

Integrated Science Assessment for Lead

Appendix 7: Hematological Effects

External Review Draft

March 2023

Health and Environmental Effects Assessment Division
Center for Public Health and Environmental Assessment
Office of Research and Development
U.S. Environmental Protection Agency

DISCLAIMER

This document is an external review draft for peer review purposes only. This information is distributed solely for the purpose of predissemination peer review under applicable information quality guidelines. It has not been formally disseminated by the Environmental Protection Agency. It does not represent and should not be construed to represent any agency determination or policy. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.

DOCUMENT GUIDE

This Document Guide is intended to orient readers to the organization of the Lead (Pb) Integrated Science Assessment (ISA) in its entirety and to the sub-section of the ISA at hand (indicated in bold). The ISA consists of the Front Matter (list of authors, contributors, reviewers, and acronyms), Executive Summary, Integrated Synthesis, and 12 appendices, which can all be found at <https://cfpub.epa.gov/ncea/isa/recordisplay.cfm?deid=357282>.

Front Matter

Executive Summary

Integrative Synthesis

Appendix 1. Lead Source to Concentration

Appendix 2. Exposure, Toxicokinetics, and Biomarkers

Appendix 3. Nervous System Effects

Appendix 4. Cardiovascular Effects

Appendix 5. Renal Effects

Appendix 6. Immune System Effects

Appendix 7. Hematological Effects

Appendix 8. Reproductive and Developmental Effects

Appendix 9. Effects on Other Organ Systems and Mortality

Appendix 10. Cancer

Appendix 11. Effects of Lead in Terrestrial and Aquatic Ecosystems

Appendix 12. Process for Developing the Pb Integrated Science Assessment

CONTENTS

LIST OF TABLES	7-v
LIST OF FIGURES	7-vi
ACRONYMS AND ABBREVIATIONS	7-vii
APPENDIX 7 HEMATOLOGICAL EFFECTS	7-1
7.1 Introduction, Summary of the 2013 Integrated Science Assessment, and Scope of the Current Review	7-1
7.1.1. Red Blood Cell Survival and Function	7-2
7.1.2. Heme Synthesis	7-2
7.2 Scope	7-3
7.3 Red Blood Cell Survival and Function	7-5
7.3.1. Epidemiologic Studies of Red Blood Cell Survival and Function	7-5
7.3.2. Toxicological Studies of Red Blood Cell Survival and Function	7-7
7.3.3. Integrated Summary of Red Blood Cell Survival and Function	7-9
7.4 Heme Synthesis	7-9
7.4.1. Epidemiologic Studies of Heme Synthesis	7-10
7.4.2. Toxicological Studies of Heme Synthesis	7-10
7.4.3. Integrated Summary of Heme Synthesis	7-11
7.5 Biological Plausibility	7-11
7.5.1. Decreased Red Blood Cell Survival and Function	7-13
7.5.2. Altered Heme Synthesis	7-14
7.6 Summary and Causality Determination	7-16
7.6.1. Causality Determination for Red Blood Cell Survival and Function	7-16
7.6.2. Red Blood Cell Survival and Function	7-16
7.6.3. Heme Synthesis	7-17
7.6.4. Causality Determination	7-18
7.7 Evidence Inventories—Data Tables to Summarize Study Details	7-24
7.8 References	7-32

LIST OF TABLES

Table 7-1	Summary of evidence indicating a causal relationship between Pb exposure and hematological effects _____	7-19
Table 7-2	Epidemiologic studies of exposure to Pb and hematological effects _____	7-24
Table 7-3	Animal toxicological studies of Pb exposure and hematological effects _____	7-29

LIST OF FIGURES

Figure 7-1	Potential biological plausibility pathways for hematological effects associated with exposure to Pb	7-12
------------	---	------

ACRONYMS AND ABBREVIATIONS

ALAD	δ-aminolevulinate dehydratase	NCE	normochromatic red blood cells
ALA	aminolevulinic acid	PCV	packed cell volume
AQCD	Air Quality Criteria Document	PECOS	Population, Exposure, Comparison, Outcome, and Study Design
BLL	blood lead level	Plt	platelets
BMI	body mass index	PND	postnatal day
BW	body weight	PS	phosphatidylserine
Ca ²⁺	calcium	Q	quartile
CAT	catalase	RBC	red blood cell
CI	confidence interval	RDW	red blood cell distribution width
e-waste	electronic waste	ROS	reactive oxygen species
EPA	Environmental Protection Agency	SD	Sprague Dawley
EPO	erythropoietin	SES	socioeconomic status
Fe ²⁺	iron ion	SOD	superoxide dismutase
GFAAS	graphite furnace atomic absorption spectroscopy	O ₂ -	superoxide
GPx	glutathione peroxidase	wk	week, weeks
GR	glutathione reductase	yr	year, years
GSH	glutathione	Zn	zinc
GSSG	glutathione disulfide	ZPP	zinc-protoporphyrin
Hb	hemoglobin		
Hct	hematocrit		
HSC	hematopoietic stem cell		
OH	hydroxyl radical		
h	hour, hours		
H ₂ O ₂	hydrogen peroxide		
ICP-MS	inductively coupled plasma mass spectrometry		
KNHANES	Korean National Health and Nutrition Examination Survey		
ln	natural logarithm		
M	male		
M/F	male/female		
MCH	mean corpuscular hemoglobin		
MCHC	mean corpuscular hemoglobin concentration		
MCV	mean corpuscular volume		
MDA	malondialdehyde		
Mg ²⁺	magnesium ion		
mo	month, months		
NAAQS	National Ambient Air Quality Standards		
NCE	normochromatic red blood cell		
NR	not reported		
OR	odds ratio		
Pb	lead		
PCE	polychromatic red blood cells		

APPENDIX 7 HEMATOLOGICAL EFFECTS

Causality Determination for Pb Exposure and Hematological Effects

This appendix characterizes the scientific evidence that supports the causality determination for lead (Pb) exposure and hematological effects. The types of studies evaluated within this appendix are consistent with the overall scope of the ISA as detailed in the Process Appendix (see Section 12.4). In assessing the overall evidence, the strengths and limitations of individual studies were evaluated based on scientific considerations detailed in Table 12-5 of the Process Appendix (Section 12.6.1). More details on the causal framework used to reach these conclusions are included in the Preamble to the ISA ([U.S. EPA 2015](https://cfpub.epa.gov/ncea/isa/recordisplay.cfm?deid=357282)). The evidence presented throughout this appendix supports the following causality determination:

Outcome	Causality Determination
Altered Heme Synthesis and Decreased Red Blood Cell Survival and Function	Causal

The Executive Summary, Integrated Synthesis, and all other appendices of this Pb ISA can be found at <https://cfpub.epa.gov/ncea/isa/recordisplay.cfm?deid=357282>.

7.1 Introduction, Summary of the 2013 Integrated Science Assessment, and Scope of the Current Review

Hematology is a subgroup of clinical pathology concerned with the morphology, physiology, and pathology of blood and blood-forming tissues. Hematological measures, when evaluated with information on other biomarkers, are informative diagnostic tests for blood-forming tissues (i.e., bone marrow, spleen, and liver) and organ function.

The 2013 Integrated Science Assessment for Lead (hereinafter referred to as the 2013 Pb ISA) issued causality determinations for hematological effects resulting from lead (Pb) exposure on red blood cell (RBC) survival and function and altered heme synthesis ([U.S. EPA 2013](https://cfpub.epa.gov/ncea/isa/recordisplay.cfm?deid=357282)). The evidence underpinning these causality determinations is summarized below. Given the interconnectedness of the effects of Pb on RBC survival and function and altered heme synthesis, this assessment presents a single causality determination for Pb exposure and hematological effects. This approach allows for a more holistic evaluation of inter-related health endpoints, including a discussion of how all individual lines of evidence contribute to the overall hematological effects causality determination.

7.1.1. Red Blood Cell Survival and Function

1 The body of epidemiologic and toxicological evidence assessed in the 2013 Pb ISA indicates a
2 “causal” relationship between Pb exposure and decreased RBC survival and function. Experimental
3 animal studies demonstrate that relevant human blood Pb levels (BLLs) from oral and inhalation exposure
4 alter several hematological parameters (e.g., RBC number), increase measures of oxidative stress (e.g.,
5 inhibition of antioxidant enzymes in RBCs), and increase cytotoxicity in RBC precursor cells. Some of
6 these effects have been observed in animal toxicological studies with exposures resulting in BLLs of 2–
7 7 µg/dL. Evidence of biologically plausible modes of action, including increased intracellular calcium
8 concentrations $[Ca^{2+}]_i$, decreased Ca^{2+}/Mg^{2+} ATPase activity, and increased phosphatidylserine (PS)
9 exposure leading to RBC destruction by macrophages, support these findings. Epidemiologic studies
10 reported associations between exposure to Pb, BLL, and altered hematological endpoints, increased
11 measures of oxidative stress, and altered hematopoiesis in adults and children. Although most of these
12 studies are limited by their lack of rigorous methodology (i.e., correlations, t tests, or chi squared
13 analyses), some studies in children did adjust for potential confounding factors, including age, sex,
14 mouthing behavior, anemia, dairy product consumption, maternal age, education, employment, marital
15 status, family structure, and socioeconomic status (SES)-related variables. Though limited in number,
16 studies that adjusted for confounders also reported consistent associations between BLL and altered
17 hematological parameters, strengthening their support for findings in experimental animals. Collectively,
18 the strong evidence from toxicological studies supported by findings from mode of action and
19 epidemiologic studies reviewed in the 2013 Pb ISA was sufficient to conclude that there is a causal
20 relationship between Pb exposures and decreased RBC survival and function.

7.1.2. Heme Synthesis

21 Available toxicological evidence evaluated in the 2013 Pb ISA indicated a causal relationship
22 between Pb exposure and altered heme synthesis ([U.S. EPA 2013](#)). Altered heme synthesis is
23 demonstrated by a small but consistent body of studies in adult animals, which report that exposures that
24 result in BLLs relevant to humans (e.g., 10 µg/dL) lead to decreased δ-aminolevulinate dehydratase
25 (ALAD) and ferrochelatase activities. Supporting this evidence is a larger body of ecotoxicological
26 studies that demonstrate decreased ALAD activity across a wide range of taxa exposed to Pb. Evidence of
27 biologically plausible modes of action, including evidence that Pb acts directly on two enzymes involved
28 in heme synthesis (ALAD and ferrochelatase), decreased RBC Hb concentration, measures of oxidative
29 stress, and evidence that administration of antioxidants reduced the effects of Pb exposure on antioxidant
30 enzymes, support these findings. Epidemiologic studies find associations in both adults and children
31 between higher BLLs and decreased ALAD and ferrochelatase activities. Although most of these studies
32 are limited by their lack of rigorous methodology and consideration of potential confounders, some
33 studies in children did incorporate potential confounding factors (i.e., age, sex, urban/rural residence,

height, weight, BMI [body mass index]). Although limited in number, studies that adjusted for confounders also reported consistent associations between BLLs and decreased ALAD and ferrochelatase, strengthening support for the findings in the animal toxicological studies. Evidence for altered heme synthesis is also provided by a large body of toxicological and epidemiologic studies that report decreased hemoglobin (Hb) concentrations in association with Pb exposure or BLL. The 2013 Pb ISA concluded that, collectively, the strong evidence from toxicological and ecotoxicological studies—supported by findings from epidemiologic studies—is sufficient to conclude a causal relationship between Pb exposures and altered heme synthesis.

This ISA determined causality for adverse effects of Pb exposure on altered heme synthesis and decreased red blood cell survival and function. Recent evidence demonstrates that Pb exposures alter several hematological parameters, decrease enzyme activity related to heme synthesis, and increase RBC oxidative stress. Biological plausibility is provided by toxicological and epidemiologic studies demonstrating increased intracellular calcium concentrations, decreased $\text{Ca}^{2+}/\text{Mg}^{2+}$ ATPase activity, and increased PS exposure. Taken together, there is sufficient evidence to conclude that there is a causal relationship between Pb exposure and hematological effects, including altered heme synthesis and decreased RBC survival and function.

The following sections provide an overview of the scope of the appendix (Section 7.2), evaluation of the scientific evidence relating Pb exposures and hematological effects (Sections 7.3 and 7.4), a discussion of biological plausibility (Section 7.5), and a summary section with the updated causality determination (Section 7.6, Table 7-1). The focus of these sections is on studies published since the completion of the 2013 Pb ISA ([U.S. EPA 2013](#)). Study-specific details, including animal type, exposure concentrations and exposure durations in experimental studies, and study design; exposure metrics; and select results in epidemiologic studies are presented in evidence inventories in Section 7.7.

7.2 Scope

The scope of this section is defined by Population, Exposure, Comparison, Outcome, and Study Design (PECOS) statements. The PECOS statement defines the objectives of the review and establishes study inclusion criteria, thereby facilitating identification of the most relevant literature to inform the Pb ISA.¹ To identify the most relevant literature, the body of evidence from the 2013 Pb ISA was considered in the development of the PECOS statements for this appendix. Specifically, well-established areas of research; gaps in the literature; and inherent uncertainties in specific populations, exposure metrics,

¹ The following types of publications are generally considered to fall outside the scope and are not included in the ISA: review articles (which typically present summaries or interpretations of existing studies rather than bringing forward new information in the form of original research or new analyses), Pb poisoning studies or clinical reports (e.g., involving accidental exposures to very high amounts of Pb described in clinical reports that may be extremely unlikely to be experienced under ambient air exposure conditions), and risk or benefits analyses (e.g., that apply concentration-response functions or effect estimates to exposure estimates for differing cases).

comparison groups, and study designs identified in the 2013 Pb ISA inform the scope of this appendix. The 2013 Pb ISA used different inclusion criteria than the current ISA, and the studies referenced therein often do not meet the current PECOS criteria (e.g., due to higher or unreported biomarker levels). Studies that were included in the 2013 Pb ISA, including many that do not meet the current PECOS criteria, are discussed in this appendix to establish the state of the evidence prior to this assessment. With the exception of supporting evidence used to demonstrate the biological plausibility of Pb-associated hematological effects, recent studies were only included if they satisfied all of the components of the following discipline-specific PECOS statements:

Epidemiologic Studies

Population: Any human population, including specific populations or lifestages that might be at increased risk of a health effect;

Exposure: Exposure to Pb¹ as indicated by biological measurements of Pb in the body, with a specific focus on Pb in blood, bone, and teeth; validated environmental indicators of Pb exposure²; or intervention groups in randomized trials and quasi-experimental studies;

Comparison: Populations, population subgroups, or individuals with relatively higher versus lower levels of the exposure metric (e.g., per unit or log unit increase in the exposure metric, or categorical comparisons between different exposure metric quantiles);

Outcome: Hematological effects including but not limited to disruption of heme synthesis and RBC survival and function; and

Study Design: Epidemiologic studies consisting of longitudinal and retrospective cohort studies, case-control studies, cross-sectional studies with appropriate timing of exposure for the health endpoint of interest, randomized trials, and quasi-experimental studies examining interventions to reduce exposures.

Experimental Studies

Population: Laboratory nonhuman mammalian animal species (e.g., mouse, rat, guinea pig, minipig, rabbit, cat, dog) of any lifestage (including preconception, in utero, lactation, peripubertal, and adult stages);

Exposure: Oral, inhalation or intravenous treatment(s) administered to a whole animal (in vivo) that results in a BLL of 30 mg/dL or below;^{3,4}

Comparators: A concurrent control group exposed to vehicle-only treatment or untreated control;

¹ Recent studies of occupational exposure to Pb were only considered insofar as they addressed a topic area that was relevant to the National Ambient Air Quality Standards review (e.g., longitudinal studies designed to examine recent versus historical Pb exposure).

² Studies that estimate Pb exposure by measuring Pb concentrations in particulate matter with a nominal mean aerodynamic diameter less than or equal to 10 μm^3 (PM₁₀) and particulate matter with a nominal mean aerodynamic diameter less than or equal to 2.5 μm^3 (PM_{2.5}) ambient air samples are only considered for inclusion if they also include a relevant biomarker of exposure. Given that size distribution data for Pb-PM are limited, it is difficult to assess the representativeness of these concentrations to population exposure [Section 2.5.3 ([U.S. EPA 2013](#))]. Moreover, data illustrating the relationships of Pb-PM₁₀ and Pb-PM_{2.5} with BLLs are lacking.

³ Pb mixture studies are included if they employ an experimental arm that involves exposure to Pb alone.

⁴ This level is approximately an order of magnitude above the upper end of the distribution of U.S. young children's BLLs. The 95th percentile of the 2011–2016 National Health and Nutrition Examination Survey distribution of BLL in children (1–5 years; n = 2,321) is 2.66 $\mu\text{g}/\text{dL}$ ([CDC 2019](#)) and the proportion of individuals with BLL that exceed this concentration varies depending on factors including (but not limited to) housing age, geographic region, and a child's age, sex and nutritional status.

Outcome: Hematological effects; and
Study design: Controlled exposure studies of animals in vivo.

7.3 Red Blood Cell Survival and Function

Toxicological and epidemiologic studies evaluated in the 2013 Pb ISA ([U.S. EPA 2013](#)) provided strong evidence that exposure to Pb affects a range of hematological outcomes related to RBC survival and function; which is consistent with epidemiologic evidence from the 2006 Pb Air Quality Criteria Document (AQCD) ([U.S. EPA 2006b](#)), demonstrating an association between high BLLs and anemia in children. Given the extensive evidence base at higher BLLs, the scope for this appendix focuses on toxicological studies conducted at lower exposure levels and epidemiologic studies in nonoccupational populations (as described in Section 7.2). Under the defined PECOS criteria, recent toxicological and epidemiologic studies provide additional support for Pb-related changes in Hb concentration and some other hematological measures of RBC survival and function. Below, recent evidence is reviewed in the context of evidence from past assessments.

7.3.1. Epidemiologic Studies of Red Blood Cell Survival and Function

The epidemiologic evidence evaluated in the 2013 Pb ISA ([U.S. EPA 2013](#)) covered a range of measures related to RBC survival and function, including RBC counts and other hematological parameters, hematopoiesis, $\text{Ca}^{2+}/\text{Mg}^{2+}$ ATPase activity, PS exposure, and RBC oxidative stress. Specifically, epidemiologic studies provided evidence that elevated BLLs in children and adults are associated with altered hematological parameters (e.g., decreased RBC counts, Hb concentration, and hematocrit [Hct] and changes in mean corpuscular volume [MCV] and mean corpuscular hemoglobin [MCH]), increased measures of oxidative stress (e.g., altered antioxidant enzyme activities [superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx)], decreased cellular glutathione (GSH), and increased lipid peroxidation), and altered hematopoiesis (e.g., decreased erythropoietin [EPO]). Notably, most of these epidemiologic studies are cross-sectional in design and conducted either in occupationally exposed populations or other populations with higher mean Pb exposures (i.e., BLLs $>10 \mu\text{g/dL}$). These studies were additionally limited by their lack of consideration of potential confounders, although some studies in children did adjust for a range of factors, including age, sex, mouthing behavior, anemia, dairy product consumption, maternal age, education, employment, marital status, family structure, and SES-related variables ([Queirolo et al. 2010](#); [Ahamed et al. 2007](#); [Riddell et al. 2007](#)). Studies that did account for potential confounders reported consistent associations between increased BLLs and decreased Hb levels, increased prevalence of anemia, and increased RBC oxidative stress (Sections 4.7.2.1 and 4.7.2.7 of the 2013 Pb ISA). As a whole, the cross-sectional study designs, higher exposures, and lack of rigorous statistical methodologies in many studies raise uncertainties in the epidemiologic evidence regarding the directionality of effects; the level, timing, frequency, and duration

1 of Pb exposure that contributed to the observed associations; and whether the observed associations are
2 independent of potential confounders.

3 Recent epidemiologic studies provide generally consistent evidence of associations between Pb
4 exposures and decreased Hb levels in children. These associations are reported at lower BLLs than in
5 studies included in the 2013 Pb ISA. Evidence for associations with other hematological parameters of
6 RBC function and survival, as well as Hb levels in adults, is less robust. Consistent with the 2013 Pb ISA,
7 most recent studies are cross-sectional analyses, which are unable to establish temporality between
8 exposure and outcome. However, recent studies include populations with lower mean BLLs and more
9 robust adjustment for potential confounders compared with studies included in the 2013 Pb ISA.
10 Measures of central tendency for BLLs used in each study, along with other study-specific details
11 including study population characteristics and select effect estimates, are highlighted in Table 7-2. An
12 overview of the recent evidence is provided below.

13 The most common hematological parameter evaluated in recent epidemiologic studies is Hb
14 levels. A number of cross-sectional studies of children in China observed decreases in Hb levels
15 associated with increases in blood Pb or erythrocyte Pb levels ([Guo et al. 2021](#); [Kuang et al. 2020](#); [Li et
16 al. 2018](#); [Liu et al. 2015](#); [Liu et al. 2012](#)). [Kuang et al. \(2020\)](#) and [Li et al. \(2018\)](#) also reported inverse
17 associations between BLLs and MCH in children, which is a measure of average Hb concentration in a
18 single erythrocyte. Notably, all of these studies had mean and/or median BLLs below 10 µg/dL, including
19 some below 5 µg/dL ([Guo et al. 2021](#); [Kuang et al. 2020](#); [Liu et al. 2012](#)). While only a few studies
20 attempted to account for potential confounding by SES ([Kuang et al. 2020](#); [Liu et al. 2015](#)), others
21 adjusted directly for iron deficiency ([Li et al. 2018](#); [Liu et al. 2012](#)), which may be the direct mechanism
22 by which SES could potentially confound the relationship between BLLs and Hb (i.e., via nutritional
23 deficiency). Although the magnitude of the observed effect estimates was not directly comparable across
24 studies, BLLs were negatively associated with Hb levels in all studies. Notably, [Liu et al. \(2012\)](#) reported
25 that a 1 µg/dL increase in BLLs was associated with a larger decrease in Hb levels when restricting their
26 sample to children with BLLs less than 10 µg/dL (−0.174 g/dL [95% confidence interval (CI): −0.27,
27 −0.078 g/dL]) compared with the full sample (−0.096 g/dL [95% CI: −0.18, −0.012 g/dL]). In studies that
28 examined quantiles of exposure, blood or erythrocyte Pb levels were only associated with Hb levels at the
29 higher quantiles ([Guo et al. 2021](#); [Liu et al. 2015](#)). For example, in a large hospital-based study in China,
30 children with BLLs between 3.33 and 4.50 µg/dL (quintile 4) and those with levels greater than
31 4.50 µg/dL (quintile 5) had decreases in Hb relative to children with BLLs less than 1.61 µg/dL
32 (−0.49 g/L [95% CI: −0.94, −0.04 g/L] and −1.25 g/L [95% CI: −1.71, −0.78 g/L], respectively);
33 however, null associations were reported for quintiles 2 (1.61–2.44 µg/dL) and 3 (2.44–3.33 µg/dL)
34 relative to the lowest quintile of exposure ([Guo et al. 2021](#)). While the clinical relevance of small mean
35 decrements in Hb across exposure quintiles is unclear, [Guo et al. \(2021\)](#) also reported monotonic
36 increases in the odds of anemia (defined as Hb levels below 110 g/L) in association with increasing blood
37 Pb quintiles, with a 45% (95% CI: 26%, 67%) increase in the odds of anemia for children in the highest

quintile of exposure relative to the lowest quintile. Similarly, [Li et al. \(2018\)](#) observed a 5% (95% CI: 0%, 11%) increase in the odds of decreased Hb levels (<115 g/L) per 1 µg/dL increase in blood Pb.

Recent studies of Pb exposure and Hb levels in adults are more limited in number and include overlapping study populations. In contrast to studies in children, [Park and Lee \(2013\)](#) reported increases in Hb associated with continuous increases in BLLs for adult participants of the 2008–2010 cycles of the Korean National Health and Nutrition Examination Survey (KNHANES). The analysis was stratified by sex and the observed associations were comparable for men and women. A similar study analyzed the same KNHANES cycles, but examined the population as a whole, rather than stratified by sex, and also categorized exposure into quartiles ([Kim and Lee 2013](#)). The authors noted similar positive associations between blood Pb and Hb levels. However, [Kim and Lee \(2013\)](#) observed inverse associations across exposure quartiles when correcting BLLs for Hct in order to estimate erythrocyte Pb. As described in the 2006 Pb AQCD ([U.S. EPA 2006b](#)), Pb exposure decreases Hct and MCV, meaning the negative effects of Pb can potentially decrease Pb levels in whole blood.

In addition to studies examining the relationship between Pb exposure and Hb levels, a few recent cross-sectional studies evaluate associations between BLLs and other hematological parameters. In a group of children in China, including some living near a battery plant or a lead/-zinc mine, [Li et al. \(2018\)](#) reported increased odds of low RBC counts and low blood platelets (Plt) in association with increased BLLs. In contrast, [Kuang et al. \(2020\)](#) observed a positive association with increased RBC counts in a convenience sample of slightly older boys in Nanjing, China, with notably lower median BLLs (2.61 µg/dL compared with 8.38 µg/dL). In an adult population, a cohort of pregnant women in Durango, Mexico, [La-Llave-León et al. \(2015\)](#) also observed a positive cross-sectional association between RBC counts and BLLs. Given this small body of studies that examine diverse populations with varying BLLs, it is difficult to discern methodological or demographic factors contributing to the inconsistent results.

7.3.2. Toxicological Studies of Red Blood Cell Survival and Function

As previously reported in the 2013 Pb ISA, epidemiologic evidence is coherent with experimental animal studies demonstrating that exposures via drinking water and oral gavage resulting in BLLs relevant to what was found in humans affect multiple hematological outcomes related to RBC survival and function ([U.S. EPA 2013](#)). Specifically, exposure to Pb has been shown to decrease RBC survival, either through direct effects on RBC membranes leading to increased fragility or through the induction of eryptosis and eventual phagocytosis by macrophages ([U.S. EPA 2013](#)). Some of these effects have been observed in animal toxicological studies with exposures resulting in 2–7 µg/dL BLL. For example, Hb concentrations in plasma was significantly decreased in male mice exposed to Pb nitrate (50 mg/kg BW in drinking water for 40 days; BLL: 1.72 ± 0.02 µg/dL) ([Sharma et al. 2010](#)). BLLs >100 µg/dL were also associated with decreased RBC survival in laboratory animals.

1 The evidence is limited and conflicting for the observed effects of Pb exposure on hematopoiesis
2 in rats and mice. For example, administration of Pb acetate (140, 250, or 500 mg/kg) via oral gavage once
3 per week for 10 weeks decreased the number of polychromatic RBCs (PCE) and increased numbers of
4 micronucleated PCEs in female rats ([Celik et al. 2005](#)). Increased micronucleated PCEs were reported in
5 female and male rats exposed to Pb acetate in drinking water for 125 days, but decreased
6 PCEs/normochromatic RBCs (NCEs) ratio was only observed in male rats ([Alghazal et al. 2008](#)).
7 However, in mice exposed to Pb acetate (1 g/L in drinking water for 90 days), PCE increased, but
8 PCE/NCE was unaffected ([Marques et al. 2006](#)).

9 In addition, the 2013 Pb ISA reported that Pb exposure significantly decreases several
10 hematological parameters. In studies reporting BLL relevant to this ISA, decreased RBC counts
11 ([Andjelkovic et al. 2019](#); [Cai et al. 2018](#); [Sharma et al. 2010](#)), Hb concentration ([Andjelkovic et al. 2019](#);
12 [Cai et al. 2018](#); [Berrahal et al. 2011](#); [Sharma et al. 2010](#); [Baranowska-Bosiacka et al. 2009](#); [Massó-](#)
13 [González and Antonio-García 2009](#)), and Hct ([Andjelkovic et al. 2019](#); [Massó-González and Antonio-](#)
14 [García 2009](#); [Massó et al. 2007](#)) were reported in laboratory studies conducted in rats. In other studies in
15 which BLLs were not reported, decreased RBC counts ([Simsek et al. 2009](#); [Marques et al. 2006](#); [Lee et al.](#)
16 [2005](#)), Hb ([Wang et al. 2010b](#); [Simsek et al. 2009](#); [Lee et al. 2005](#)), Hct ([Molina et al. 2011](#); [Marques et](#)
17 [al. 2006](#); [Lee et al. 2005](#)), MCV ([Wang et al. 2010b](#)), MCH ([Wang et al. 2010b](#); [Simsek et al. 2009](#)), and
18 mean corpuscular hemoglobin concentration (MCHC) ([Wang et al. 2010b](#); [Simsek et al. 2009](#)) were
19 reported in laboratory studies conducted in rats and mice. Some toxicological studies found no evidence
20 of hematological effects ([Gautam and Flora 2010](#); [Lee et al. 2006](#)).

21 Recent studies also report effects of Pb exposure on hematological parameters at BLLs relevant to
22 this ISA. Administration of Pb acetate in drinking water (0.2%; BLL = 9.3 ± 0.98 µg/dL) for 84 days
23 resulted in RBC hemolysis, and significantly decreased RBC lifespan and number, and Hb levels, but had
24 no effect on Plt number in blood collected from Sprague Dawley rats ([Cai et al. 2018](#)). In a different
25 study, administration of Pb acetate in drinking water (0.150 mg/kg; BLL = 14.7 µg/dL) for 1 day
26 decreased RBCs, Plts, Hb concentration, and Hct, but had no effect on MCV, MCH and MCHC in whole
27 blood collected from adult male Wistar rats ([Andjelkovic et al. 2019](#)). Pb acetate treatment increased Hct
28 and RBC distribution width, decreased MCHC, and had no effect on RBC number, Hb, MCV, and MCH
29 in adult male C57BJ mice exposed via drinking water (200 ppm; BLL = 21.6 µg/dL) for 45 days ([Corsetti](#)
30 [et al. 2017](#)). The potential effects of Pb exposure in rats were also investigated through a combination of
31 lactational and drinking water exposures. Beginning on postnatal day (PND) 1, dams were given drinking
32 water containing Pb acetate (50 mg/L). On PND 21, male pups were subsequently administered Pb
33 acetate (50 mg/L) in drinking water for an additional 40 or 65 days, at which time they were sacrificed.
34 Hct was significantly reduced in mice exposed until PND 40 (BLL = 12.67 ± 1.68 µg/dL) whereas Hct
35 and Hb were significantly decreased at PND 65 (BLL = 7.49 ± 0.78 ug/dL) ([Berrahal et al. 2011](#)). Study-
36 specific details, including animal species, strain, sex, and BLLs are highlighted in Table 7-3.

7.3.3. Integrated Summary of Red Blood Cell Survival and Function

1 Experimental animal studies evaluated in the 2013 Pb ISA ([U.S. EPA 2013](#)) demonstrate that Pb
2 exposures resulting in BLLs relevant to humans (i.e., <10 µg/dL) alter several hematological parameters,
3 increase measures of oxidative stress, and increase cytotoxicity in RBC precursor cells. While
4 epidemiologic evidence synthesized in the last review was generally coherent with results from the animal
5 studies, most of the epidemiologic studies evaluated are cross-sectional, were conducted in populations
6 with higher mean Pb exposures (i.e., BLLs >10 µg/dL), did not thoroughly consider potential
7 confounders, and lacked rigorous statistical methodology. As a result, there were considerable
8 uncertainties in the epidemiologic evidence regarding the directionality of effects; the level, timing,
9 frequency, and duration of Pb exposure that contributed to the observed associations; and whether the
10 observed associations are independent of potential confounders.

11 Though limited in number, recent PECOS-relevant animal toxicological studies continue to
12 support the findings from the last review. Specifically, these studies consistently report the effects of Pb
13 on hematological parameters, including mostly consistent evidence of a Pb-related decrease in Hb. Recent
14 epidemiologic studies expand on the evidence presented in the 2013 Pb ISA and provide additional
15 support for the experimental evidence. Although the recent studies are also cross-sectional, they include
16 populations with much lower BLL means (<10 µg/dL) and include more robust adjustment for potential
17 confounding, addressing important uncertainties from the last review. The most consistent epidemiologic
18 evidence indicates an association between BLLs and decreased Hb levels in children, which is coherent
19 with the evidence from recent experimental animal studies. While the clinical relevance of small mean
20 decrements in Hb is unclear, a few of the recent epidemiologic studies include analyses linking increased
21 BLLs to increased prevalence of anemia. Recent epidemiologic studies of Hb levels in adults were more
22 limited in number and less consistent than those in children. Additionally, of the relatively few studies
23 examining RBC counts, the results were also inconsistent.

7.4 Heme Synthesis

24 Toxicological and ecotoxicological studies evaluated in the 2006 Pb AQCD ([U.S. EPA 2006b](#))
25 and the 2013 Pb ISA ([U.S. EPA 2013](#)) provided strong evidence that exposure to Pb affects heme
26 synthesis. A limited number of epidemiologic studies contributed compelling supporting evidence. Given
27 the extensive evidence base at higher BLLs, the scope for this appendix focuses on toxicological studies
28 conducted at lower exposure levels and epidemiologic studies in nonoccupational populations (as
29 described in Section 7.2). Below, recent evidence is reviewed in the context of evidence from past
30 assessments.

7.4.1. Epidemiologic Studies of Heme Synthesis

1 The epidemiologic studies evaluated in the 2013 Pb ISA ([U.S. EPA 2013](#)) provided evidence that
2 BLLs in children and adults are associated with decreased activity of enzymes involved in the heme
3 synthesis pathway, including ALAD and ferrochelatase. Similar to studies of RBC function and survival,
4 most studies on heme synthesis are cross-sectional in design and conducted either in occupationally
5 exposed populations and/or in populations with higher mean Pb exposures (i.e., BLL >20 µg/dL). These
6 studies were additionally limited by their lack of consideration of potential confounders, although some
7 studies adjusted for or consider potential confounding factors (i.e., age, sex, urban/rural residence, height,
8 weight, BMI ([Wang et al. 2010a](#); [Ahamed et al. 2007](#); [Ahamed et al. 2006](#))). Studies that did account for
9 potential confounders reported consistent associations between increased BLLs and decreased ALAD
10 activity (Section 4.7.3.1 of the 2013 Pb ISA). Evidence for altered heme synthesis is also provided by the
11 epidemiologic studies discussed in Section 7.3.1 that report decreased Hb concentrations in association
12 with increased Pb exposure or BLLs. Decreased RBC survival and hematopoiesis can be expected to
13 occur simultaneously, and any effect on Hb levels is likely a combination of the two processes. Outside of
14 the recent studies on Hb concentrations discussed in Section 7.3.1, there are no recent PECOS-relevant
15 epidemiologic studies examining the relationship between Pb exposure and heme synthesis.

7.4.2. Toxicological Studies of Heme Synthesis

16 Pb-induced alterations in heme synthesis occurring at BLLs relevant to this ISA (e.g., 10 µg/dL)
17 have been demonstrated convincingly by a small but consistent body of evidence. In brief, Pb exposure in
18 rats inhibits several enzymes involved in heme synthesis, most notably ALAD, the enzyme that catalyzes
19 the second, rate-limiting step in heme biosynthesis ([Rendón-Ramirez et al. 2007](#); [Terayama et al. 1986](#)).
20 Pb exposure has also been shown to inhibit ferrochelatase, a mitochondrial iron (Fe)-sulfur (S) containing
21 enzyme that incorporates Fe²⁺ into protoporphyrin IX to create heme ([Rendón-Ramirez et al. 2007](#)).

22 Toxicological studies have found that Pb exposures result in increases in markers of oxidative
23 stress. For example, [Lee et al. \(2005\)](#) reported increased RBC malondialdehyde (MDA), SOD and CAT
24 levels accompanied by significant decreases in GSH and GPx in rats exposed to Pb (25 mg/kg) once a
25 week for 4 weeks. In a second drinking water study performed in rats, administration of Pb acetate
26 (750 mg/kg in drinking water for 11 weeks) resulted in decreased concentrations of plasma Vitamin C,
27 Vitamin E, nonprotein thiol, and RBC-GSH, with simultaneous increased activity of SOD and GPx
28 ([Kharoubi et al. 2008](#)). Effects on measures of oxidative stress were also observed in in vitro studies
29 including increased MDA and decreased SOD and CAT in RBCs ([Ciubar et al. 2007](#)), and decreased
30 glutathione reductase (GR) activity in human RBCs ([Coban et al. 2007](#)), and decreased GSH and
31 increased glutathione disulfide (GSSG).

1 There were no recent toxicology studies investigating the effects of Pb exposure on heme
2 synthesis that satisfied the PECOS criteria described in Section 7.2 available for this review.

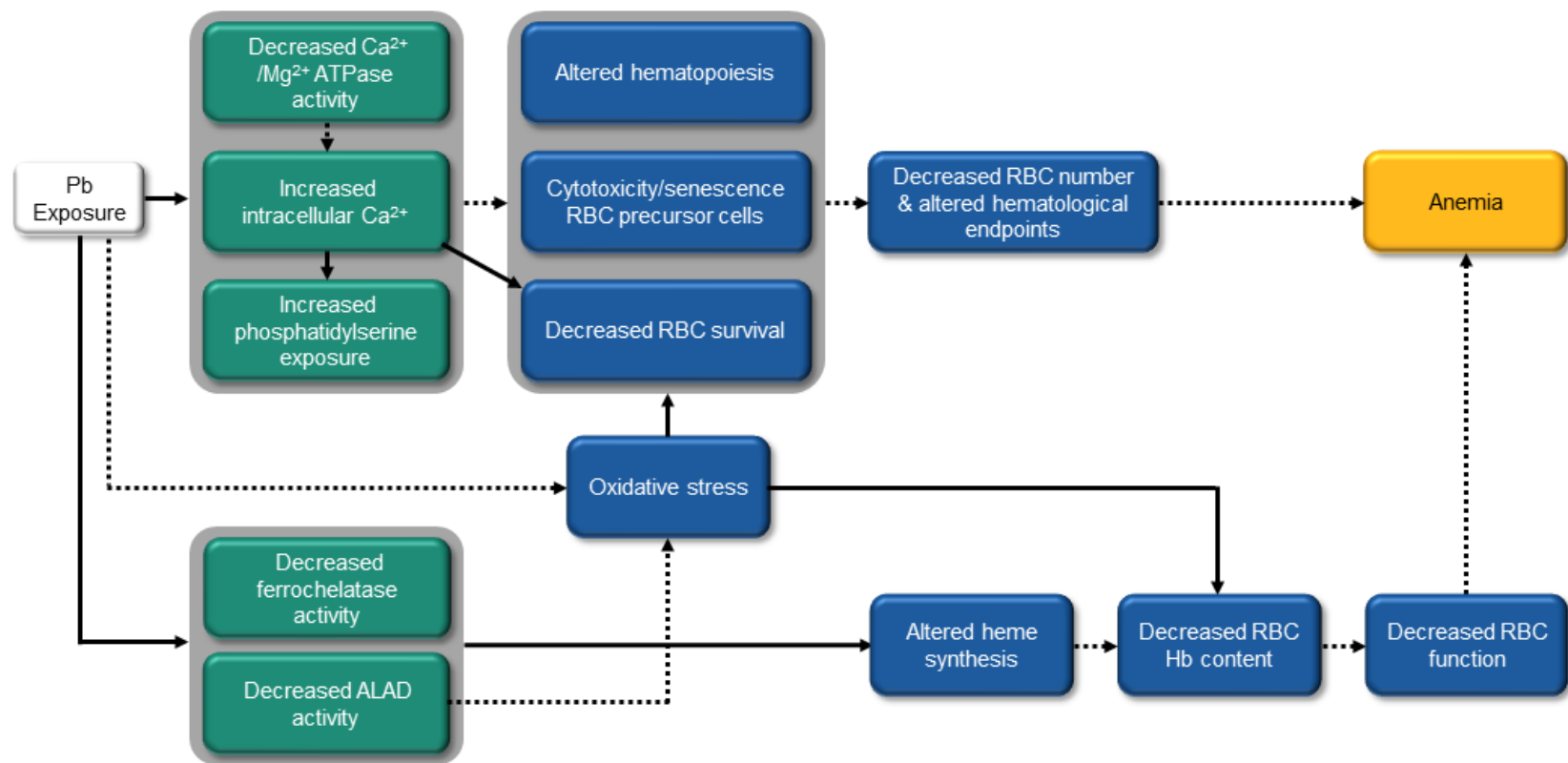
7.4.3. Integrated Summary of Heme Synthesis

3 A small number of animal toxicological studies evaluated in the 2013 Pb ISA provide consistent
4 evidence that Pb exposures affect heme synthesis, including Pb-induced decreases in ALAD ([Rendón-
5 Ramirez et al. 2007](#); [Terayama et al. 1986](#)) and ferrochelatase activities ([Rendón-Ramirez et al. 2007](#)).
6 The toxicological evidence was supported by a larger body of ecotoxicological studies that demonstrate
7 ALAD inhibition in Pb-exposed aquatic and terrestrial invertebrates and vertebrates (Sections 6.3.4.3,
8 6.4.5.2, 6.4.5.3, and 6.4.15.2 of the 2013 Pb ISA). Ecological evidence from previous reviews
9 consistently observed Pb-induced ALAD inhibition in multiple species, including birds and fish ([U.S.
10 EPA 2013, 2006b](#)). Some cross-sectional epidemiologic studies evaluated in previous reviews provide
11 supporting evidence that concurrent BLLs are associated with decreased ALAD and ferrochelatase
12 activities in both adults and children. The majority of these studies, however, are limited by the lack of
13 rigorous methodology and consideration of potential confounding.

14 Recent evidence provided by epidemiologic and toxicological studies of Pb exposure and Hb
15 levels provides additional support for Pb-related impairment of heme synthesis. These studies are
16 discussed in more detail in Section 7.3.

7.5 Biological Plausibility

17 This section describes biological pathways that potentially underlie effects on hematology
18 measures resulting from exposure to Pb. Figure 7-1 depicts the proposed pathways as a continuum of
19 upstream events connected by arrows that may lead to downstream events observed in epidemiologic
20 studies. Evidence supporting these proposed pathways was derived from Sections 7.3 and 7.4 of this ISA,
21 evidence reviewed in the 2013 Pb ISA ([U.S. EPA 2013](#)), and recent evidence collected from studies that
22 may not meet the current PECOS criteria but contain mechanistic information supporting these pathways.
23 Discussion of how exposure to Pb may lead to hematological effects contributes to an understanding of
24 the biological plausibility of epidemiologic results. Note that the structure of the biological plausibility
25 section and the role of biological plausibility in contributing to the weight-of-evidence analysis used in
26 the 2013 Pb ISA are discussed below.



ALAD = δ -aminolevulinatase; Ca^{2+} = calcium ion; Mg^{2+} = Magnesium ion; RBC = red blood cell.

Note: The boxes represent the effects for which there is experimental or epidemiologic evidence related to Pb exposure, and the arrows indicate a proposed relationship between those effects. Solid arrows denote evidence of essentiality as provided, for example, by an inhibitor of the pathway used in an experimental study involving Pb exposure. Dotted arrows denote a possible relationship between effects. Shading around multiple boxes is used to denote a grouping of these effects. Arrows may connect individual boxes, groupings of boxes, and individual boxes within groupings of boxes. Progression of effects is generally depicted from left to right and color coded (white, exposure; green, initial effect; blue, intermediate effect; orange, effect at the population level or a key clinical effect). Here, population-level effects generally reflect results of epidemiologic studies. When there are gaps in the evidence, there are complementary gaps in the figure and the accompanying text below. The structure of the biological plausibility sections and the role of biological plausibility in contributing to the weight-of-evidence analysis used in the 2022 Pb ISA are discussed in Section 7.6.

Figure 7-1 Potential biological plausibility pathways for hematological effects associated with exposure to Pb

Careful review of the available evidence indicates that exposure to Pb has the potential to modulate the hematological parameters leading to decreased RBC survival and function and altered heme synthesis. These deficits converge, promoting the development of anemia, a condition that occurs when the number of RBCs and/or the concentration of Hb in RBCs is abnormally low. Below, evidence from peer-reviewed toxicology studies providing biological plausibility for Pb-associated effects on hematological parameters is reviewed.

7.5.1. Decreased Red Blood Cell Survival and Function

As described below, there is strong evidence that Pb impacts a series of hematological parameters along a cascade of events that results in decreased RBC function and survival, possibly leading to anemia (Figure 7-1). As reviewed in the 2013 Pb ISA, Pb uptake into human RBCs occurs through a passive anion transport mechanism, and once Pb is in the cell, little leaves ([Bergdahl et al. 1997](#); [Simons 1993](#); [Simons 1986](#)). While the precise mechanisms responsible for decreasing RBC lifespan and mobility are unknown, occupational Pb exposure has been shown to decrease intracellular free Ca^{2+} levels and decrease $\text{Ca}^{2+}/\text{Mg}^{2+}$ ATPase activity in RBCs in workers ([Abam et al. 2008](#); [Quintanar-Escorza et al. 2007](#)). These changes are associated with fragility and morphological alterations in RBCs in Pb-exposed workers. Pb-induced increases in intracellular Ca^{2+} levels also play a role in the activity of phospholipid scramblases and flippases in RBCs, increasing access to PS by tissue macrophages and triggering splenic sequestration and destruction of RBCs, leading to reduced numbers of RBCs in circulation ([Jang et al. 2011](#)). Importantly, [Ahyayauch et al. \(2018\)](#) showed that inhibiting Ca^{2+} increase stimulated by Pb results in decreased flippase activity and prevented destruction of RBCs. This pathway is depicted in Figure 7-1 by solid lines linking increased intracellular Ca^{2+} to increased PS exposure and decreased RBC survival. In addition, phagocytosis of Pb-exposed RBC by human renal proximal tubular cells was mediated by PS ([Kwon and Chung 2016](#)). Heme-regulated eIF2 α kinase was shown to protect RBC from Pb-induced hemolytic stress in mice ([Wang et al. 2015](#)). Pb-induced hemolysis was also documented in other recent studies ([Hossain et al. 2015](#); [Mrugesh et al. 2011](#)). Furthermore, Pb exposure reduced the number of RBCs and Hb levels in rats ([Ibrahim et al. 2012](#)). Consistent with the pattern seen in epidemiology studies, the effects of Pb exposure on RBC number and Hb level were more pronounced in 3-month-old Wistar rats than in adult animals ([Daku et al. 2019](#)). In addition, [Cai et al. \(2018\)](#) reported that Pb administration reduced RBC lifespan in mice. These findings support the conclusion that Pb alters RBC survival and function, consistent with the larger body of evidence showing measures of decreased hematological parameters (i.e., RBC number, Hb, Hct, MCV, and/or MCH) in children ([Guo et al. 2021](#); [Kuang et al. 2020](#); [Li et al. 2018](#); [Liu et al. 2015](#); [Liu et al. 2012](#)) and animals ([Odo et al. 2020](#); [Andjelkovic et al. 2019](#); [Cai et al. 2018](#); [Corsetti et al. 2017](#); [Lakshmi et al. 2013](#); [Berrahal et al. 2011](#); [Sharma et al. 2010](#); [Wang et al. 2010b](#); [Baranowska-Bosiacka et al. 2009](#); [Massó-González and Antonio-García 2009](#); [Simsek et al. 2009](#); [Massó et al. 2007](#)).

Pb exposure also has the potential to disrupt normal hematopoiesis. Erythropoietin (EPO) is a glycoprotein hormone excreted by the kidney to promote the development of RBCs in bone marrow. As reviewed in the 2006 Pb AQCD, Pb exposure has been observed to alter EPO production in children ([Graziano et al. 2004](#); [Factor-Litvak et al. 1999](#); [Factor-Litvak et al. 1998](#)). Available data support the postulation that observed increases in EPO in younger children reflect bone marrow hyperactivity to counteract RBC destruction, whereas the lack of EPO elevation in older children may reflect a transitional period in which increasing renal and bone marrow toxicity leads to observed decreases in EPO later in life ([U.S. EPA 2006a](#)). In addition to altering levels of a key hormone involved in hematopoiesis, Pb exposure has the potential to alter hematopoiesis by causing cytotoxicity ([Alghazal et al. 2008](#); [Marques et al. 2006](#); [Çelik et al. 2005](#)) and senescence ([Cai et al. 2018](#); [Nagano et al. 2015](#)) of RBC precursors. [Baktybaeva \(2011\)](#) reported that intraperitoneal injection of Pb acetate resulted in reduced bone marrow hematopoiesis in mice. Furthermore, Pb is known to reduce erythropoiesis, causing anemia in children ([Dai et al. 2017](#); [Ahamed et al. 2011](#)). Altered EPO levels and RBC precursor cytotoxicity have the potential to alter the number of RBCs in circulation which may lead to anemia.

7.5.2. Altered Heme Synthesis

Although the mechanisms that could lead to Pb-induced anemia are not fully understood, as reviewed in EPA's 2006 AQCD ([U.S. EPA 2006b](#)), Pb is known to act directly on two enzymes involved in heme synthesis: ALAD and ferrochelatase. ALAD, a cytoplasmic enzyme requiring zinc (Zn) for enzymatic activity, catalyzes the rate-limiting step in heme biosynthesis. Inhibition of ALAD activity has been reported in adults ([Wang et al. 2010a](#)) and children ([Dai et al. 2017](#); [Wang et al. 2010a](#); [Ahamed et al. 2007](#)) as well as in animal toxicology studies reporting BLLs relevant to this ISA (i.e., 24.7 µg/dL) ([Rendón-Ramírez et al. 2007](#); [Terayama et al. 1986](#)). ALAD activity was also reduced in animal studies reporting BLLs higher than those meeting the PECOS criteria in this ISA ([Mani et al. 2020](#); [Velaga et al. 2014](#); [Whittaker et al. 2011](#); [Gautam and Flora 2010](#); [Lee et al. 2005](#)). Pb exposure has also been shown to inhibit ferrochelatase, a mitochondrial iron (Fe)-sulfur (S)containing enzyme that incorporates Fe²⁺ into protoporphyrin IX to create heme ([Rendón-Ramírez et al. 2007](#)). Pb inhibits the insertion of Fe²⁺ into the protoporphyrin ring and instead, Zn is inserted into the ring creating Zn-protoporphyrin (ZPP). Evidence for altered heme synthesis is supported by a large body of evidence collected from occupationally exposed adults ([Ukajiofo et al. 2009](#); [Khan et al. 2008](#); [Patil et al. 2006](#); [Karita et al. 2005](#)), children ([Queirolo et al. 2010](#); [Shah et al. 2010](#); [Olivero-Verbel et al. 2007](#); [Riddell et al. 2007](#)), and Pb-exposed experimental animal models ([Andjelkovic et al. 2019](#); [Cai et al. 2018](#); [Berrahal et al. 2011](#); [Baranowska-Bosiacka et al. 2009](#); [Massó-González and Antonio-García 2009](#); [Simsek et al. 2009](#); [Rendón-Ramírez et al. 2007](#); [Marques et al. 2006](#); [Terayama et al. 1986](#)) reporting decreased Hb concentrations in association with Pb exposure or increased BLLs. Further demonstrating the role Pb plays in the activity of these important enzymes, chelation therapy restored ALAD activity and reduced ZPP levels in blood harvested from rats exposed to Pb via drinking water for 90 days ([Ata et al. 2018](#)).

Oxidative stress is caused by an imbalance between production and elimination of reactive oxygen species (ROS) in cells or tissues that exceed the capacity of antioxidant defense mechanisms. ROS are unstable, highly reactive molecules formed from molecular oxygen and include, for example, superoxide (O_2^-), hydroxyl radical (OH), and hydrogen peroxide (H_2O_2). Although ROS play an important role in healthy biological systems (e.g., cell signaling, cellular differentiation, immune responses), unregulated ROS can cause direct and indirect damage to nucleic acids, proteins, and lipids leading to cytotoxicity, tissue injury, and even disruption of normal physiology ([Auten and Davis 2009](#)). Oxidative stress is involved in both arms of the pathway leading to anemia shown in Figure 7-1, including effects on RBC MDA, SOD, CAT, GSH, and GPx levels ([Kharoubi et al. 2008](#); [Lee et al. 2005](#)). Effects of Pb exposure on measures of oxidative stress—including MDA and decreased SOD, CAT, GR, and GSSG—were also observed in vitro ([Ciubar et al. 2007](#); [Coban et al. 2007](#)).

Supporting the role of oxidative stress in the development of anemia, administration of antioxidants reduced the effects of Pb exposure on levels of GSH ([Alcaraz-Contreras et al. 2011](#)), MDA ([Alcaraz-Contreras et al. 2011](#)) and Hb ([Farooq et al. 2016](#); [Sajitha et al. 2016](#); [Sarkar et al. 2015](#); [Eshginia and Marjani 2013](#)) and reduced the effects of Pb exposure on SOD activity ([Eshginia and Marjani 2013](#)), ROS production ([Nagano et al. 2015](#); [Sarkar et al. 2015](#)), hematopoietic stem cell (HSC) number ([Nagano et al. 2015](#)), HSC colony formation ([Cai et al. 2018](#)), HSC senescence markers ([Cai et al. 2018](#); [Nagano et al. 2015](#)), RBC number ([Farooq et al. 2016](#); [Sajitha et al. 2016](#); [Sarkar et al. 2015](#)), and PS exposure ([Sarkar et al. 2015](#)) in animal studies. Conflicting evidence on the effects of antioxidant treatment on ALAD activity was reported in the literature ([Sajitha et al. 2016](#); [Alcaraz-Contreras et al. 2011](#)), whereas the evidence for effects of Pb exposure on Hb levels was consistent; thus, a solid arrow connects oxidative stress to decreased RBC Hb content in Figure 7-1. Further demonstrating the role of oxidative stress in the development of anemia, decreased ALAD activity results in the accumulation of δ -aminolevulinic acid (δ -ALA) in blood and urine, where it undergoes tautomerization and autoxidation. Oxidized δ -ALA leads to the generation of ROS (i.e., O_2^- , OH, H_2O_2 , and an aminolevulinic acid [ALA]) radicals ([Hermes-Lima et al. 1991](#); [Monteiro et al. 1991](#); [Monteiro et al. 1989](#); [Monteiro et al. 1986](#)). Reflecting the strength of the evidence described above, the pathway connecting oxidative stress to altered hematopoiesis, cytotoxicity/senescence RBC precursor cells, and decreased RBC survival is depicted as a solid line.

Decreased RBC Hb content and oxidative stress associated with Pb exposure have been demonstrated to alter RBC function. This conclusion is supported by direct evidence for binding of Pb to key enzymes in the heme synthesis pathway, Pb-induced oxidative stress resulting in decreased RBC Hb content and effects on hematopoiesis, RBC precursor cells, and RBC survival. Decreased RBC number coupled with impaired RBC function, if of sufficient magnitude, leads to Pb-induced anemia.

7.6 Summary and Causality Determination

7.6.1. Causality Determination for Red Blood Cell Survival and Function

1 The 2013 Pb ISA presented causality determinations for two groups of hematological endpoints:
2 heme synthesis and RBC survival and function. Although there are enzymes and hematological
3 parameters that are distinct indicators of these processes, the potential biological plausibility pathways in
4 which exposure to Pb may result in hematological effects demonstrate a spectrum of events, which can be
5 challenging to attribute to a unique line of evidence (Figure 7-1). For example, altered heme synthesis can
6 decrease Hb levels, which in turn has been shown to alter RBC function. Because of this
7 interconnectedness, this assessment presents a single causality determination for Pb exposure and heme
8 synthesis and RBC survival and function. This approach allows for a more holistic evaluation of inter-
9 related health endpoints, including a discussion of how all individual lines of evidence contribute to the
10 overall causality determination. The key evidence, as it relates to the causal framework, is outlined below,
11 and summarized in Table 7-1.

7.6.2. Evidence for Red Blood Cell Survival and Function

12 The 2013 Pb ISA concluded that there is a “causal relationship” between Pb exposure and
13 decreased RBC survival and function ([U.S. EPA 2013](#)). This causality determination was made on the
14 basis of a strong body of evidence from experimental animal studies demonstrating that Pb exposures
15 alter several hematological parameters (e.g., Hb, Hct, MCV, MCH), induce oxidative stress (e.g., alter
16 antioxidant enzyme activities [SOD, CAT, GPx], decrease cellular GSH, and increase lipid peroxidation),
17 and increase cytotoxicity in RBC precursor cells in rodents exposed to various forms of Pb via drinking
18 water and gavage resulting in BLLs ≤ 30 $\mu\text{g/dL}$ ([Molina et al. 2011](#); [Baranowska-Bosiacka et al. 2009](#);
19 [Lee et al. 2005](#)). Consistent results were observed in several additional studies in rodents that did not
20 report BLLs. Epidemiologic evidence was coherent with results from the evaluated toxicological studies
21 but was subject to more uncertainties. Notably, the epidemiologic evidence consisted of cross-sectional
22 studies that were conducted in populations with higher mean Pb exposures (i.e., BLLs >10 $\mu\text{g/dL}$), did not
23 thoroughly consider potential confounders, and lacked rigorous statistical methodology. These limitations
24 precluded strong conclusions on the directionality of effects; the level, timing, frequency, and duration of
25 Pb exposure that contributed to the observed associations; and whether the observed associations are
26 independent of potential confounders. Although there were substantial uncertainties in the epidemiologic
27 evidence, animal toxicological evidence established a clear basis for temporality of exposure to Pb and
28 effects on RBCs. Additional support for these findings was provided by toxicological and epidemiologic
29 studies demonstrating increased intracellular calcium concentrations, decreased $\text{Ca}^{2+}/\text{Mg}^{2+}$ ATPase
30 activity, and increased PS exposure, establishing biological plausibility for Pb-induced changes in RBC
31 survival.

1 Although limited in number, recent PECOS-relevant animal toxicological studies continue to
2 support the findings from the last review. The most consistent evidence comes from studies that report
3 decreased Hb levels in rodents following Pb exposures, resulting in BLLs ranging from 7.5 to 14.7 µg/dL
4 ([Andjelkovic et al. 2019](#); [Cai et al. 2018](#); [Berrahal et al. 2011](#)). Other recent toxicological studies noted
5 Pb-induced decrements in Hct ([Andjelkovic et al. 2019](#)), packed cell volume (PCV) ([Berrahal et al.](#)
6 [2011](#)), and hematopoiesis ([Andjelkovic et al. 2019](#)). Recent epidemiologic studies expand on the evidence
7 presented in the 2013 Pb ISA and are coherent with the experimental evidence. Although the recent
8 studies are also cross-sectional, they include populations with much lower BLL means (<10 µg/dL) and
9 include more robust adjustment for potential confounding, addressing important uncertainties from the
10 last review. The most consistent epidemiologic evidence indicates an association between increased BLL
11 and decreased Hb levels in children (Section 7.3.1), which is in line with the evidence from recent
12 experimental animal studies. While the clinical relevance of small mean decrements in Hb across
13 exposure quintiles is unclear, a few of the recent epidemiologic studies observed increases in the odds of
14 prevalent anemia in children associated with increasing quantiles of BLLs ([Guo et al. 2021](#); [Li et al.](#)
15 [2018](#)). Recent epidemiologic studies of Hb in adults were more limited in number and less consistent than
16 those in children. Additionally, the relatively few studies examining RBC counts were also inconsistent.

7.6.3. Evidence for Heme Synthesis

17 The 2013 Pb ISA concluded there is a “causal relationship” between Pb exposure and altered
18 heme synthesis ([U.S. EPA 2013](#)). This determination was based on a small but consistent body of studies
19 in adult animals reporting that Pb exposures via drinking water and gavage (resulting in BLLs relevant to
20 this ISA) for 15 days to 9 months decreased ALAD ([Rendón-Ramírez et al. 2007](#); [Terayama et al. 1986](#))
21 and ferrochelatase activities ([Rendón-Ramírez et al. 2007](#)). Notably, [Rendón-Ramírez et al. \(2007\)](#)
22 observed effects on ALAD and ferrochelatase activities in albino Wistar rats at mean BLLs of 24.7 µg/dL
23 after Pb administration drinking water for 15 or 30 days. Supporting this toxicological evidence was a
24 larger body of ecotoxicological studies that demonstrate altered heme synthesis in Pb-exposed aquatic and
25 terrestrial invertebrates and vertebrates. Ecological evidence from previous reviews consistently observed
26 Pb-induced ALAD inhibition in multiple species, including birds and fish ([U.S. EPA 2013](#), [2006b](#)).
27 Cross-sectional epidemiologic studies provided supporting evidence that concurrent elevated BLLs are
28 associated with decreased ALAD and ferrochelatase activities in both adults and children. However, the
29 majority of these studies are limited by the lack of rigorous methodology and consideration of potential
30 confounding. Although there were limitations in the epidemiologic evidence, some studies in children did
31 control for or consider potential confounding, and effects in adults and children in these studies are
32 coherent with effects observed in animal toxicological studies.

33 The relationship between Pb exposure and altered heme synthesis was further supported by cross-
34 sectional epidemiologic studies indicating that increased BLLs were associated with decreased Hb in
35 children and occupationally exposed adults. These findings were consistent with several toxicological

1 studies that observed decreased Hb levels in laboratory animals exposed to Pb. Decreased Hb levels can
2 be a direct indicator of decreased heme synthesis.

3 Recent PECOS-relevant studies are limited in number and focus mainly on Hb levels but continue
4 to provide support for Pb-related alterations in heme synthesis. Notably, recent epidemiologic studies
5 indicating an association between increased BLLs and decreased Hb include more robust statistical
6 methods, expanded consideration of potential confounders, and populations with much lower BLLs than
7 the studies included in the previous reviews (mean or median BLLs ranging from 3.04 to 8.38 µg/dL;
8 Section 7.3.1). The recent epidemiologic evidence is coherent with recent toxicological studies, which
9 observed Hb decrements in Pb-exposed mice ([Andjelkovic et al. 2019](#); [Cai et al. 2018](#)). While the cross-
10 sectional nature of the epidemiologic studies introduces uncertainty about the temporality of the exposure
11 and outcome, animal toxicological evidence establishes clear temporality of exposure to Pb and altered
12 heme synthesis.

7.6.4. Causality Determination

13 In summary, there is coherent evidence across toxicological and epidemiologic studies that Pb
14 exposures alter several hematological parameters, decrease enzyme activity related to heme synthesis, and
15 increase RBC oxidative stress. Although all evaluated epidemiologic studies are cross-sectional,
16 toxicological studies establish temporality between exposure to Pb and effects on heme synthesis and
17 RBCs. Additionally, recent epidemiologic and animal studies continue to demonstrate evidence of a
18 causal relationship at relevant BLLs, and recent epidemiologic studies address uncertainties from
19 previous reviews by expanding adjustment for potential confounders and using more robust statistical
20 methods (i.e., multivariable regression models). Because of the contribution of bone Pb levels to
21 concurrent BLLs, associations with concurrent BLLs may reflect an effect of past and/or recent Pb
22 exposures. Therefore, there is uncertainty regarding the timing, duration, and level of Pb exposure
23 associated with observed hematological effects in children and adults. Biological plausibility for the
24 observed associations is provided by toxicological and epidemiologic studies demonstrating increased
25 intracellular calcium concentrations, decreased $\text{Ca}^{2+}/\text{Mg}^{2+}$ ATPase activity, and increased PS exposure,
26 which collectively can lead to fragility, morphological alterations in RBCs, and RBC destruction. **Taken**
27 **together, there is sufficient evidence to conclude that there is a causal relationship between Pb**
28 **exposure and hematological effects, including altered heme synthesis and decreased RBC survival**
29 **and function.**

Table 7-1 Summary of evidence indicating a causal relationship between Pb exposure and hematological effects

Rationale for Causality Determination ^a	Key Evidence ^b	Key References ^b	Pb Biomarker Levels Associated with Effects ^c
Red Blood Cell Survival and Function			
Consistent evidence from toxicological studies with relevant exposures	Large body of studies with generally consistent findings for decreased RBC survival and function in rodents:	See Section 7.3.2	
			Mean BLLs (± SD):
	Decreased plasma Hb concentration	Cai et al. (2018)	9.3 ± 0.98 µg/dL
		Andjelkovic et al. (2019)	29.0 ± 43.1 µg/dL
		Berrahal et al. (2011)	12.67 ± 1.68 µg/dL PND 40; 7.49 ± 0.78 µg/dL PND 65
		Sharma et al. (2010)	1.72 ± 0.02 µg/dL
		Massó-González and Antonio-García (2009)	22.8 ± 0.50 µg/dL
		Baranowska-Bosiacka et al. (2009)	7.11 ± 1.7 µg/dL
	Decreased Hct	Andjelkovic et al. (2019)	29.0 ± 14.7 µg/dL
		Massó et al. (2007)	22.8 ± 0.50 µg/dL
		Massó-González and Antonio-García (2009)	22.8 ± 0.50 µg/dL
		Berrahal et al. (2011)	12.67 ± 1.68 µg/dL PND 40; 7.49 ± 0.78 µg/dL PND 65
	Decreased RBCs	Andjelkovic et al. (2019)	29.0 ± 14.7 µg/dL

Rationale for Causality Determination ^a	Key Evidence ^b	Key References ^b	Pb Biomarker Levels Associated with Effects ^c
		Cai et al. (2018)	9.3 ± 0.98 µg/dL
		Sharma et al. (2010)	1.72 ± 0.02 µg/dL
	Decreased PCV	U.S. EPA (2013)	
	Increased eryptosis	U.S. EPA (2013)	
		U.S. EPA (2006b)	
	Decreased hematopoiesis	U.S. EPA (2013)	
		U.S. EPA (2006b)	
	Increased oxidative stress	In vitro:	
		Ciubar et al. (2007)	
		Coban et al. (2007)	
		Shin et al. (2007)	
Consistent evidence from multiple epidemiologic studies of children with relevant BLLs provides coherence with toxicological evidence.	Cross-sectional studies provide support for experimental evidence with consistent associations between blood Pb and decreased in Hb in children. Recent studies adjusted for a number of relevant potential confounders, including age, sex, BMI, SES factors, and nutrition.	Guo et al. (2021) Kuang et al. (2020) Li et al. (2018) Liu et al. (2015) Liu et al. (2012)	Mean BLLs: 3.07–3.21 µg/dL 3.04 µg/dL 8.38 µg/dL (Median) 7.33 µg/dL 4.30 µg/dL (Median)

Rationale for Causality Determination ^a	Key Evidence ^b	Key References ^b	Pb Biomarker Levels Associated with Effects ^c
	Cross-sectional studies provide generally consistent evidence for associations between blood Pb and altered hematological parameters (e.g., RBC counts, Hct, MCV, and MCH), measures of oxidative stress (e.g., SOD, CAT, GPx, GSH, and lipid peroxidation), and hematopoiesis (e.g., decreased erythropoietin). Evidence base limited by lack of adjustment for potential confounders and populations with higher BLLs.	(U.S. EPA 2013)	>10 µg/dL
Biological Plausibility	Evidence of increased [Ca ²⁺] _i and decreased Ca ²⁺ /Mg ²⁺ ATPase activity in the RBCs of exposed workers. [Ca ²⁺] _i levels highly correlated with blood Pb even among unexposed controls.	See Section 7.5.2	
Altered RBC membrane ion transport			
PS exposure	[Ca ²⁺] _i levels increased in RBCs from healthy volunteers when exposed in vitro to Pb. [Ca ²⁺] _i associated with increased RBC fragility and alterations in RBC morphology. Consistent evidence from in vivo and in vitro studies that Pb exposure increases PS exposure on RBC membranes via modulation of [Ca ²⁺] _i concentrations. Increased PS exposure leads to eryptosis and phagocytosis by macrophages.		
Heme Synthesis			
Consistent evidence in animals with relevant exposures	A small, but consistent toxicology evidence base indicates decreased heme synthesis in rodents with relevant Pb concentrations and routes of exposure.	Rendón-Ramírez et al. (2007) Terayama et al. (1986)	BLL: 24.7 ± 2.4 µg/dL Exposures: 500–5,000 ppm in drinking water, 15–30 days as adults
		Ahamed et al. (2006)	Mean BLL: 7.40 and 13.27 µg/dL

Rationale for Causality Determination ^a	Key Evidence ^b	Key References ^b	Pb Biomarker Levels Associated with Effects ^c
Coherence in a limited number of epidemiologic studies with relevant BLLs	Cross-sectional studies that considered potential confounding by age, sex, urban/rural residence, height, weight, BMI found consistent associations with lower ALAD and ferrochelatase activities in children.	Ahamed et al. (2007)	BLL: >10 µg/dL compared with <10 µg/dL
	Concurrent BLL associated with lower ALAD and higher ZPP in adults with consideration for potential confounding by age, sex, smoking status, and alcohol use.	Wang et al. (2010a)	Mean BLL: 6.71 µg/dL
Support from toxicological and epidemiologic evidence for decreases in Hb, a direct marker of decreased heme synthesis	Consistent evidence in animals with relevant Pb exposures for decreases in Hb levels.	Baranowska-Bosiacka et al. (2009) . Sharma et al. (2010) Section 7.4.2	Adult animals: BLL 1.7 µg/dL after 40-day Pb exposure
	Consistent associations between concurrent BLLs and decreased Hb in children. Associations observed at low BLLs with thorough consideration of potential confounders.	Guo et al. (2021) Kuang et al. (2020) Li et al. (2018) Liu et al. (2015) Liu et al. (2012)	Mean BLLs: 3.07–3.21 µg/dL 3.04 µg/dL 8.38 µg/dL (Median) 7.33 µg/dL 4.30 µg/dL (Median)
Biological Plausibility	Altered Ion Status: Evidence that Pb competitively inhibits the binding of Zn ions necessary for ALAD activity. Pb also inhibits the incorporation of Fe ²⁺ into protoporphyrin IX by ferrochelatase, resulting in Zn-protoporphyrin production.	See Section 7.5.2	

Rationale for Causality Determination ^a	Key Evidence ^b	Key References ^b	Pb Biomarker Levels Associated with Effects ^c
--	---------------------------	-----------------------------	--

ALAD = δ -aminolevulinate dehydratase; BLL = blood lead level; BMI = body mass index; Ca^{2+} = calcium; CAT = catalase; Fe^{2+} = iron; GPx = glutathione peroxidase; GSH = glutathione; Hb = hemoglobin; Hct = hematocrit; i = inorganic; MCV = mean corpuscular volume; Mg^{2+} = magnesium; Pb = lead; PCV = packed cell volume; PND = postnatal day; PS = phosphatidylserine; RBC = red blood cell; SES = socioeconomic status; SOD = superoxide dismutase; Zn = zinc; ZPP = Zn-protoporphyrin.

^aBased on aspects considered in judgments of causality and weight of evidence in causal framework in Table I and Table II of the Preamble to the ISAs ([U.S. EPA 2015](#)).

^bDescribes the key evidence and references, supporting or contradicting, contributing most heavily to causality determination and, where applicable, to uncertainties or inconsistencies. References to earlier sections indicate where the full body of evidence is described.

^cDescribes the Pb biomarker levels at which the evidence is substantiated.

7.7 Evidence Inventories—Data Tables to Summarize Study Details

Table 7-2 Epidemiologic studies of exposure to Pb and hematological effects

Reference and Study Design	Study Population	Exposure Assessment	Outcome	Confounders	Effect Estimates and 95% CIs ^a
Children					
† Guo et al. (2021) Guangdong China 2014-2017 Cross-sectional	Guangdong Women and Children's Hospital n: 17486 Children 0–5 yr old visiting hospital for routine health examination	Blood BLLs were measured using atomic absorption spectrometry Age at Measurement: 0–5 yr Means: Males: 3.21 µg/dL; Females: 3.07 µg/dL	Hb Hb (g/L) measured using an automated hematology analyzer Age at Outcome: 0–5 yr	Age and sex	Hb Mean Difference (g/L) Q1 (<1.61) Reference Q2 (1.61–2.44) –0.05 (–0.51, 0.40) Q3 (2.44–3.33) –0.02 (–0.48, 0.43) Q4 (3.33–4.50) –0.48 (–0.94, –0.04) Q5 (>4.50) –1.25 (–1.71, –0.78) Anemia (OR) Q1 (<1.61) Reference Q2 (1.61–2.44) 1.08 (0.94, 1.23) Q3 (2.44–3.33) 1.16 (1, 1.33) Q4 (3.33–4.50) 1.25 (1.07, 1.43) Q5 (>4.50)

Reference and Study Design	Study Population	Exposure Assessment	Outcome	Confounders	Effect Estimates and 95% CIs ^a
					1.45 (1.26, 1.67)
† Kuang et al. (2020) Nanjing China 2012 Cross-sectional	n: 395 Convenience sample of children 7–11 yr old	Blood Blood Pb was measured in venous whole blood using ICP-MS Age at Measurement: 7–11 yr old Mean: 3.04 µg/dL; Median: 2.61 µg/dL	Hematological Parameters RBC, Hb, Hct, MCV, MCH, and MCHC measured by a whole cell analyzer Age at Outcome: 7–11 yr old	Picky eaters and passive smoking (age, gender, parents' education, and parents' occupation also considered)	RBC Count (10¹²/L) <i>Boys:</i> 0.02 (-0.01, 0.04) <i>Girls:</i> 0.01 (-0.02, 0.03) Hb (g/L) <i>Boys:</i> -0.12 (-0.22, -0.02) <i>Girls:</i> -0.08 (-0.23, 0.07) Hct (%) <i>Boys:</i> -0.04 (-0.08, -0.01) <i>Girls:</i> -0.02 (-0.07, 0.02) MCV (fL) -0.04 (-0.06, -0.02) MCH (pg) -0.01 (-0.02, -0.00) MCHC (g/L) 0.26 (-0.64, 1.15)
† Liu et al. (2012) Changzhou City China Cross-sectional	China Jintan Child Cohort Study n: 140 Convenience sample of preschool age children	Blood Blood Pb was measured in whole blood using GFAAS Age at Measurement: Median age: 3 yr old Median: 4.3 µg/dL Maximum: 11.4 µg/dL	Hb Hb measured in whole blood using a photoelectric colorimeter Age at Outcome: Median age: 3 yr old	Age, sex, height, weight, iron deficiency	Hb (g/dL)* <i>Full Population</i> -0.096 (-0.18, -0.012) <i>Blood Pb < 10 µg/dL</i> -0.174 (-0.27, -0.078)

Reference and Study Design	Study Population	Exposure Assessment	Outcome	Confounders	Effect Estimates and 95% CIs ^a
† Li et al. (2018) Hubei and Hunan Provinces China 2012-2017 Cross-sectional	Blood Lead Intervention Program n: 758 Children Ages 5–8 yr recruited from four counties in two provinces One county in each province had high environmental Pb levels (battery plant and mining)	Blood Blood Pb was measured in venous whole blood using GFAAS Age at Measurement: 5–8 yr Median: 8.38 µg/dL 75 th : 13.51 µg/dL 90 th : 18.77 µg/dL 95 th : 21.82 µg/dL	Hematological Parameters Hb, MCH, RBCs, and Plt measured in venous whole blood using an automated hematology analyzer Age at Outcome: 5–8 yr	Age, sex, BMI, environmental Pb exposure level, and serum iron, zinc, and calcium	ORs Decreased Hb (<115 g/L) 1.05 (1.00, 1.11) Decreased RBC (<4 × 10¹²/L for boys; <3.5 × 10¹²/L for girls) 1.11 (1.05, 1.16) Decreased Plt (<100 × 10⁹/L) 1.11 (1.05, 1.16) Decreased MCH (<27 pg) 1.11 (1.05, 1.16)
† Liu et al. (2015) Guiyu, Chendian, and Chaonan China 2006–2011 Cross-sectional	n: 855 Children 3–7 yr old from e-waste processing area or control industrial areas without high environmental Pb exposures	Blood Blood Pb was measured using GFAAS. Blood Pb was divided by Hct as a fraction of the whole blood to estimate erythrocyte Pb Age at Measurement: 3–7 yr old Median: Blood Pb: 7.33 µg/dL; Erythrocyte Pb: 19.3 µg/dL	Hb Hb, MCH, RBCs, and Plt measured in venous whole blood using an automated hematology analyzer Age at Outcome: 3–7 yr old	Age, sex, residence area, and SES	Hb (g/L) Mean Difference Q1 (5.98–13.52)* Reference Q2 (13.52–19.35)* –0.02 (–1.89, 1.52) Q3 (19.35–28.42)* –3.01 (–4.71, 1.31) Q4 (28.42–101.01)* –3.97 (–5.68, –2.27) *Erythrocyte Pb (µg/dL)
Adults					

Reference and Study Design	Study Population	Exposure Assessment	Outcome	Confounders	Effect Estimates and 95% CIs ^a
† Park and Lee (2013) South Korea 2008-2010 Cross-sectional	KNHANES n: 4522 General population, ≥20 yr old	Blood Blood Pb was measured in venous whole blood using GFAAS Age at Measurement: ≥20 yr Geometric Means: Males: 2.46 µg/dL; Females: 1.98 µg/dL	Hb Blood Hb (g/dL) measured using an automated hematology analyzer Age at Outcome: ≥20 yr	Age, BMI, education, smoking and drinking status, and rural/urban residence	Hb (g/dL)* <i>Men</i> 0.04 (0.03, 0.06) <i>Women</i> 0.04 (0.02, 0.06) *Not standardized. Per ln(Pb) increase
† Kim and Lee (2013) South Korea 2008-2010 Cross-sectional	KNHANES n: 5951 General population, ≥20 yr old	Blood Blood Pb was measured in whole blood using GFAAS. Blood Pb was divided by Hct as a fraction of the whole blood to estimate erythrocyte Pb Age at Measurement: ≥20 yr old Median: Blood Pb: 2.31 µg/dL; Erythrocyte Pb: 5.4 µg/dL 75 th : Blood Pb: 3.01 µg/dL; Erythrocyte Pb: 6.9 µg/dL	Hb Hb measured in whole blood using an automated hematology analyzer Age at Outcome: ≥20 yr old	Sex, age, obesity, residence area, education level, smoking and drinking status, serum ferritin, and serum creatinine	Hb (g/L) Mean Difference Q1 (<1.73) Reference Q2 (1.73–2.31) 0.13 (0.03, 0.23) Q3 (2.31–3.01) 0.33 (0.23, 0.42) Q4 (>3.01) 0.42 (0.30, 0.53) Q1 (<4.1)* Reference Q2 (4.1–5.4) –0.06 (0.15, 0.03) Q3 (5.4–6.9) –0.06 (–0.15, 0.03) Q4 (>6.9) –0.14 (–0.25, –0.04) *Erythrocyte Pb (µg/dL)

Reference and Study Design	Study Population	Exposure Assessment	Outcome	Confounders	Effect Estimates and 95% CIs ^a
† La-Llave-León et al. (2015) Durango Mexico 2007-2008 Cross-Sectional	n: 292 Pregnant women, 14–41 yr old	Blood Blood Pb was measured in venous whole blood using GFAAS Age at Measurement: 14–41 yr old Mean: 2.79 µg/dL	Hematological Parameter RBC, Hb, Hct, MCV, MCH, and MCHC measured using an automated hematology analyzer Age at Outcome: 14–41 yr old	BMI, gestational age, age, parity, gestations, and household monthly income per person	RBC Count (× 10⁶ µg/dL) 0.034 (0.013, 0.056)

BMI = body mass index; BW = body weight; CI = confidence interval; e-waste = electronic waste; GFAAS = graphite furnace atomic absorption spectroscopy; Hb = hemoglobin; Hct = hematocrit; ICP-MS = inductively coupled plasma mass spectrometry; KNHANES = Korean National Health and Nutrition Examination Survey; ln = natural log; MCH = mean corpuscular hemoglobin; MCHC = mean corpuscular hemoglobin concentration; MCV = mean corpuscular volume; OR = odds ratio; Pb = lead; Plt = platelet; PND = postnatal day; Q = quartile; RBC = red blood cell; RDW = red blood cell distribution width; SES = socioeconomic status; yr = years.

^a Effect estimates are standardized to a 1 µg/dL increase in blood Pb level or a 10 µg/g increase in bone Pb level, unless otherwise noted. For studies that report results corresponding to a change in log-transformed Pb biomarkers, effect estimates are assumed to be linear within the 10th to 90th percentile interval of the biomarker and standardized accordingly.

†Studies published since the 2013 Pb ISA.

Table 7-3 Animal toxicological studies of Pb exposure and hematological effects

Study	Species (Stock/Strain), n, Sex	Timing of Exposure	Exposure Details (Concentration, Duration)	BLL as Reported (µg/dL) ^a	Endpoints Examined
Berrahal et al. (2011)	Rat (Wistar) Control (vehicle), M, n = 12–16 50 mg/L Pb, M, n = 12–16	PND 1 to PND 21: Lactational PND 21 to PND 40 or PND 65: Drinking water	Dams were given 50 mg/L Pb acetate in drinking water until weaning on PND 21. Male offspring received 50 mg/L Pb acetate in drinking water from PND 21 to PND 40 or PND 65. Control animals received tap water.	PND 40: 1.76 ± 0.33 µg/dL for 0 µg/dL 12.67 ± 1.68 µg/dL for 50 mg/L PND 65: 2.06 ± 0.35 µg/dL for 0 µg/dL 7.49 ± 0.78 µg/dL for 50 mg/L	Hct, Hb
Basha et al. (2012)	Rat (Wistar) Control (vehicle), M, n = 8 0.2% Pb, M, n = 8	PND 1 to PND 21	Damns given Pb acetate in drinking water or Pb acetate containing water supplemented with 0.02% calcium, zinc, and iron. Control group received deionized water as vehicle (no supplement). Pups exposed through lactation.	PND 45: 0.42 ± 0.04 µg/dL for 0%, 52.5 ± 0.67 µg/dL for 0.2%, 21.1 ± 1.12 µg/dL for 0.2% + supplementation PND 12 mo: 0.56 ± 0.08 µg/dL for 0%, 16.4 ± 1.95 µg/dL for 0.2%, 7.2 ± 0.56 µg/dL for 0.2% + supplementation PND 24 mo: 0.46 ± 0.02 µg/dL for 0%, 12.2 ± 0.76 µg/dL for 0.2%, 4.8 ± 0.5 µg/dL for 0.2% for 0.2% + supplementation	RBC, Hb

Study	Species (Stock/Strain), n, Sex	Timing of Exposure	Exposure Details (Concentration, Duration)	BLL as Reported (µg/dL) ^a	Endpoints Examined
Zou et al. (2015)	Mouse (ICR) Control (vehicle), M, n = 10 250 mg/L Pb, M, n = 10	3 wk exposure	Rats received 250 mg/L Pb acetate in redistilled drinking water for 3 weeks. The rats were 30 d old when acquired, but the authors did not specify the age at the time of treatment.	PND 58: 1.8 µg/dL for 0 mg/L 21.7 µg/dL for 250 mg/L	RBC, MCHC
Corsetti et al. (2017)	Mouse (C57BL.6) Control (vehicle), M, n = 8 200 ppm Pb, M, n = 8	d 30 to d 75	Mice were exposed via drinking water for 45 consecutive days. Control animals were exposed to drinking water containing acetic acid (1 mL/L).	<5 µg/dL for 0 ppm 21.6 µg/dL for 200 ppm	RBC, Hb, Hct, MCV, MCH, MCHC, RDW %, Plt
Andjelkovic et al. (2019)	Rat (Wistar) Control (vehicle), M, n = 8 0.2% Pb, M, n = 6	NR	Rats (250 g), age at time of dosing not reported, were exposed to a single dose of 150 mg Pb/kg BW Pb acetate via oral gavage. Control animals were given "water."	24.9 ± 1.9 µg/kg for 0 mg Pb/kg BW (2.6 ± 2.0 µg/dL) 291.2 ± 139 µg/kg for 150 mg Pb/kg BW (29.0 ± 14.7 µg/dL)	RBC, Hb, Hct, MCV, MCH, MCHC, Plt

Study	Species (Stock/Strain), n, Sex	Timing of Exposure	Exposure Details (Concentration, Duration)	BLL as Reported (µg/dL) ^a	Endpoints Examined
Cai et al. (2018)	Rat (Sprague Dawley) Control (vehicle), M/F, n = 5 0.2% Pb, M/F, n = 5	8–10 wk to 20–22 wk	Rats were 8– 10 wk old when acquired. Whether, or not the rats were allowed to acclimate to the facility prior to study initiation was not reported. The number of males and females not reported. Control animals received tap water. The exposure period was 12 wk, assumed rats were exposed 7 d/wk for a total of 84 d.	20.5 ± 0.68 µg/L for 0% (2.2 ± 6.4 µg/dL) 87.4 ± 9.2 µg/L for 0.2% (9.3 ± 0.98 µg/dL)	Hb, Plt, Erythrocyte life span, RBC

BLL = blood lead level; BW = body weight; d = day; Hb = hemoglobin; Hct = hematocrit; M = male; M/F = male/female; MCH = mean corpuscular hemoglobin; MCHC = mean corpuscular hemoglobin concentration; MCV = mean corpuscular volume; mo = month; NR = not reported; Pb = lead; Plt = platelet; PND = postnatal day; RBC = red blood cell; RDW = red blood cell distribution width; wk = weeks.

^aIf applicable, reported values for BLL were converted to µg/dL using WebPlot Digitizer (<https://apps.automeris.io/wpd/>) and are shown in parenthesis.

7.8 References

- Abam, E; Okediran, BS; Odukoya, OO; Adamson, I; Ademuyiwa, O. (2008). Reversal of ionoregulatory disruptions in occupational lead exposure by vitamin C. *Environ Toxicol Pharmacol* 26: 297-304. <http://dx.doi.org/10.1016/j.etap.2008.05.008>.
- Ahamed, M; Akhtar, MJ; Verma, S; Kumar, A; Siddiqui, MKJ. (2011). Environmental lead exposure as a risk for childhood aplastic anemia. 5: 38-43. <http://dx.doi.org/10.5582/bst.2011.v5.1.38>.
- Ahamed, M; Singh, S; Behari, JR; Kumar, A; Siddiqui, MKJ. (2007). Interaction of lead with some essential trace metals in the blood of anemic children from Lucknow, India. *Clin Chim Acta* 377: 92-97. <http://dx.doi.org/10.1016/j.cca.2006.08.032>.
- Ahamed, M; Verma, S; Kumar, A; Siddiqui, MKJ. (2006). Delta-aminolevulinic acid dehydratase inhibition and oxidative stress in relation to blood lead among urban adolescents. *Hum Exp Toxicol* 25: 547-553. <http://dx.doi.org/10.1191/0960327106het657oa>.
- Ahyayauch, H; García-Arribas, AB; Sot, J; González-Ramírez, EJ; Busto, JV; Monasterio, BG; Jiménez-Rojo, N; Contreras, FX; Rendón-Ramírez, A; Martín, C; Alonso, A; Goñi, FM. (2018). Pb(II) induces scramblase activation and ceramide-domain generation in red blood cells. *Sci Rep* 8: 7456. <http://dx.doi.org/10.1038/s41598-018-25905-8>.
- Alcaraz-Contreras, Y; Garza-Ocañas, L; Carcaño-Díaz, K; Ramírez-Gómez, XS. (2011). Effect of glycine on lead mobilization, lead-induced oxidative stress, and hepatic toxicity in rats. *J Toxicol* 2011: 430539. <http://dx.doi.org/10.1155/2011/430539>.
- Alghazal, MA; Šutiaková, I; Kovalkovičová, N; Legáth, J; Falis, M; Pistl, J; Sabo, R; Beňová, K; Sabová, L; Václav, P. (2008). Induction of micronuclei in rat bone marrow after chronic exposure to lead acetate trihydrate. *Toxicol Ind Health* 24: 587-593. <http://dx.doi.org/10.1177/0748233708100089>.
- Andjelkovic, M; Djordjevic, AB; Antonijevic, E; Antonijevic, B; Stanic, M; Kotur-Stevuljevic, J; Spasojevic-Kalimanovska, V; Jovanovic, M; Boricic, N; Wallace, D; Bulat, Z. (2019). Toxic effect of acute cadmium and lead exposure in rat blood, liver, and kidney. *Int J Environ Res Public Health* 16: 274. <http://dx.doi.org/10.3390/ijerph16020274>.
- Ata, SA; Abu-Dari, KI; Tutunji, MF; Mubarak, MS. (2018). Reversing the adverse biochemical effects in lead-intoxicated rats by N,N' - bis[(1,2-didehydro-1-hydroxy-2-thioxopyrid-4-yl)-carbonyl]- L-lysine. *J Trace Elem Med Biol* 50: 93-99. <http://dx.doi.org/10.1016/j.jtemb.2018.06.008>.
- Auten, RL; Davis, JM. (2009). Oxygen toxicity and reactive oxygen species: the devil is in the details [Review]. *Pediatr Res* 66: 121-127. <http://dx.doi.org/10.1203/PDR.0b013e3181a9eafb>.
- Baktybaeva, LK. (2011). [Effect of new compound BIV-30 on bone-marrow hemopoiesis under conditions of lead-acetate-induced myelosuppression]. *Eksp Klin Farmakol* 74: 26-28.
- Baranowska-Bosiacka, I; Dziedziejko, V; Safranow, K; Gutowska, I; Marchlewicz, M; Dołęgowska, B; Rać, ME; Wiszniewska, B; Chlubek, D. (2009). Inhibition of erythrocyte phosphoribosyltransferases (APRT and HPRT) by Pb²⁺: A potential mechanism of lead toxicity. *Toxicology* 259: 77-83. <http://dx.doi.org/10.1016/j.tox.2009.02.005>.
- Basha, DC; Basha, SS; Reddy, GR. (2012). Lead-induced cardiac and hematological alterations in aging Wistar male rats: alleviating effects of nutrient metal mixture. *Biogerontology* 13: 359-368. <http://dx.doi.org/10.1007/s10522-012-9380-9>.
- Bergdahl, IA; Grubb, A; Schütz, A; Desnick, RJ; Wetmur, JG; Sassa, S; Skerfving, S. (1997). Lead binding to δ -aminolevulinic acid dehydratase (ALAD) in human erythrocytes. *Basic Clin Pharmacol Toxicol* 81: 153-158. <http://dx.doi.org/10.1111/j.1600-0773.1997.tb02061.x>.

- Berrahal, AA; Lasram, M; El Elj, N; Kerkeni, A; Gharbi, N; El-Fazâa, S. (2011). Effect of age-dependent exposure to lead on hepatotoxicity and nephrotoxicity in male rats. *Environ Toxicol* 26: 68-78. <http://dx.doi.org/10.1002/tox.20530>.
- Cai, SZ; Zhou, Y; Liu, J; Li, CP; Jia, DY; Zhang, MS; Wang, YP. (2018). Alleviation of ginsenoside Rg1 on hematopoietic homeostasis defects caused by lead-acetate. *Biomed Pharmacother* 97: 1204-1211. <http://dx.doi.org/10.1016/j.biopha.2017.10.148>.
- CDC (Centers for Disease Control and Prevention). (2019). Fourth National Report on Human Exposure to Environmental Chemicals, Updated Tables, January 2019, Volume 1. Centers for Disease Control and Prevention, National Health and Nutrition Examination Survey. https://www.cdc.gov/exposurereport/pdf/FourthReport_UpdatedTables_Volume1_Jan2019-508.pdf.
- Çelik, A; Ögenler, O; Çömelekoğlu, Ü. (2005). The evaluation of micronucleus frequency by acridine orange fluorescent staining in peripheral blood of rats treated with lead acetate. *Mutagenesis* 20: 411-415. <http://dx.doi.org/10.1093/mutage/gei055>.
- Ciubar, R; Ciofrangeanu, C; Mitran, V; Cimpean, A; Stanescu, A; Iordachescu, D. (2007). The effect of lead ions on human erythrocytes in vitro. *Rev Chim* 58: 895-897.
- Coban, TA; Senturk, M; Ciftci, M; Kufrevioglu, OI. (2007). Effects of some metal ions on human erythrocyte glutathione reductase: An in vitro study. *Protein Pept Lett* 14: 1027-1030. <http://dx.doi.org/10.2174/092986607782541060>.
- Corsetti, G; Romano, C; Stacchiotti, A; Pasini, E; Dioguardi, FS. (2017). Endoplasmic reticulum stress and apoptosis triggered by sub-chronic lead exposure in mice spleen: A histopathological study. *Biol Trace Elem Res* 178: 86-97. <http://dx.doi.org/10.1007/s12011-016-0912-z>.
- Dai, Y; Huo, X; Zhang, Y; Yang, T; Li, M; Xu, X. (2017). Elevated lead levels and changes in blood morphology and erythrocyte CR1 in preschool children from an e-waste area. *Sci Total Environ* 592: 51-59. <http://dx.doi.org/10.1016/j.scitotenv.2017.03.080>.
- Daku, AB; Mustapha, S; Salisu, AI; El-Ta'alu, AB. (2019). Age-related effects of lead poisoning on some haematological parameters in adult wistar rats. *Niger J Physiol Sci* 34: 125-130.
- Eshginia, S; Marjani, A. (2013). The effect of vitamin C on the erythrocyte antioxidant enzymes in intoxicated-lead rat offsprings. *JCDR* 7: 1078-1081. <http://dx.doi.org/10.7860/JCDR/2013/5310.3059>.
- Factor-Litvak, P; Slavkovich, V; Liu, X; Popovac, D; Preteni, E; Capuni-Paracka, S; Hadzialjevic, S; Lekic, V; Lolocono, N; Kline, J; Graziano, J. (1998). Hyperproduction of erythropoietin in nonanemic lead-exposed children. *Environ Health Perspect* 106: 361-364. <http://dx.doi.org/10.1289/ehp.98106361>.
- Factor-Litvak, P; Wasserman, G; Kline, JK; Graziano, J. (1999). The Yugoslavia prospective study of environmental lead exposure. *Environ Health Perspect* 107: 9-15. <http://dx.doi.org/10.2307/3434284>.
- Farooq, Y; Farooq, MA; Hussain, A. (2016). Role of ascorbic acid supplement in amelioration of anaemia in lead intoxication. *J Pak Med Assoc* 66: 1073-1076.
- Gautam, P; Flora, SJS. (2010). Oral supplementation of gossypin during lead exposure protects alteration in heme synthesis pathway and brain oxidative stress in rats. *Nutrition* 26: 563-570. <http://dx.doi.org/10.1016/j.nut.2009.06.008>.
- Graziano, J; Slavkovich, V; Liu, XH; Factor-Litvak, P; Todd, A. (2004). A prospective study of prenatal and childhood lead exposure and erythropoietin production. *J Occup Environ Med* 46: 924-929. <http://dx.doi.org/10.1097/01.jom.0000137721.95544.4f>.
- Guo, Y; Deng, YH; Ke, HJ; Wu, JL. (2021). Iron status in relation to low-level lead exposure in a large population of children aged 0-5 years. *Biol Trace Elem Res* 199: 1253-1258. <http://dx.doi.org/10.1007/s12011-020-02253-1>.
- Hermes-Lima, M; Pereira, B; Bechara, EJH. (1991). Are free radicals involved in lead poisoning? [Review]. *Xenobiotica* 21: 1085-1090. <http://dx.doi.org/10.3109/00498259109039548>.

- Hossain, S; Bhowmick, S; Islam, S; Rozario, L; Jahan, S; Hassan, M; Sarkar, M; Choudhury, BK; Ahmed, S; Shahjalal, H. (2015). Oral administration of Ganoderma lucidum to lead-exposed rats protects erythrocytes against hemolysis: Implicates to anti-anemia. *eCAM* 2015: 463703. <http://dx.doi.org/10.1155/2015/463703>.
- Ibrahim, NM; Eweis, EA; El-Beltagi, HS; Abdel-Mobdy, YE. (2012). Effect of lead acetate toxicity on experimental male albino rat. *Asian Pacific Journal of Tropical Biomedicine* 2: 41-46. [http://dx.doi.org/10.1016/S2221-1691\(11\)60187-1](http://dx.doi.org/10.1016/S2221-1691(11)60187-1).
- Jang, WH; Lim, KM; Kim, K; Noh, JY; Kang, S; Chang, YK; Chung, JH. (2011). Low level of lead can induce phosphatidylserine exposure and erythrophagocytosis: A new mechanism underlying lead-associated anemia. *Toxicol Sci* 122: 177-184. <http://dx.doi.org/10.1093/toxsci/kfr079>.
- Karita, K; Yano, E; Dakeishi, M; Iwata, T; Murata, K. (2005). Benchmark dose of lead inducing anemia at the workplace. *Risk Anal* 25: 957-962. <http://dx.doi.org/10.1111/j.1539-6924.2005.00652.x>.
- Khan, DA; Qayyum, S; Saleem, S; Khan, FA. (2008). Lead-induced oxidative stress adversely affects health of the occupational workers. *Toxicol Ind Health* 24: 611-618. <http://dx.doi.org/10.1177/0748233708098127>.
- Kharoubi, O; Slimani, M; Krouf, D; Seddik, L; Aoues, A. (2008). Role of wormwood (*Artemisia absinthium*) extract on oxidative stress in ameliorating lead induced haematotoxicity. *African Journal of Traditional, Complementary and Alternative Medicines* 5: 263-270. <http://dx.doi.org/10.4314/ajtcam.v5i3.31282>.
- Kim, Y; Lee, BK. (2013). Increased erythrocyte lead levels correlate with decreased hemoglobin levels in the Korean general population: Analysis of 2008-2010 Korean National Health and Nutrition Examination Survey data. *Int Arch Occup Environ Health* 86: 741-748. <http://dx.doi.org/10.1007/s00420-012-0811-3>.
- Kuang, W; Chen, Z; Shi, K; Sun, H; Li, H; Huang, L; Bi, J. (2020). Adverse health effects of lead exposure on physical growth, erythrocyte parameters and school performances for school-aged children in eastern China. *Environ Int* 145: 106130. <http://dx.doi.org/10.1016/j.envint.2020.106130>.
- Kwon, SY; Chung, JH. (2016). Erythrophagocytosis of lead-exposed erythrocytes by renal tubular cells may contribute to lead-induced nephrotoxicity [Abstract]. *Toxicol Lett* 259(Suppl.): S239. <http://dx.doi.org/10.1016/j.toxlet.2016.07.574>.
- La-Llave-León, O; Lugo-Soto, R; Aguilar-Durán, M; Estrada-Martínez, S; Salas-Pacheco, JM; Sandoval-Carrillo, A; Castellanos-Juárez, FX; Barraza-Salas, M; Vázquez-Alanís, F; García-Vargas, G. (2015). Relationship between blood lead levels and hematological indices in pregnant women. *Women Health* 55: 90-102. <http://dx.doi.org/10.1080/03630242.2014.972019>.
- Lakshmi, BVS; Sudhakar, M; Aparna, M. (2013). Protective potential of Black grapes against lead induced oxidative stress in rats. *Environ Toxicol Pharmacol* 35: 361-368. <http://dx.doi.org/10.1016/j.etap.2013.01.008>.
- Lee, MK; Cho, SY; Kim, DJ; Jang, JY; Shin, KH; Park, SA; Park, EM; Lee, JS; Choi, MS; Lee, JS; Kim, MJ. (2005). Du-zhong (*Eucommia ulmoides* Oliv.) cortex water extract alters heme biosynthesis and erythrocyte antioxidant defense system in lead-administered rats. *J Med Food* 8: 86-92. <http://dx.doi.org/10.1089/jmf.2005.8.86>.
- Lee, MY; Shin, JH; Han, HS; Chung, JH. (2006). In vivo effects of lead on erythrocytes following chronic exposure through drinking water. *Arch Pharm Res* 29: 1158-1163. <http://dx.doi.org/10.1007/BF02969308>.
- Li, C; Ni, ZM; Ye, LX; Chen, JW; Wang, Q; Zhou, YK. (2018). Dose-response relationship between blood lead levels and hematological parameters in children from central China. *Environ Res* 164: 501-506. <http://dx.doi.org/10.1016/j.envres.2018.03.018>.
- Liu, C; Huo, X; Lin, P; Zhang, Y; Li, W; Xu, X. (2015). Association between blood erythrocyte lead concentrations and hemoglobin levels in preschool children. *Environ Sci Pollut Res Int* 22: 9233-9240. <http://dx.doi.org/10.1007/s11356-014-3992-3>.
- Liu, J; McCauley, L; Yan, C; Shen, X; Pinto-Martin, JA. (2012). Low blood lead levels and hemoglobin concentrations in preschool children in China. *Toxicol Environ Chem* 94: 423-426. <http://dx.doi.org/10.1080/02772248.2011.628001>.

- Mani, MS; Joshi, MB; Shetty, RR; Dsouza, VL; Swathi, M; Kabekkodu, SP; Dsouza, HS. (2020). Lead exposure induces metabolic reprogramming in rat models. *Toxicol Lett* 335: 11-27. <http://dx.doi.org/10.1016/j.toxlet.2020.09.010>.
- Marques, CC; Nunes, AC; Pinheiro, T; Lopes, PA; Santos, MC; Viegas-Crespo, AM; Ramalhinho, MG; Mathias, ML. (2006). An assessment of time-dependent effects of lead exposure in algerian mice (*Mus spretus*) using different methodological approaches. *Biol Trace Elem Res* 109: 75-89. <http://dx.doi.org/10.1385/BTER:109:1:075>.
- Massó-González, EL; Antonio-García, MT. (2009). Natural antioxidants protect against lead-induced damage during pregnancy and lactation in rat's pups. *Ecotoxicol Environ Saf* 72: 2137-2142. <http://dx.doi.org/10.1016/j.ecoenv.2009.03.013>.
- Massó, EL; Corredor, L; Antonio, MT. (2007). Oxidative damage in liver after perinatal intoxication with lead and/or cadmium. *J Trace Elem Med Biol* 21: 210-216. <http://dx.doi.org/10.1016/j.jtemb.2007.03.002>.
- Molina, RM; Phattanasudee, S; Kim, J; Thompson, K; Wessling-Resnick, M; Maher, TJ; Brain, JD. (2011). Ingestion of Mn and Pb by rats during and after pregnancy alters iron metabolism and behavior in offspring. *Neurotoxicology* 32: 413-422. <http://dx.doi.org/10.1016/j.neuro.2011.03.010>.
- Monteiro, HP; Abdalla, DSP; Augusto, O; Bechara, EJH. (1989). Free radical generation during δ -Aminolevulinic acid autoxidation: Induction by hemoglobin and connections with porphyriopathies. *Arch Biochem Biophys* 271: 206-216. [http://dx.doi.org/10.1016/0003-9861\(89\)90271-3](http://dx.doi.org/10.1016/0003-9861(89)90271-3).
- Monteiro, HP; Abdalla, DSP; Faljoni-Alário, A; Bechara, EJH. (1986). Generation of active oxygen species during coupled autoxidation of oxyhemoglobin and δ -aminolevulinic acid. *Biochim Biophys Acta* 881: 100-106. [http://dx.doi.org/10.1016/0304-4165\(86\)90102-9](http://dx.doi.org/10.1016/0304-4165(86)90102-9).
- Monteiro, HP; Bechara, EJH; Abdalla, DSP. (1991). Free radicals involvement in neurological porphyrias and lead poisoning [Review]. *Mol Cell Biochem* 103: 73-83. <http://dx.doi.org/10.1007/BF00229595>.
- Mrugesh, T; Dipa, L; Manishika, G. (2011). Effect of lead on human erythrocytes: An in vitro study. *Acta Pol Pharm* 68: 653-656.
- Nagano, S; Takahashi, Y; Yamamoto, K; Masutani, H; Fujiwara, N; Liu, J; Jia, D; Cai, S; Li, CP; Zhang, MS; Zhang, Y; Yan, CH; Wang, YP. (2015). Mitochondria defects are involved in lead-acetate-induced adult hematopoietic stem cell decline. *Toxicol Lett* 235: 37-44. <http://dx.doi.org/10.1016/j.toxlet.2015.03.007>.
- Odo, RI; Mbegbu, EC; Anyanwu, LN. (2020). Effect of aqueous ginger (*Zingiber officinale*) extract on sperm quality and haematology in lead acetate-treated male albino rats. *Tropical Journal of Pharmaceutical Research* 19: 1481-1485. <http://dx.doi.org/10.4314/tjpr.v19i7.21>.
- Olivero-Verbel, J; Duarte, D; Echenique, M; Guette, J; Johnson-Restrepo, B; Parsons, PJ. (2007). Blood lead levels in children aged 5-9 years living in Cartagena, Colombia. *Sci Total Environ* 372: 707-716. <http://dx.doi.org/10.1016/j.scitotenv.2006.10.025>.
- Park, S; Lee, BK. (2013). Body fat percentage and hemoglobin levels are related to blood lead, cadmium, and mercury concentrations in a Korean adult population (KNHANES 2008-2010). *Biol Trace Elem Res* 151: 315-323. <http://dx.doi.org/10.1007/s12011-012-9566-7>.
- Patil, AJ; Bhagwat, VR; Patil, JA; Dongre, NN; Ambekar, JG; Jailkhani, R; Das, KK. (2006). Effect of lead (Pb) exposure on the activity of superoxide dismutase and catalase in battery manufacturing workers (BMW) of Western Maharashtra (India) with reference to heme biosynthesis. *Int J Environ Res Public Health* 3: 329-337. <http://dx.doi.org/10.3390/ijerph2006030041>.
- Queirolo, EI; Ettinger, AS; Stoltzfus, RJ; Kordas, K. (2010). Association of anemia, child and family characteristics with elevated blood lead concentrations in preschool children from Montevideo, Uruguay. *Arch Environ Occup Health* 65: 94-100. <http://dx.doi.org/10.1080/19338240903390313>.
- Quintanar-Escorza, MA; González-Martínez, MT; Navarro, L; Maldonado, M; Arévalo, B; Calderón-Salinas, JV. (2007). Intracellular free calcium concentration and calcium transport in human erythrocytes of lead-exposed workers. *Toxicol Appl Pharmacol* 220: 1-8. <http://dx.doi.org/10.1016/j.taap.2006.10.016>.

- Rendón-Ramírez, A; Cerbón-Solórzano, J; Maldonado-Vega, M; Quintanar-Escorza, MA; Calderón-Salinas, JV. (2007). Vitamin-E reduces the oxidative damage on δ -aminolevulinic dehydratase induced by lead intoxication in rat erythrocytes. *Toxicol In Vitro* 21: 1121-1126.
<http://dx.doi.org/10.1016/j.tiv.2007.04.019>.
- Riddell, TJ; Solon, O; Quimbo, SA; Tan, CMC; Butrick, E; Peabody, JW. (2007). Elevated blood-lead levels among children living in the rural Philippines. *Bull World Health Organ* 85: 674-680.
<http://dx.doi.org/10.2471/blt.06.036137>.
- Sajitha, GR; Augusti, KT; Jose, R. (2016). Prophylactic effects of garlic oil and onion oil fractions as compared to vitamin E on rats orally fed with lead acetate solution. *Indian J Clin Biochem* 31: 260-269.
<http://dx.doi.org/10.1007/s12291-015-0526-9>.
- Sarkar, A; Sengupta, D; Mandal, S; Sen, G; Chowdhury, KD; Sadhukhan, GC. (2015). Treatment with garlic restores membrane thiol content and ameliorates lead induced early death of erythrocytes in mice. *Environ Toxicol* 30: 396-410. <http://dx.doi.org/10.1002/tox.21901>.
- Shah, F; Kazi, TG; Afridi, HI; Baig, JA; Khan, S; Kolachi, NF; Wadhwa, SK; Shah, AQ. (2010). Environmental exposure of lead and iron deficit anemia in children age ranged 1-5 years: A cross sectional study. *Sci Total Environ* 408: 5325-5330. <http://dx.doi.org/10.1016/j.scitotenv.2010.07.091>.
- Sharma, V; Sharma, A; Kansal, L. (2010). The effect of oral administration of *Allium sativum* extracts on lead nitrate induced toxicity in male mice. *Food Chem Toxicol* 48: 928-936.
<http://dx.doi.org/10.1016/j.fct.2010.01.002>.
- Shin, JH; Lim, KM; Noh, JY; Bae, ON; Chung, SM; Lee, MY; Chung, JH. (2007). Lead-induced procoagulant activation of erythrocytes through phosphatidylserine exposure may lead to thrombotic diseases. *Chem Res Toxicol* 20: 38-43. <http://dx.doi.org/10.1021/tx060114+>.
- Simons, TJ. (1986). Passive transport and binding of lead by human red blood cells. *J Physiol* 378: 267-286.
<http://dx.doi.org/10.1113/jphysiol.1986.sp016219>.
- Simons, TJB. (1993). Lead transport and binding by human erythrocytes in vitro. *Pflugers Arch* 423: 307-313.
<http://dx.doi.org/10.1007/BF00374410>.
- Simsek, N; Karadeniz, A; Kalkan, Y; Keles, ON; Unal, B. (2009). *Spirulina platensis* feeding inhibited the anemia- and leucopenia-induced lead and cadmium in rats. *J Hazard Mater* 164: 1304-1309.
<http://dx.doi.org/10.1016/j.jhazmat.2008.09.041>.
- Terayama, K; Maehara, N; Muratsugu, M; Makino, M; Yamamura, K. (1986). Effect of lead on electrophoretic mobility of rat erythrocytes. *Toxicology* 40: 259-265.
- U.S. EPA (U.S. Environmental Protection Agency). (2006a). Air quality criteria for lead [EPA Report]. (EPA/600/R-05/144aF-bF). Research Triangle Park, NC.
<https://cfpub.epa.gov/ncea/risk/recordisplay.cfm?deid=158823>.
- U.S. EPA (U.S. Environmental Protection Agency). (2006b). Air quality criteria for lead (Final report, 2006): Volume I of II [EPA Report]. (EPA/600/R-05/144aF). Washington, DC.
<http://cfpub.epa.gov/ncea/CFM/recordisplay.cfm?deid=158823>.
- U.S. EPA (U.S. Environmental Protection Agency). (2013). Integrated science assessment for lead [EPA Report]. (EPA/600/R-10/075F). Washington, DC. <https://nepis.epa.gov/Exe/ZyPURL.cgi?Dockkey=P100K82L.txt>.
- U.S. EPA (U.S. Environmental Protection Agency). (2015). Preamble to the Integrated Science Assessments [EPA Report]. (EPA/600/R-15/067). Research Triangle Park, NC: U.S. Environmental Protection Agency, Office of Research and Development, National Center for Environmental Assessment, RTP Division.
<https://cfpub.epa.gov/ncea/isa/recordisplay.cfm?deid=310244>.
- Ukaejiofo, EO; Thomas, N; Ike, SO. (2009). Haematological assessment of occupational exposure to lead handlers in Enugu urban, Enugu State, Nigeria. *Niger J Clin Pract* 12: 58-64.

- Velaga, MK; Daughtry, LK; Jones, AC; Yallapragada, PR; Rajanna, S; Rajanna, B. (2014). Attenuation of lead-induced oxidative stress in rat brain, liver, kidney and blood of male Wistar rats by Moringa oleifera seed powder. *J Environ Pathol Toxicol Oncol* 33: 323-337. <http://dx.doi.org/10.1615/JEnvironPatholToxicolOncol.2014011656>.
- Wang, Q; Zhao, HH; Chen, JW; Hao, QL; Gu, KD; Zhu, YX; Zhou, YK; Ye, LX. (2010a). δ -Aminolevulinic acid dehydratase activity, urinary δ -aminolevulinic acid concentration and zinc protoporphyrin level among people with low level of lead exposure. *Int J Hyg Environ Health* 213: 52-58. <http://dx.doi.org/10.1016/j.ijheh.2009.08.003>.
- Wang, TL; Kao, TH; Inbaraj, BS; Su, YT; Chen, BH. (2010b). Inhibition effect of poly(γ -glutamic acid) on lead-induced toxicity in mice. *J Agric Food Chem* 58: 12562-12567. <http://dx.doi.org/10.1021/jf1034509>.
- Wang, X; Wang, L; Liu, S. (2015). Heme-regulated eIF2 α kinase plays a crucial role in protecting erythroid cells against Pb-induced hemolytic stress. *Chem Res Toxicol* 28: 460-469. <http://dx.doi.org/10.1021/tx500422q>.
- Whittaker, MH; Wang, G; Chen, XQ; Lipsky, M; Smith, D; Gwiazda, R; Fowler, BA. (2011). Exposure to Pb, Cd, and As mixtures potentiates the production of oxidative stress precursors: 30-day, 90-day, and 180-day drinking water studies in rats. *Toxicol Appl Pharmacol* 254: 154-166. <http://dx.doi.org/10.1016/j.taap.2010.10.025>.
- Zou, Y; Feng, W; Wang, W; Chen, Y; Zhou, Z; Li, Q; Zhao, T; Mao, G; Wu, X; Yang, L. (2015). Protective effect of porcine cerebral hydrolysate peptides on learning and memory deficits and oxidative stress in lead-exposed mice. *Biol Trace Elem Res* 168: 429-440. <http://dx.doi.org/10.1007/s12011-015-0329-0>.