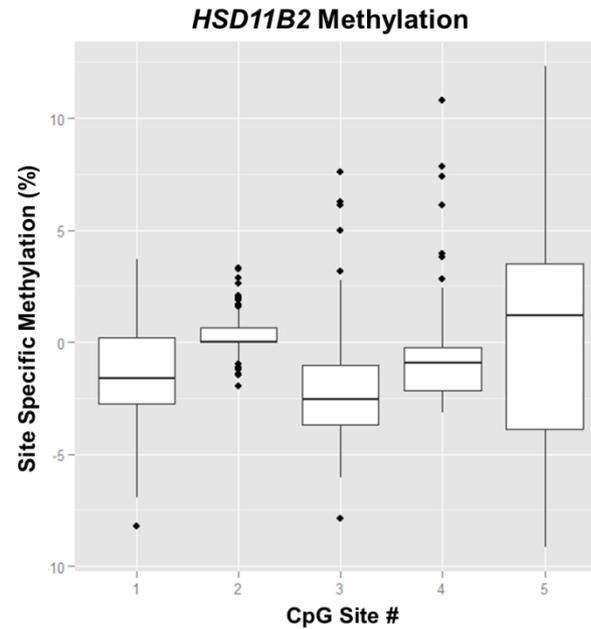
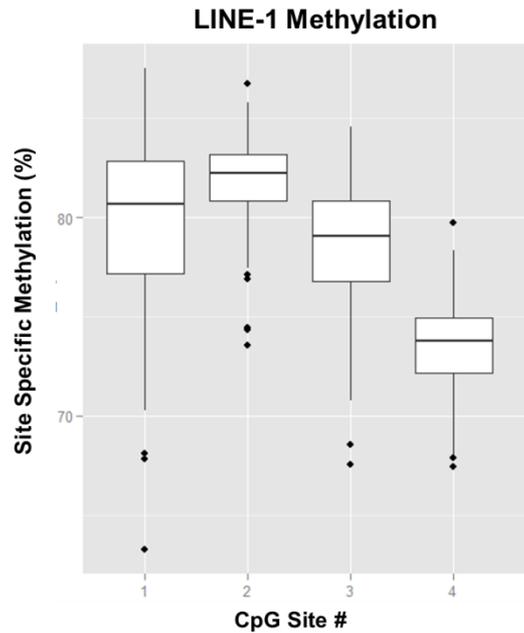


Epigenetic changes may occur from the early life exposure that can propagate through development, and influence metabolite levels in peri-puberty.

# DNA Methylation on Candidate Genes

- Methylation quantified in blood leukocyte DNA from peri-puberty of the ELEMENT cohort (n=250)
- Candidate genes/regions selected
  - *LINE-1*
    - Representative of global repetitive element methylation
    - Hypermethylation of repetitive elements necessary to suppress retrotransposition and maintain genomic stability
  - *IGF2* and *H19*
    - Reciprocally imprinted genes important for *in utero* growth
    - *IGF2* promotes growth, *H19* inhibits growth
  - *HSD11B2*
    - Protects against the growth-inhibitor, cortisol, during *in utero* development
- Methylation at these regions associated with *in utero* exposures

# Methylation Levels at the Candidate Genes



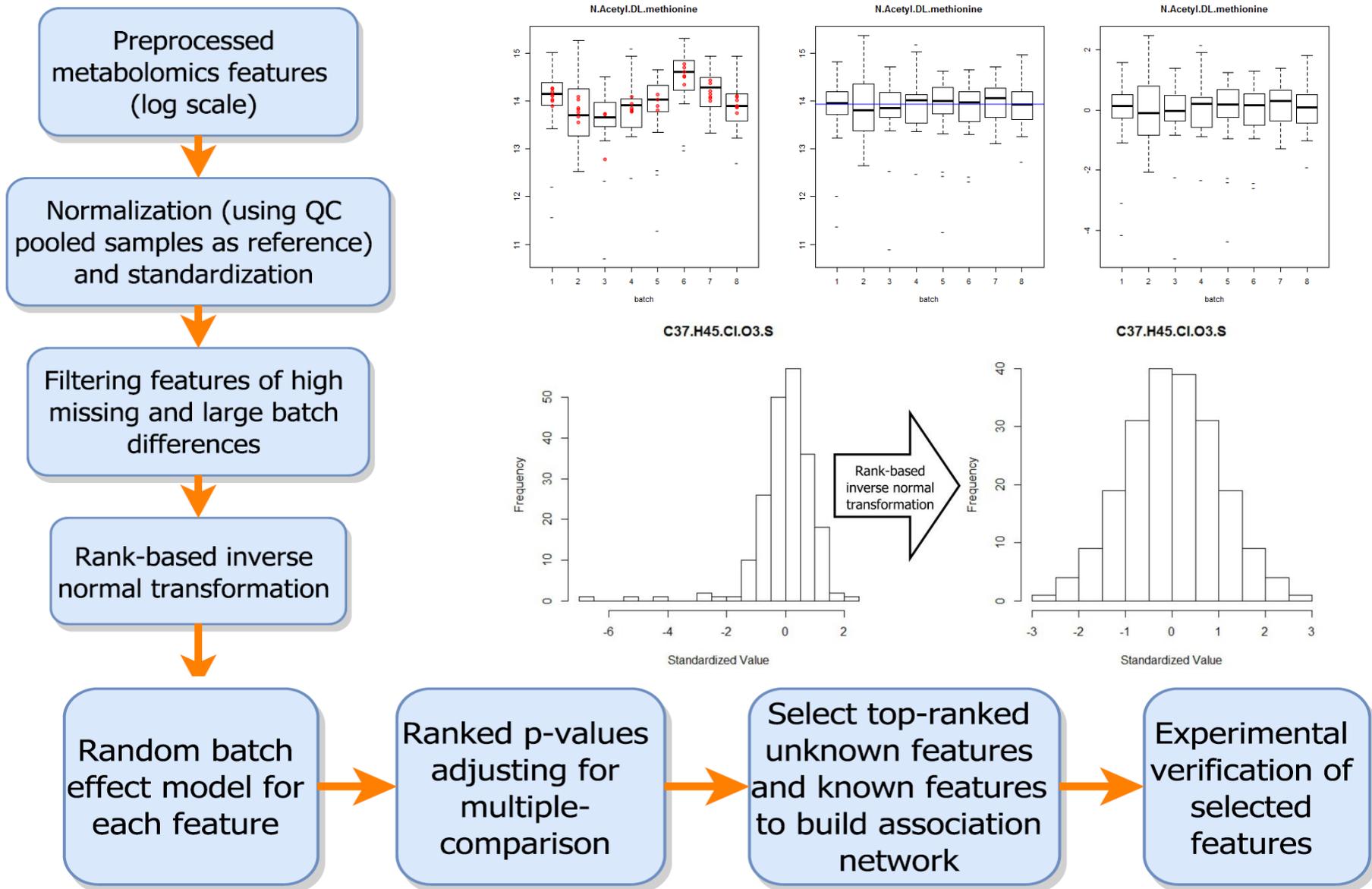
-Pyrosequencing used for  
LINE-1, *HSD11B2*, *H19*

-Sequenom EpiTYPER  
used for *IGF2*

-Data adjusted for  
experimental batch

-Wide intra-region variability  
across CpG sites with the  
exception of *H19*

# Metabolomics: Select Candidate Metabolomics Features



# High-dimensional Feature Screening

- **Random Batch Effects Model:**

$$\text{Metabolite}_i = \beta_0 + \beta_1 \text{Pb}^{\text{prenatal}} + \beta_2 \text{sex} + \beta_3 \text{age} + \beta_4 \text{Pb}^{\text{concurrent}} + b_i + \epsilon_i$$

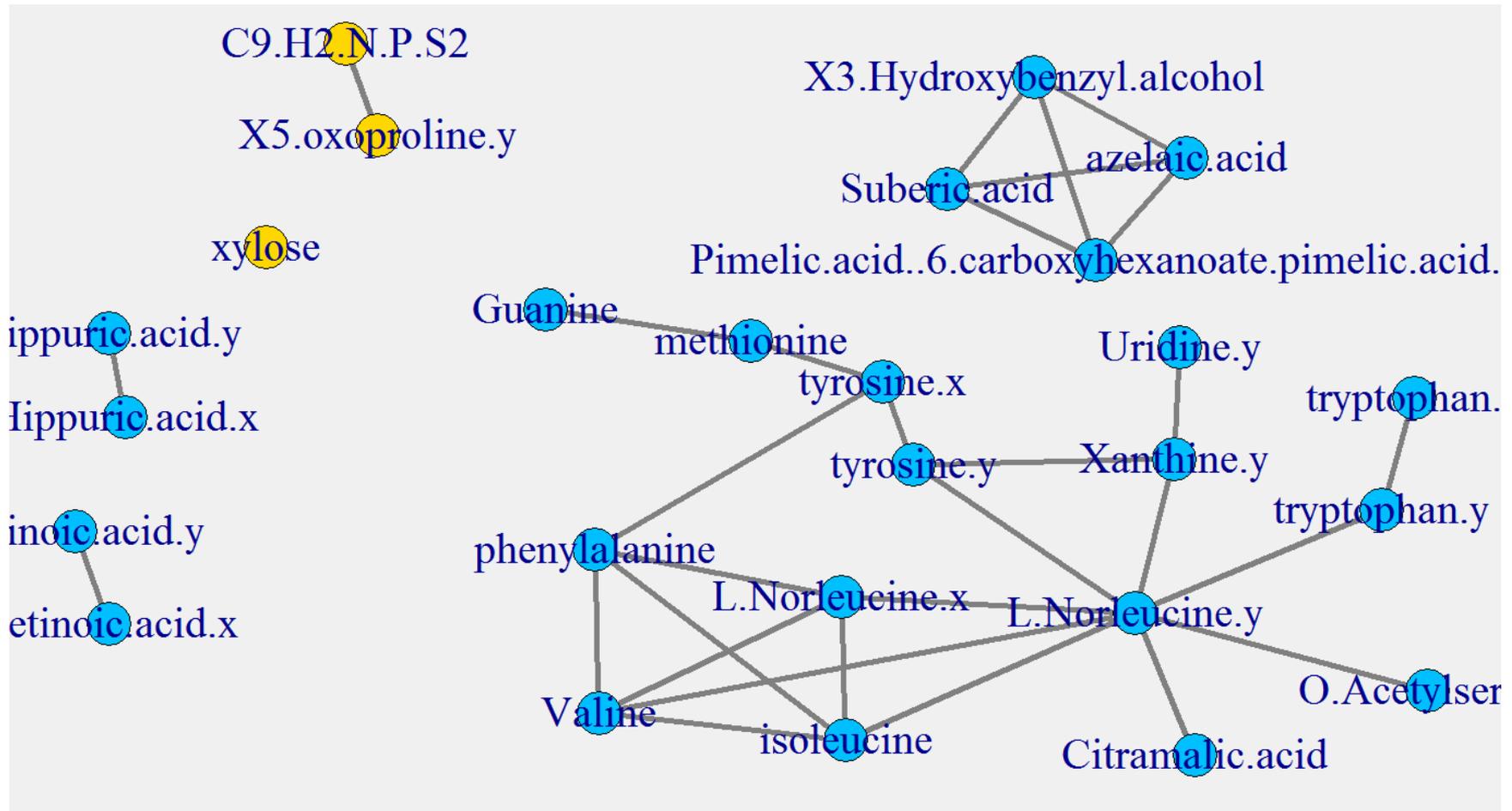
$i=1,2,\dots,8$  denotes the  $i$ -th batch

Prenatal Pb = maternal patella bone Pb

Concurrent Pb = blood Pb

- **Screening Test:**  $H_0: \beta_1=0$  (or  $H_0: \beta_4=0$ )
- **High-dimensional Problem:**  $\sim 10,000$  features are screened simultaneously
- **Significance level:**  $q$  value (FDR adjusted  $p$ -value) 0.05
- **Three metabolomics features are detected via the screening on  $H_0: \beta_1=0$**

# Three Candidates Detected in Network of Known Metabolite Features



# Statistical Models for Integrative Analysis of Concurrent Measures

## In Sex-Stratified Analyses:

$$(1) \text{metabolite}_i = \beta_0 + \beta_1 \text{age} + \beta_2 \text{methylation} + \beta_3 \text{Pb}^{\text{prenatal}} + b_i + \varepsilon_i$$

$$(2) \text{metabolite}_i = \beta_0 + \beta_1 \text{age} + \beta_2 \text{methylation} + \beta_3 \text{Pb}^{\text{prenatal}} + \beta_4 \text{Pb}^{\text{concurrent}} + b_i + \varepsilon_i$$

**Metabolites tested:** C9.H2.N.P.S2, X5.oxoproline, xylose

**Methylation:** percent methylation at one of the following:

- 1) 4 individual CpG sites in LINE-1
- 2) 5 individual CpG sites in *HSD11B2*
- 3) Average of all sites in *H19*
- 4) 5 CpG units in *IGF2* (some units are the average of 2 sites)

# DNA Methylation as a Predictor of Metabolites among Girls

p<0.1 associations:

**C9.H2.N.P.S2 Models**

LINE-1 #1	0.00 [-0.06, 0.06]
LINE-1 #2	0.00 [-0.13, 0.12]
LINE-1 #3	-0.01 [-0.09, 0.07]
LINE-1 #4	0.05 [-0.03, 0.14]
HSD11B2 #1	0.04 [-0.07, 0.14]
HSD11B2 #2	0.04 [-0.19, 0.26]
HSD11B2 #3	0.02 [-0.08, 0.12]
HSD11B2 #4	-0.06 [-0.20, 0.08]
HSD11B2 #5	-0.03 [-0.08, 0.02]
IGF2 #1&2	0.00 [-0.01, 0.02]
IGF2 #3	0.01 [-0.02, 0.01]
IGF2 #4	0.02 [-0.02, 0.05]
IGF2 #5&6	0.05 [-0.01, 0.11] *
IGF2 #7	0.03 [-0.01, 0.06]
H19 Average #1-4	-0.01 [-0.04, 0.03]

IGF2 CpGs 5&6 and ↑C9.H2.N.P.S2

**Oxoproline Models**

LINE-1 #1	-0.02 [-0.08, 0.04]
LINE-1 #2	-0.09 [-0.20, 0.02]
LINE-1 #3	-0.03 [-0.11, 0.04]
LINE-1 #4	-0.01 [-0.09, 0.08]
HSD11B2 #1	0.06 [-0.04, 0.16]
HSD11B2 #2	0.02 [-0.19, 0.22]
HSD11B2 #3	0.03 [-0.06, 0.12]
HSD11B2 #4	-0.06 [-0.16, 0.04]
HSD11B2 #5	0.01 [-0.03, 0.06]
IGF2 #1&2	0.00 [-0.01, 0.02]
IGF2 #3	-0.01 [-0.02, 0.01]
IGF2 #4	0.02 [-0.01, 0.05]
IGF2 #5&6	0.04 [-0.01, 0.09]
IGF2 #7	0.00 [-0.03, 0.03]
H19 Average #1-4	-0.02 [-0.05, 0.02]

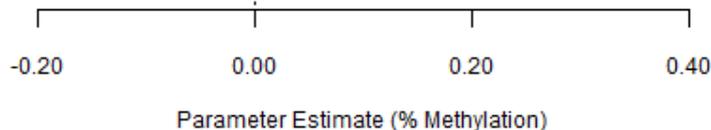
LINE-1 CpG 3 and ↓Xylose

**Xylose Models**

LINE-1 #1	-0.01 [-0.07, 0.04]
LINE-1 #2	-0.07 [-0.17, 0.04]
LINE-1 #3	-0.07 [-0.14, -0.01] **
LINE-1 #4	0.01 [-0.07, 0.09]
HSD11B2 #1	0.07 [-0.02, 0.15]
HSD11B2 #2	0.16 [-0.03, 0.34] *
HSD11B2 #3	-0.02 [-0.10, 0.06]
HSD11B2 #4	-0.10 [-0.20, 0.00] **
HSD11B2 #5	0.01 [-0.04, 0.05]
IGF2 #1&2	0.00 [-0.01, 0.01]
IGF2 #3	-0.01 [-0.02, 0.01]
IGF2 #4	0.02 [-0.02, 0.05]
IGF2 #5&6	0.03 [-0.02, 0.08]
IGF2 #7	0.01 [-0.01, 0.04]
H19 Average #1-4	0.00 [-0.04, 0.03]

HSD11B2 CpG 2 and ↑Xylose

HSD11B2 CpG 4 and ↓Xylose



**C9.H2.N.P.S2 Models**

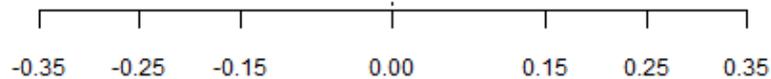
LINE-1 #1	0.02	[-0.03, 0.07]
LINE-1 #2	-0.02	[-0.12, 0.07]
LINE-1 #3	0.00	[-0.07, 0.06]
LINE-1 #4	0.01	[-0.09, 0.10]
HSD11B2 #1	-0.04	[-0.14, 0.07]
HSD11B2 #2	0.12	[-0.11, 0.35]
HSD11B2 #3	0.00	[-0.08, 0.09]
HSD11B2 #4	0.04	[-0.08, 0.16]
HSD11B2 #5	0.00	[-0.04, 0.05]
IGF2 #1&2	-0.01	[-0.02, 0.01]
IGF2 #3	0.01	[-0.01, 0.02]
IGF2 #4	-0.01	[-0.04, 0.01]
IGF2 #5&6	0.00	[-0.04, 0.04]
IGF2 #7	0.00	[-0.02, 0.03]
H19 Average #1-4	-0.03	[-0.07, 0.02]

**Oxoprolin Models**

LINE-1 #1	0.01	[-0.04, 0.06]
LINE-1 #2	-0.01	[-0.10, 0.08]
LINE-1 #3	0.01	[-0.05, 0.07]
LINE-1 #4	0.03	[-0.07, 0.12]
HSD11B2 #1	0.01	[-0.08, 0.11]
HSD11B2 #2	0.11	[-0.11, 0.33]
HSD11B2 #3	0.06	[-0.03, 0.14]
HSD11B2 #4	0.04	[-0.08, 0.16]
HSD11B2 #5	0.01	[-0.04, 0.05]
IGF2 #1&2	-0.01	[-0.02, 0.01]
IGF2 #3	0.01	[-0.01, 0.02]
IGF2 #4	-0.02	[-0.04, 0.01]
IGF2 #5&6	-0.01	[-0.05, 0.04]
IGF2 #7	0.00	[-0.02, 0.03]
H19 Average #1-4	-0.02	[-0.06, 0.02]

**Xylose Models**

LINE-1 #1	0.04	[-0.01, 0.10]
LINE-1 #2	0.03	[-0.07, 0.13]
LINE-1 #3	0.01	[-0.06, 0.08]
LINE-1 #4	0.01	[-0.09, 0.12]
HSD11B2 #1	0.01	[-0.10, 0.12]
HSD11B2 #2	-0.10	[-0.35, 0.14]
HSD11B2 #3	0.03	[-0.06, 0.13]
HSD11B2 #4	0.06	[-0.08, 0.19]
HSD11B2 #5	-0.03	[-0.09, 0.02]
IGF2 #1&2	0.01	[0.00, 0.03] *
IGF2 #3	0.01	[-0.01, 0.02]
IGF2 #4	0.01	[-0.02, 0.04]
IGF2 #5&6	0.02	[-0.02, 0.07]
IGF2 #7	0.01	[-0.02, 0.04]
H19 Average #1-4	-0.03	[-0.08, 0.02]



Parameter Estimate (% Methylation)

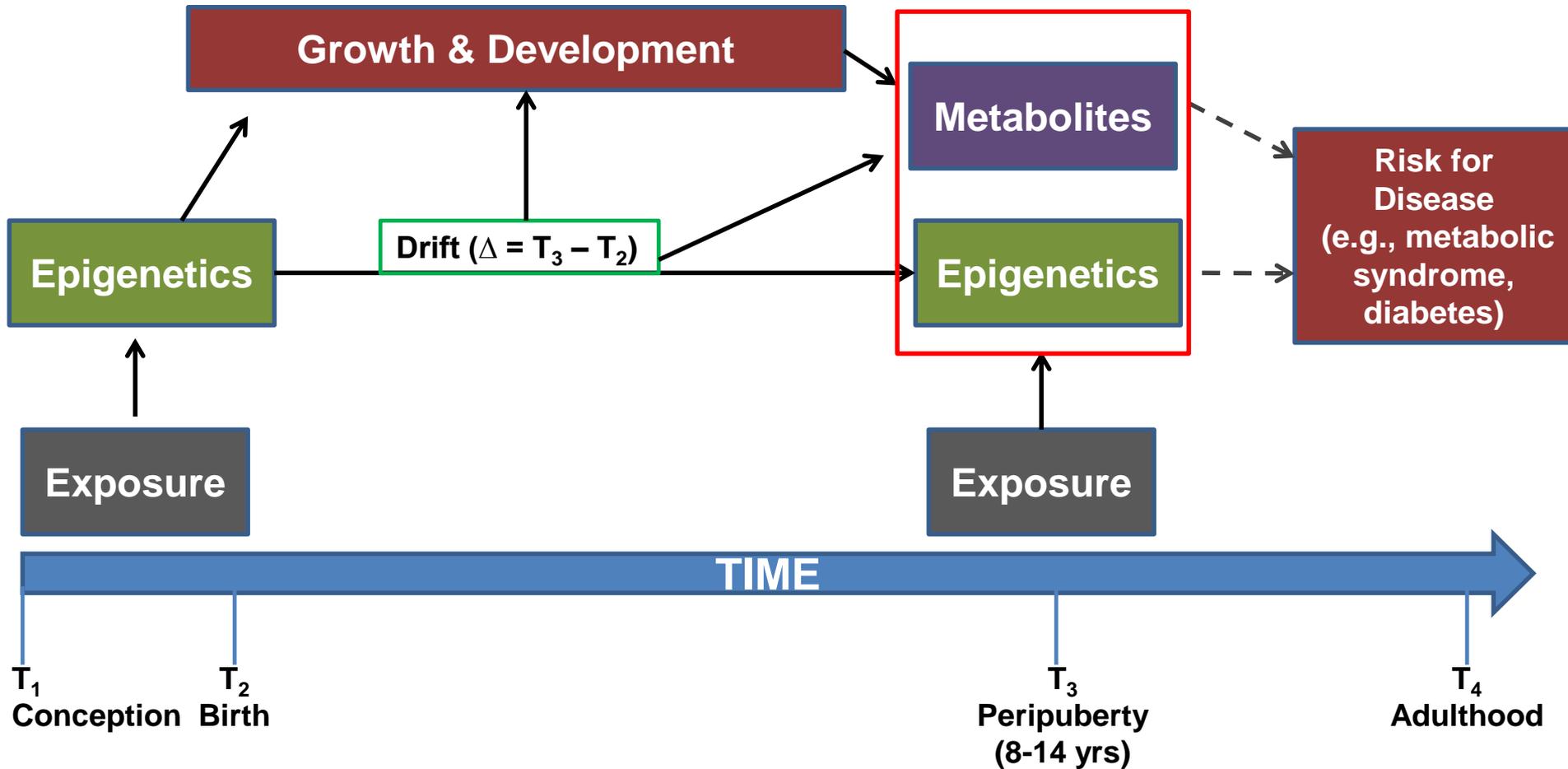
# DNA Methylation as a Predictor of Metabolites among Boys

p<0.1 association:

IGF2 CpG 1&2 and  
↑Xylose

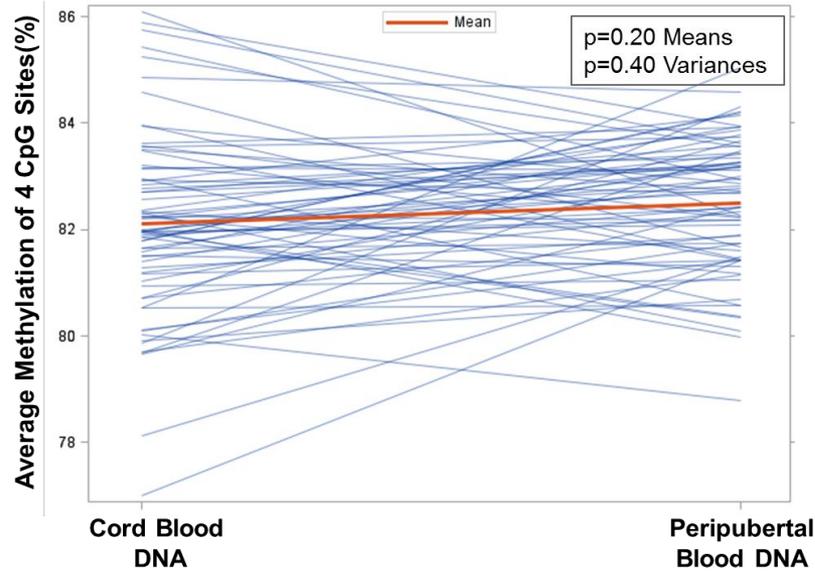
# Does Epigenetic Drift Play a Role?

Epigenetic changes from the early life exposure propagate through development, and influence metabolites in peri-puberty

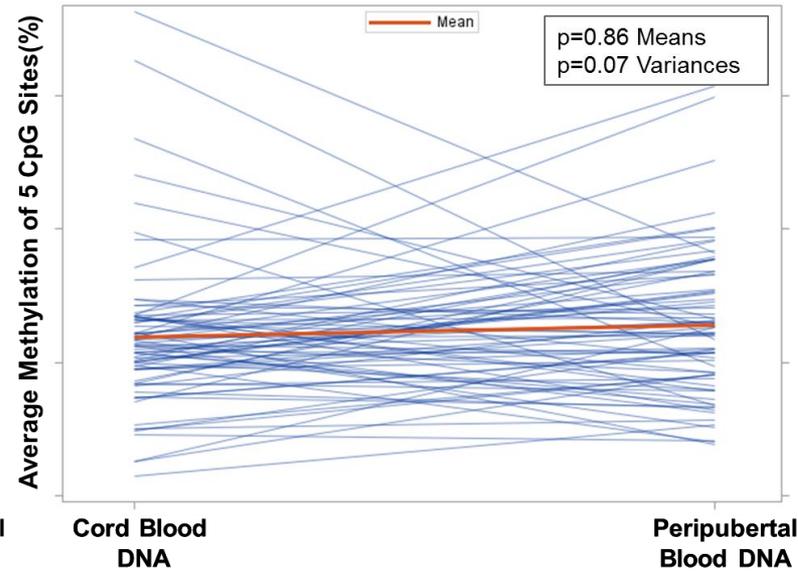


# DNA Methylation Drift in the ELEMENT Cohort (n=78)

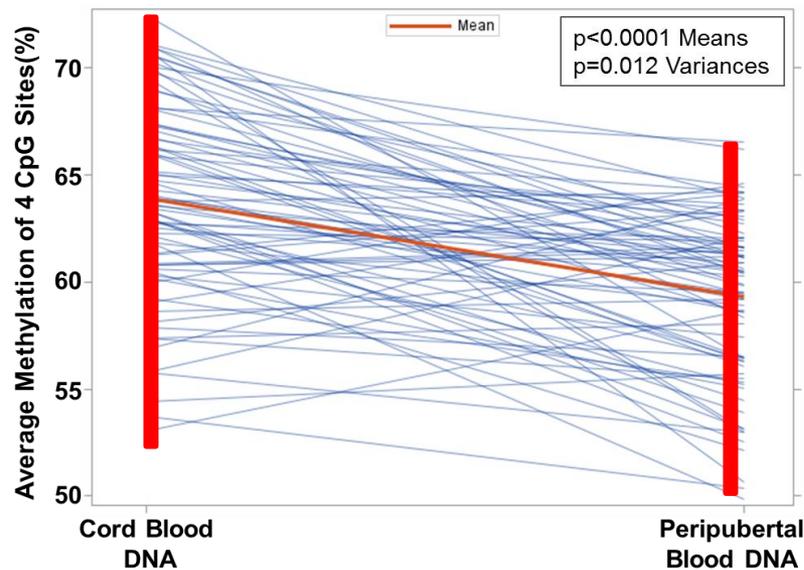
## LINE-1 Methylation



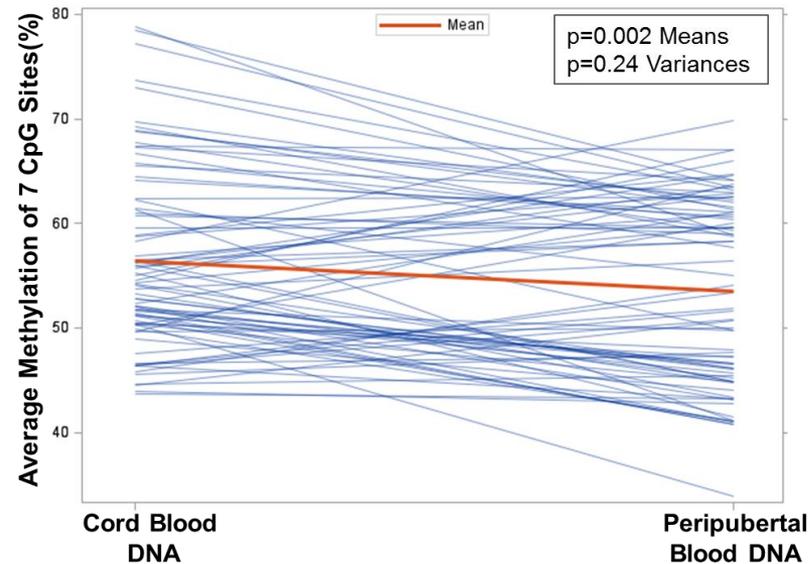
## HSD11B2 Methylation



## H19 Methylation



## IGF2 Methylation



*-H19* methylation mean and variance significantly decreased with time.

*-IGF2* methylation significantly increased with time.

# Incorporating Epigenetic Drift

$$\text{(Drift \#1) metabolite}_i = \beta_0 + \beta_1 \text{ age} + \beta_2 \Delta \text{methylation} + \beta_3 \text{Pb}^{\text{prenatal}} + b_i + \varepsilon_i$$

$$\begin{aligned} \text{(Drift \#2) metabolite}_i = & \beta_0 + \beta_1 \text{ age} + \beta_2 \Delta \text{methylation} + \beta_3 \Delta \text{variability} \\ & + \beta_4 \text{Pb}^{\text{prenatal}} + b_i + \varepsilon_i \end{aligned}$$

**Metabolites tested:** C9.H2.N.P.S2, X5.oxoproline, xylose

$\Delta$  **methylation:** (methylation<sub>j</sub> at t<sub>3</sub> – mean methylation at t<sub>3</sub>) - (methylation<sub>j</sub> at t<sub>2</sub> – mean methylation at t<sub>2</sub>)

$\Delta$  **variability:** (methylation<sub>j</sub> at t<sub>3</sub> – mean methylation at t<sub>3</sub>)<sup>2</sup> - (methylation<sub>j</sub> at t<sub>2</sub> – mean methylation at t<sub>2</sub>)<sup>2</sup>

**t**<sub>2</sub> = birth (cord blood leukocyte DNA)

**t**<sub>3</sub> = peri-puberty (blood leukocyte DNA)

# Association of Methylation Drift on Metabolites

- Methylation Drift:

Among those that passed statistical test of a significant mean change between two times, methylation changes of *IGF2#1 & 2* and *IFG2#7* are associated with *xylose*.

- Methylation Variability Drift:

Among those that passed statistical test of a significant variance change between two times, the change of variability of *IGF2 5&6* is associated with *xylose*.

# Concluding Remarks

- Integrative analysis involves high-dimensional data, in which most of them are noise. To reduce false discoveries, good data is of most importance.
- Statistical design of cohort studies remains a difficult problem, in particular there are many confounding factors involved in such studies.
- Multiple steps are required in data processing and data analysis, which incurs accumulation of human errors along the process, and any findings must be put for validation.
- It remains unknown whether or not, if so and how, to combine site-specific methylations or combine metabolite features.

# Future Directions

- Apply structural equation model for high-dimensional mediators to assess mediation effect of biomarkers.
- Utilize data mining techniques to identify biomarkers sensitive to past exposures and predictive to outcomes related to exposures.
- Derive epigenetic or metabolomics change as markers of cumulative exposures or cumulating risk for disease development.
- Analyze genome-wide methylation (being collected by the 450K platform through the P01 Admin Supplement) and full metabolomics data to find new biomarkers or mechanistic pathways.

# University of Michigan CEHC Team

- **CEHC Directors:** Karen E. Peterson, Vasantha Padmanabhan
- **Project/Core Leaders:** Dana Dolinoy, John Meeker, Alison Miller, Peter Song
- **Instituto Nacional de Salud Pública PI:** Mara Tellez-Rojo
- **Investigators/Consultants:** Alejandra Cantoral, Jorge Chavarro, Adrienne Ettinger, Howard Hu, Joyce Lee, Sub Pennathur, Lourdes Schnaas, Brisa Sánchez
- **Center Staff:** Lindsey Mitchell, Tamara Jones, Samantha Milewski
- **Postdoctoral Fellows:** Christopher Faulk, Kelly Ferguson, Jaclyn Goodrich, Deborah Watkins
- **Graduate Students:** Emily Hector, Joe Kochmanski, Lisa Marchlewicz, Meghan Moynihan, Lu Tang, Zhenzhen Zhang
- **INSP Field Staff**

## Acknowledgements:

Jackie Goodrich, Lu Tang, Emily Hector, Lisa Marchlewicz, Karen Peterson, Dana Dolinoy,

**Michigan Regional Comprehensive**

**Metabolomics Resource Core :**

Alla Karnovosky, Charles Burant